


ram

MSc Report Joon and Riya[1].docx

 Delhi Technological University

Document Details

Submission ID

trn:oid:::27535:124510582

Submission Date

Dec 15, 2025, 1:47 PM GMT+5:30

Download Date

Dec 15, 2025, 1:52 PM GMT+5:30

File Name

MSc Report Joon and Riya[1].docx

File Size

69.1 KB

27 Pages

8,329 Words

53,306 Characters





4% Overall Similarity

The combined total of all matches, including overlapping sources, for each database.




Filtered from the Report

- Bibliography
- Quoted Text
- Cited Text
- Small Matches (less than 10 words)

Match Groups

-  **11 Not Cited or Quoted 4%**
Matches with neither in-text citation nor quotation marks
-  **0 Missing Quotations 0%**
Matches that are still very similar to source material
-  **0 Missing Citation 0%**
Matches that have quotation marks, but no in-text citation
-  **0 Cited and Quoted 0%**
Matches with in-text citation present, but no quotation marks

Top Sources

- 3%  Internet sources
- 3%  Publications
- 0%  Submitted works (Student Papers)

Integrity Flags

0 Integrity Flags for Review

No suspicious text manipulations found.

Our system's algorithms look deeply at a document for any inconsistencies that would set it apart from a normal submission. If we notice something strange, we flag it for you to review.

A Flag is not necessarily an indicator of a problem. However, we'd recommend you focus your attention there for further review.

Match Groups

- 11 Not Cited or Quoted 4%**
Matches with neither in-text citation nor quotation marks
- 0 Missing Quotations 0%**
Matches that are still very similar to source material
- 0 Missing Citation 0%**
Matches that have quotation marks, but no in-text citation
- 0 Cited and Quoted 0%**
Matches with in-text citation present, but no quotation marks

Top Sources

- 3% Internet sources
- 3% Publications
- 0% Submitted works (Student Papers)

Top Sources

The sources with the highest number of matches within the submission. Overlapping sources will not be displayed.

- 1

Internet

ijfas.com

<1%
- 2

Internet

link.springer.com

<1%
- 3

Publication

Kiran Suresh Mawale, Parvatam Giridhar. "Green approaches for the synthesis of ...

<1%
- 4

Internet

www.ncbi.nlm.nih.gov

<1%
- 5

Internet

www.iiste.org

<1%
- 6

Publication

"Nanobiotechnology for Abiotic Stress Adaptation and Mitigation in Agricultural ...

<1%
- 7

Publication

Elsayed A. Elkhatab, Hala M. Hamadeen. "Role of nanofertilizers in improving abio...

<1%

Application of Silver based nanoparticle for seed priming of medicinal and herbal plants

Abstract

Medicinal plants have been essential to the global evolution of human cultures. Many abiotic and biotic processes affect the growth and development of plants. Seedling establishment and germination are important phases in the plant life cycle and efficient seed germination encourages successful establishment and deep root system of plants. Through the production of various physiological and metabolic changes, priming has been shown to have a noteworthy impact. This review summarises the studies of various medicinal and herbal plants by using nano-priming of seeds.

A literature search was conducted with the help of electronic databases like Google Scholar, PubMed, Scopus, Sci Finder, and ONOS. The search was conducted using the keywords Medicinal Plants, Seed priming, nanotechnology, silver nanoparticle, germination.

The studies have shown that the use of silver nanoparticles (AgNPs) as nanopriming agent for enhancing seed germination is a step towards sustainable agriculture. This process is suitable for small seed plants.

This study gathered information on the seed germination using silver nanoparticles on various medicinal plants.

Introduction

Medicinal and herbal plants represent the most ancient form of medication, used for thousands of years in traditional medicine in many countries around the world (Marrelli, 2021). The empirical knowledge about their beneficial effects was transmitted over the centuries within human communities (Khan, 2014). Medicinal plants not only serve as complements or substitute for modern medical treatments, which are often inadequately available but also enhance the health and security of local people (Agidew, 2022). The medicinal importance of some plants is mainly due to the presence of active ingredients such as terpenes, flavonoids, coumarins, carotenoids, essential oils, and amino acids (Khalaki et al., 2021; Ahmad et al., 2018).

The cultivation of medicinal and herbal plants is currently confronted with several difficulties, such as changeable climatic circumstances that can negatively impact seedling development, seed germination, and crop output, such as drought, salt, heavy metal buildup in soil, and climate change (Shelar et al., 2023; Yadav et al., 2020; Imran et al., 2021; He et al., 2018). The uncertainty of the global climate with erratic rainfall patterns is the major cause of the frequent onset of drought stress around the world (Lobell et al., 2011). Over 6% of the world's land is spoiled by salinity which negatively affects crop survival by minimizing the growth and yield of staple food up to 70% (FAO, 2016; Schilling, 2016).

Drought-induced economic losses were estimated to be about 29 billion dollars during 2005 to 2015, and it is predicted to become more persistent and extensive in the coming decades (Schilling, 2016; Trenberth et al., 2014). By 2050, about 50% of arable lands are expected to be under drought stress (Marthandan et al., 2020). Drought can occur in all growth stages, but the first and foremost effect is on seed germination (Kasim et al., 2013; Kaya et al., 2006) where water entrance into the seed decreases due to hydraulic reduction; and thereby, all the physiological and metabolic germination processes are affected (Fahad et al., 2017). Impaired germination and establishment under drought stress have been studied in several crops viz., peas (Bareke, 2018), barnyard millet (Okcu et al., 2005), rice (Wu et al., 2019), and sunflower (Islam et al., 2018).

Efficient seed germination is important for increasing the production of forage and medicinal plants in rangeland fields (Humera Razzaq et al., 2017). The main role of the seed is to protect the embryo and sense environmental information to couple germination with seasons compatible with the completion of the plant life cycle (Azimi et al., 2014). The germination process is completed in three stages. In the stage 1, seed germinates upon absorption of water in the cells of the seed as they hydrate, and become rigid or turgid. In

stage 2, the areas of growth, cell division, and differentiation at the root and shoot meristems or tips are where the sugars and amino acids are focused. In stage 3, the seed coat bursts due to the swelling in cells. The primary root or radicle emerges downward, and the stem grows upward (Carrera-Castaño et al., 2020).

There is an urgent need to develop a sustainable technology that can contribute to the green revolution to address these growing concerns and to restore the damage caused to the ecosystem (Mahra et al., 2024). Seed priming is an innovative technique to improve seed germination rates, seedling growth, and crop yield, as well as provide resistance to various plant stresses like drought, salinity, and heavy metal toxicity in a sustainable way (Shelar et al., 2021). Priming is the process of pre-treating seeds before planting those plants using traditional methods such as pre-soaking and coating (Maroufi et al., 2011). Priming creates a physiological state in the seed that strengthens its growth capacity against biotic or abiotic stresses (Nile et al., 2022). However, many factors affect the performance of seed priming such as plant species, priming duration, temperature, priming media, and their concentration and storage conditions (Conrath, 2011; Rhaman et al., 2020).

Priming using nanoparticles (nano-priming) has been proven to be more promising than traditional priming approaches for achieving feasible agricultural yields (Rhaman et al., 2020). Nano-priming uses particles with a size of less than 100 nm (nanoparticles, NPs), and "priming" relates to the development of stress tolerance under moderate and recurring stress (Nile et al., 2022; Chandrasekaran et al., 2020). The literature search has shown that nanomaterials prepared with silver (Rajwade et al., 2020), gold (Prasad et al., 2017), copper (Usman et al., 2020), palladium (Kamle et al., 2020), selenium (Liu et al., 2021), zinc oxide (Pramanik et al., 2020), magnesium oxide (Jiang et al., 2020), titanium dioxide (Shang et al., 2019), and iron oxide (Pulizzi, 2019) have been proven to promote seed germination and improve crop yields (Figure 1).

Silver nanoparticles (AgNPs) are the most widely used nanoparticles with a variety of uses because of their unique properties (Yan and Chen 2019). The applications of AgNPs have received high focus and promotion in the medical and pharmaceutical fields (Khan et al., 2023). Among numerous monometallic NPs, biogenic AgNPs are frequently applied because of their characteristics such as electrical conductivity, optical polarization, and SER scattering (Abasi et al., 2022). The potential uses of Ag-NPs in catalysis (Kamat 2002), plasmonics (Maier et al. 2001), optoelectronics (Boncheva et al. 2002), biological sensors (Mirkin et al. 1996; Han et al. 2001), antimicrobial activities (Savithramma et al. 2011; Rai et al. 2009), DNA sequencing (Cao et al. 2001), climate change and contamination control

(Shan et al. 2009), clean water technology (Savage and Diallo 2005), energy generation (Zech et al. 2006), information storage (Caruthers et al. 2007), and biomedical applications (Hullmann 2007) have all attracted a lot of interest in the formation of Ag-NPs. By showcasing its potential during the past ten years, the production of NPs has given us amazing advancements in the field of nanotechnology (Samberg et al. 2010).

Therefore, the current study's objectives are to investigate the potential utility of varying AgNPs priming concentrations on the morpho-physiological and biochemical characteristics of herbal and medicinal plants under salt stress, including growth, biomass, oxidative damage, and antioxidant system.

Basics of Seed Priming

Priming is the process of pre-treating seeds before planting those plants using traditional methods such as pre-soaking and coating (Torre-Roche et al., 2020; Waqas et al., 2019). Seed priming is controlled hydration of seeds to a level that allows pre-germinative metabolic activity to continue, but interrupts actual emergence of the radicle (Raj and Raj, 2019). Although seedlings raised from primed seeds have been reported to exhibit modifications in water contents, improved cell cycle regulation, management of oxidative stress and reserve food mobilization, the efficacy of seed priming highly depends on the plant species and the priming method (Raj and Raj, 2019; Johnson and Puthur, 2021). Several seed and agricultural firms have used the priming technology as a revolutionary strategy for achieving a consistent crop standby (Sivasubramaniam et al., 2011; Waqas et al., 2019; Raj and Raj, 2019).

Priming initiates cross-tolerance that assists improved germination and seedling establishment under harsh environmental conditions (Chen et al., 2012). This has been demonstrated in many crop plants, such as zucchini (*Cucurbita pepo* L.), onion (*Allium cepa* L.), pepper (*Capsicum annum* L.), tomato (*Lycopersicon esculentum* L.), zinnia (*Zinnia elegans* Jacq.) etc. (Anand et al., 2019; Szopinska and Polityvka, 2016; Silva et al., 2015; Zhao et al., 2018; Valivand et al., 2019). In mung bean plants, a faster seedling establishment resulting from priming may contribute to a total increase in yield up to 45% (Rashid et al., 2004). Increased seed vigour observed in primed seeds of Arabidopsis (Gallardo et al., 2001; Rajjou et al., 2006), alfalfa (*Medicago sativa* L.) (Yacoubi et al., 2013), wheat (Fercha et al., 2013; Fercha et al., 2014).

Proteome analyses of seed priming and germination have proven invaluable in identifying changes between primed and unprimed seeds in various plants (Wang et al.,

2015). It is regulated by the interaction of the surrounding environmental conditions, the seeds' physiological state, and the germ (Khaeim et al., 2022; Vishal and Kumar, 2018). Temperature, light, pH, water availability, and soil moisture most affect seed germination among abiotic factors (Khaeim et al., 2022; Rizzardi et al., 2009). Figure 2 shows that **plants produced from primed seeds often exhibit a faster growth than unprimed ones**. The beneficial impact of priming on plant growth may be due to an improved nutrient use efficiency allowing a higher relative growth rate (Debbarma and Das, 2017; Muhammad et al., 2015). A higher growth of seedlings issued from primed seeds may also be analyzed in relation to a direct impact of pretreatment on cell cycle regulation and cell elongation processes (Debarma and Das, 2017; Chen and Arora, 2013). The growth parameters of chickpea were significantly affected by seed priming (Debbarma and Das, 2017; Vikas and Mahender, 2012).

Seed germination with Silver Nanoparticles

Nanotechnology utilizes particles less than 100 nm in size, and it has a promising role in transforming agriculture (Fraceto et al., 2016). The two techniques for NP synthesis are "Top-down" and "Bottom-up." The most flexible method for producing metal nanoparticles top-down is evaporation-condensation (Rafique et al., 2017; Mathur et al., 2017). Nanoparticles exhibit high surface-area-to-volume ratios, enhanced reactivity, and controlled release capabilities, making them valuable for precision agriculture (Kędziora et al., 2018).

The excessive use of chemical fertilizers can be reduced by utilizing nanomaterials in agriculture (Upadhyaya et al., 2017). The mechanism behind high seed germination in nano-priming is the greater penetration *via* seed coat that improves nutrient and water uptake efficiency of the seed (Dutta, 2018). Studies reported that seed priming with calcium-phosphate, SiO₂, ZnO, and Ag nanoparticles enhanced germination and seedling development (Ghafari and Razmjoo, 2013).

One of the most promising methods for increasing agricultural output in both favourable and unfavourable environmental circumstances is the application of nanoparticles (NPs) (Biswas et al., 2023; Zulfiqar and Ashraf, 2021). It has been documented that nanoprimer controls physio-biochemical reactions in abiotically stressed crops (Figure 3). AgNPs (Thiruvengadam et al., 2015; Baskar et al., 2015), AuNPs (Alshehddi and Bokhari, 2020), CuNPs (Chung et al., 2019; Nguyen et al., 2022), FeNPs (Kornarzyński et al., 2020), FeS₂NPs (Srivastava et al., 2014), TiO₂NPs (Gohari et al., 2020; Faraji and Sepehri, 2019), ZnNPs (Montanha et al., 2020), ZnONPs (Faizan et al., 2021; Taran et al., 2017; Abdel Latef et al., 2017), and carbon NPs like fullerene (Shafiq et al., 2019) and carbon nanotubes

(Rahimi et al., 2016) have all been used as seed pretreatment. In maize plants, seed priming using gold nanoparticles (AuNPs) frequently improves water absorption (Rajwade et al., 2020).

It has been reported that commercially available silver nanoparticles (AgNPs) with sizes ranging from 50 to 100 nm can lessen the negative effects of stress caused by 120 and 150 mM NaCl in *Pennisetum glaucum* (Parveen and Rao 2014). Additional instances of plants that show improved development and a favorable physiological response after being exposed to silver nanoparticles include *Phaseolus vulgaris*, *Zea mays*, (Salama 2012) *Phytolacca Americana*, *Panicum virgatum* (Yin et al., 2012). In *Boswellia ovalifoliolata* (Savithramma et al., 2012), treatment with AgNPs has been demonstrated to have a favorable effect on seed germination. In *Brassica nigra* seeds and seedlings, it was similarly discovered that an AgNP dose ranging from 0.2 to 1.6 mg/L inhibited lipase activity, soluble and reducing sugar content, and seed germination (Amooaghaiea and Tabatabaiea 2015). Additionally, it has been discovered that 10 mg/L AgNPs hinder *Hordeum vulgare* seed germination and shorten the shoot length of barley (*Hordeum vulgare*) and flax (*Linum usitatissimum*) (Temsah and Joner 2012).

To increase plant tolerance to salinity, AgNPs enhanced plant height, proline contents, ion homeostasis, antioxidant enzyme activities, and phenolic and flavonoid contents (Biswas et al., 2023; Khan et al., 2020). While higher and lower concentrations of AgNPs may have a negative impact on plant growth, exposure to particular concentrations may improve plant growth when compared to non-exposed plants (Kaveh et al., 2013; Geisler-Lee et al., 2013; Qian et al., 2013). The optimum concentration of AgNPs to induce a growth response in *Brassica juncea* seedlings was found to be 50 ppm, out of the utilized concentrations of 0, 25, 50, 100, 200, and 400 ppm (Almutairi and Alharbi, 2015). The root length, fresh weight, shoot length, and vigour index of seedlings had positive impact at this concentration. The treated seedlings' vigour index increased by 133% and their root length increased by 326% as a result of this dosage (Sharma et al., 2012). Seedling biomass was shown to increase when *Arabidopsis thaliana* plants were treated with 1 or 2.5 mg/L of AgNPs, while seedling biomass decreased when larger doses were applied (Kaveh et al., 2013).

In order to investigate drought stress on lentil germination five levels of silver nanoparticles (0, 10, 20, 30 and 40 $\mu\text{g mL}^{-1}$) on Lentil (*Lens culinaris Medic*) were taken AgNP enhanced the radical length and the highest length (4.45 cm) observed in severe stress by applying 10 $\mu\text{g mL}^{-1}$ of the nanoparticle. When plants were treated with 20 $\mu\text{g/mL}$ of

silver nanoparticles (AgNPs), the radicle (embryonic root) had the highest dry weight, which was 0.07 grams (Hojjat and Ganjali 2016).

Studies also showed that the addition of 10 or 50 mg·dm⁻³ of AgNPs changes the composition of essential oils produced by lavender plant (Jadczak et al., 2020). For instance, broad bean (*Vicia faba L.*) seeds exposed to citrate-coated silver nanoparticles (ca. 40 nm) at 10 ppm for 6 h showed higher growth, physiological, and biological traits than non-nano treatments (Alhammad et al., 2023). With alfalfa (*Medicago sativa L.*), the results were also encouraging when exposed to AgNPs (approximately 50 nm) for 3 h (Song et al., 2022). AgNPs exhibited strong antimicrobial activity against a variety of diseases. For example, they significantly limit the growth of fungi that cause rice blast disease and the colony formation of *Magnaporthe grisea* (Vishwanathan and Negi 2021). (Matras 2022) reported that AgNPs had antifungal efficacy against *Alternaria alternata* and *Botrytis cinerea*. According to the study, the highest inhibition of fungal hyphal development was caused by the 15 mg/L concentration of AgNPs. However, it was demonstrated that AgNPs had no discernible effects on the castor bean plant, *Ricinus communis L.*, in terms of seed germination, root length, or shoot length (Yasur and Rani 2013). *Cucumis sativus* and *Lactuca sativa* seed germination was observed to be unaffected by 100 mg/L AgNPs (Ingle et al., 2020). *M. incognita* invasive larvae viability was 100% reduced and the egg hatch process was 100% inhibited after incubation with a very low quantity of silver compounds (0.05 ppm) (Furmanczyk et al., 2025).

Satureja hortensis L., is a medicinal plant endemic to Iran, which is well-known in the folk medicine for its therapeutic uses as herbal tea and as an analgesic and antiseptic substance due to the presence of secondary metabolites including terpenoids, phenolics, flavonoids, steroids, and tannins (Hosainzadegan and Delfan, 2009). During recent years, antibacterial, antioxidant, antifungal, antidiabetic, antinociceptive, antihyperlipidemic, antibiofilm, anti-inflammatory, antispasmodic and antidiarrhea effects and as well as triglyceride-lowering potential have been reported for *S. hortensis* (Hosainzadegan and Delfan, 2009). Priming of *S. hortensis L.* (summer savory) seeds before planting had a positive physiological and biochemical impact. This likely resulted in improved seed germination, enhanced stress tolerance, and better overall plant growth by activating beneficial metabolic processes before the seeds were sown in the seedbed. In *S. hortensis*, the highest seed germination percentage was observed when 80 ppm of AgNPs were applied, particularly under conditions of low salinity. Conversely, the lowest germination percentage

was recorded under high salinity conditions 120 MmL^{-1} when AgNPs were not applied (Nejatzadeh, 2021).

According to some studies, a high concentration of AgNPs had detrimental effects on photosynthesis, lowered the amount of chlorophyll overall, and dramatically raised the parameters of oxidative stress (Song et al., 2022; Abasi et al., 2022). The aquatic plant *Lemna gibba* has been shown to exhibit growth inhibition, as evidenced by a notable decline in frond numbers that is dependent on AgNPs concentration (Oukarroum et al., 2013). According to (Stampoulis et al., 2009), the biomass and transpiration rates of *Cucurbita pepo* (zucchini) were reduced by 41% and 57%, respectively, when exposed to 100 nm AgNPs at 100 and 500 mg/L. AgNPs dramatically reduced plant biomass, plant tissue nitrate-nitrogen content, chlorophyll a/b, and chlorophyll fluorescence (Fv/Fm) in an aquatic macrophyte (*Spirodela polyrhiza*, larger duckweed), according to a study by Jiang et al. (2012). Plant toxicity was solely examined in pure cultures in any of these investigations.

AgNPs seed priming has become a viable method for improving crop development and physiological responses to abiotic stressors, such as salinity. In order to help seeds overcome environmental obstacles and improve germination rates and seedling establishment, AgNPs enhance water uptake, enzyme activity, and oxidative stress tolerance. Optimizing the concentration and application techniques is essential to avoiding possible toxicity, though. However, concerns regarding nanoparticle toxicity, environmental persistence, and regulatory challenges necessitate further research to optimize their safe and sustainable application in modern agriculture.

A simple way to improve seed germination and seedling establishment and consequently field performance of medicinal plants is seed priming (Dalil, 2014). Most of the medicinal plants have some problems in seed germination and stand establishment in the field (Zare et al., 2011). Since germination and seedling establishment are critical stages in the plant life cycle (Cheng and Bradford, 1999), offering the solutions for improvement of seed germination and seedling establishment will help to better performance in cultivation of medicinal plants. One of the simple techniques which can improve seedling vigor and establishment and consequently field performance of plants is seed priming or physiological advancement of the seed (Mc Donald, 2000). In comparison to the control seedlings, AgNPs increased the amount of chlorophyll and enhanced the cellular electron exchange and photosynthetic quantum efficiencies in medicinal plant (Sharma et al., 2012).

According to several reports, flavonoids are in charge of the environmentally friendly synthesis of AgNPs (Mustapha et al. 2022). AgNPs produced synthetically have been utilized

a food packaging, wastewater treatment, and biological uses as an antibacterial agent (Vanlalveni et al. 2021). This method is successful in small seed plants such as many medicinal plants that have great economic value with quick and uniform emergence requirements (Ellis and Roberts, 1981). Majority of studies focus on agricultural plants and some focus on flowering/medicinal plants have been compiled in Table 1 (*Psophocarpus tetragonolobus* (L.), *Vigna mungo* (L.) Hepper, *Satureja hortensis* L., *Sanguisorba minor*, *Lavandula angustifolia*, *Allium cepa* L, *Phaseolus vulgaris* L., *Echinops macrochaetus*, *Medicago sativa* L., *Linum usitatissimum* L., *Ocimum basilicum* L., *Helianthus annuus* L., *Solanum lycopersicum*, *Vicia faba*, *Capsicum annuum* L.).

Table 1: Various plants studied under silver nanoparticles seed priming:

Plant studied	Seed condition	Concentration of NPs	Aspect studied	Impact of NPs	References
<i>Psophocarpus tetragonolobus</i> (L.) (Legume)	Seedling growth stress	50 mg	Seed germination	Increased upto 88.33%	(Kumar et al., 2020)
<i>Vigna mungo</i> (L.) Hepper	Poor seed vigour, Low germination rates	1.5 mM	Germination percentage Root length Shoot length Vigour index	Increased upto 21% Increased upto 19% Increased upto 23% Increased upto 35%	(Krishnasamy et al., 2024)
<i>Satureja hortensis</i> L.	salinity stress	80 ppm	Seed germination Salinity stress	Improved salinity tolerance with increased seed germination	(Nejatzadeh, 2021)

<i>Sanguisorba minor</i>	Drought stress	30 mg/l	Biomass production	Increased by 25%	(Farmahini Farahani et al., 2022)
<i>Lavandula angustifolia</i>	Lack of enhanced metabolic and defensive activities	2 mg·dm ⁻³	activity of antioxidant enzymes and total polyphenolic capacity	Increased activity of APX and SOD	(Jadczak et al., 2020)
<i>Allium cepa L.</i>		25 ppm	Seed germination and vigour index	Stimulated early germination and significant increase in Vigour Index	(Patidar et al., 2024)
<i>Phaseolus vulgaris L.</i>		25 mg/L 50 mg/L	Root length Flavonoids Protein Chlorophyll content Shoot length	Maximum shoot and root length elongation was observed at 25 mg/L of AgNPs concentration. Secondary metabolites increased	(Ahmed and Murtaza, 2024)
<i>Echinops macrochaetus</i>	Salinity stress	40 µmol/L	Biomass Shoot length Chlorophyll content	Faster germination with increase in biomass, shoot length and chlorophyll	(Khan et al., 2024)

				content was observed at 40 $\mu\text{mol/L}$ AgNPs	
<i>Medicago sativa L.</i>		25 mg/l	Seed germination Root and shoot length	Significant increase in water absorption rate. Thus, increasing seed germination along with increased root and shoot length.	(Song et al., 2022)
<i>Linum usitatissimum L.</i>	Salinity stress	AgNPs combined with <i>C. testosteroni</i>	Overall growth and yield in saline condition.	increases production of photosynthetic pigments, proteins, sugars, and proline	(Khalofah et al., 2021)
<i>Ocimum basilicum L.</i>		200 mg/L	germination percentage and root length	Germination increased upto 85% and root length increased by 19.4 mm.	(Sencan et al., 2024)
<i>Helianthus annuus L.</i>		25 mg L ⁻¹	Overall growth and plant characteristics	Improved morphological characteristic, seed quality,	(Haq et al., 2024)

				and oil content, secondary metabolites	
<i>Solanum lycopersicum</i>		25–50 µg/mL	seed germination, seedling growth	increase in the seed germination percentage	(Sonawane et al., 2021)
<i>Vicia faba</i>	Aged seeds	100 ppm	vigor and mitotic index	Increased mitotic cell cycle and seedling growth	(Younis et al., 2019)
<i>Capsicum annuum L.</i>		30-50 ppm	root length, shoot length, seedling length, and germination vigour index	primed seedlings increased root and shoot length, seedling length, and germination vigour index	(Mawale and Giridhar, 2024)

Conclusions

Medicinal and herbal plants have been essential to the global evolution of human cultures. Their proper growth helps in development of socio-economic aspects of any society. Seedling establishment and germination are important phases in their life cycle. The efficient seed germination encourages successful establishment and deep root system. Seed priming is the process of treating seeds with chemical or biological priming agents to display better growth and to amplify abiotic stress resistance. The use of nanoprimering has been proven to be more promising than traditional priming approaches for achieving feasible agricultural yields. This article discussed the use of silver nanoparticles (AgNPs) as nanoprimering agent to enhance the seed germination of many medicinal and herbal plants. The future lies in the

utility of silver nanoparticle in seed priming with proper applications, appropriate ‘stop’ time, and better prior standardization. The seed priming adoption at field level is the need for future studies.

References

- Abasi, F., Raja, N.I., Mashwani, Z.U.R., Amjad, M.S., Ehsan, M., Mustafa, N., Haroon, M., Proćków, J., 2022. Biogenic silver nanoparticles as a stress alleviator in plants: A mechanistic overview. *Molecules* 27(11), 3378. <https://doi.org/10.3390/molecules27113378>
- Abdel Latef, A.A.H., Abu Alhmad, M.F., Abdelfattah, K.E., 2017. The possible roles of priming with ZnO nanoparticles in mitigation of salinity stress in lupine (*Lupinus termis*) plants. *Journal of Plant Growth Regulation* 36, 60-70. <https://doi.org/10.1007/s00344-016-9618-x>.
- Agidew, M.G., 2022. Phytochemical analysis of some selected traditional medicinal plants in Ethiopia. *Bulletin of the National Research Centre* 46(1), 87. <https://doi.org/10.1186/s42269-022-00770-8>
- Ahmad, B., Shabbir, A., Jaleel, H., Khan, M.M.A., Sadiq, Y., 2018. Efficacy of titanium dioxide nanoparticles in modulating photosynthesis, peltate glandular trichomes and essential oil production and quality in *Mentha piperita* L. *Current Plant Biology* 13, 6-15. <https://doi.org/10.1016/j.cpb.2018.04.002>
- Ahmed, M.J., Murtaza, G., 2024. Differential growth and biochemical responses induced by benevolent silver nanoparticles in germinating seeds and seedling of *Phaseolus vulgaris* L. *Environment, Development and Sustainability*, 1-16. <https://doi.org/10.1007/s10668-024-05208-x>
- Alhammad, B.A., Abdel-Aziz, H.M.M., Seleiman, M.F., Tourky, S.M.N., 2023. How Can Biological and Chemical Silver Nanoparticles Positively Impact Physio-Chemical and Chloroplast Ultrastructural Characteristics of *Vicia faba* Seedlings?. *Plants* 12, 2509. <https://doi.org/10.3390/plants1213250>
- Almutairi, Z. M., Alharbi, A., 2015. Effect of silver nanoparticles on seed germination of crop plants. *International Journal of Nuclear and Quantum Engineering* 9(6), 689-693. <https://doi.org/10.24297/jaa.v4i1.4295>
- Alshehddi, L.A.A., Bokhari, N., 2020. Influence of gold and silver nanoparticles on the germination and growth of *Mimusops laurifolia* seeds in the South-Western regions in Saudi Arabia. *Saudi Journal of Biological Sciences* 27(1), 574-580. <https://doi.org/10.1016/j.sjbs.2019.11.013>.

- Amooaghaiea, R., Tabatabaiea, F., Ahadia, A.M., 2015. Role of hematin and sodium nitroprusside in regulating *Brassica nigra* seed germination under nanosilver and silver nitrate stresses. *Ecotoxicology and Environmental Safety* 113, 259-270. <https://doi.org/10.1016/j.ecoenv.2014.12.017>.
- Anand, A., Kumari, A., Thakur, M., Koul, A., 2019. Hydrogen peroxide signaling integrates with phytohormones during the germination of magnetoprimed tomato seeds. *Scientific Reports* 9, 8814. <https://doi.org/10.1038/s41598-019-45102-5>
- Azimi, R., Borzelabad, M.J., Feizi, H., Azimi, A., 2014. Interaction of SiO₂ nanoparticles with seed prechilling on germination and early seedling growth of tall wheatgrass (*Agropyron elongatum* L.). *Polish Journal of Chemical Technology* 16(3), 25-29.
- Barboza da Silva, C., Marcos-Filho, J., Jourdan, P., Bennett, M.A., 2015. Performance of bell pepper seeds in response to drum priming with addition of 24-Epibrassinolide. *HortScience* 50, 873–878. <https://doi.org/10.21273/HORTSCI.50.6.873>
- Bareke, T., 2018. Biology of seed development and germination physiology. *Advances in Plants & Agriculture Research* 8, 336–346. DOI: 10.15406/apar.2018.08.00336
- Baskar, V., Venkatesh, J., Park, S.W., 2015. Impact of biologically synthesized silver nanoparticles on the growth and physiological responses in *Brassica rapa* ssp. *pekinensis*. *Environmental Science and Pollution Research* 22, 17672-17682. <https://doi.org/10.1007/s11356-015-4864-1>.
- Biswas, S., Seal, P., Majumder, B., Biswas, A.K., 2023. Efficacy of seed priming strategies for enhancing salinity tolerance in plants: An overview of the progress and achievements. *Plant Stress*, 100186. <https://doi.org/10.1016/j.stress.2023.100186>
- Boncheva, M., Gracias, DH., Jacobs, HO., Whitesides, GM., 2002. Biomimetic self-assembly of a functional asymmetrical electronic device. *Proceedings of the National Academy of Sciences* 99(8), 4937–4940. <https://doi.org/10.1073/pnas.032667599>
- Cao, Y., Jin, R., Mirkin, CA., 2001. DNA-modified core-shell Ag/Au nanoparticles. *Journal of the American Chemical Society* 123(32), 7961–7962. <https://doi.org/10.1021/ja011342n>
- Carrera-Castaño, G., Calleja-Cabrera, J., Pernas, M., Gómez, L., Oñate-Sánchez, L., 2020. An updated overview on the regulation of seed germination. *Plants* 9(6), 703. <https://doi.org/10.3390/plants9060703>.
- Caruthers, SD., Wickline, SA., Lanza, GM., 2007. Nanotechnological applications in medicine. *Current Opinion in Biotechnology* 18(1), 26–30. <https://doi.org/10.1016/j.copbio.2007.01.006>

- Chandrasekaran, U., Luo, X., Wang, Q., Shu, K., 2020. Are there unidentified factors involved in the germination of nanoprimered seeds ?. *Frontiers in Plant Science* 11, 832. <https://doi.org/10.3389/fpls.2020.00832>
- Chen, K., Arora, R., 2013. Priming memory invokes seed stress-tolerance. *Environmental and Experimental Botany* 94, 33-45. <https://doi.org/10.1016/j.envexpbot.2012.03.005>
- Chen, K., Fessehaie, A., Arora, R., 2012. Dehydrin metabolism is altered during seed osmopriming and subsequent germination under chilling and desiccation in *Spinacia oleracea* L. cv. Bloomsdale: possible role in stress tolerance. *Plant Science* 183, 27-36.
- Cheng, Z., Bradford, K.J., 1999. Hydrothermal time analysis of tomato seed germination responses to priming treatments. *Journal of Experimental of Botany* 33, 89–99. <https://doi.org/10.1093/jxb/50.330.89>
- Chung IllMin, C.I., Kaliyaperumal Rekha, K.R., Baskar Venkidasamy, B.V., Muthu Thiruvengadam, M.T., 2019. Effect of copper oxide nanoparticles on the physiology, bioactive molecules, and transcriptional changes in *Brassica rapa ssp. rapa* seedlings. *Water, Air, and Soil Pollution* 230, 48. <https://doi.org/10.1007/s11270-019-4084-2>.
- Conrath, U., 2011. Molecular aspects of defense priming. *Trends in Plant Science* 16(10), 524–531. DOI: 10.1016/j.tplants.2011.06.004
- Dalil, B., 2014. Response of medicinal plants to seed priming: a review. *International Journal of Plant, Animal and Environmental Sciences* 4(2), 741-745.
- De La Torre-Roche, R., Cantu, J., Tamez, C., Zuverza-Mena, N., Hamdi, H., Adisa, I.O., Elmer, W., Gardea-Torresdey, J., White, J.C., 2020. Seed biofortification by engineered nanomaterials: a pathway to alleviate malnutrition? *Journal of Agricultural and Food Chemistry* 68(44), 12189-12202. <https://doi.org/10.1021/acs.jafc.0c04881>
- Debbarna, M., Das, S.P., 2017. Priming of Seed: Enhancing Growth and Development. *International Journal of Current Microbiology and Applied Sciences* 6(12), 2390-2396. <https://doi.org/10.20546/ijemas.2017.612.276>
- Dutta, P., 2018. Seed priming: new vistas and contemporary perspectives. In: *Advances in seed priming*. Springer, Singapore, 3–22. https://doi.org/10.1007/978-981-13-0032-5_1
- Ellis, R.H., Roberts, E.H., 1981. The quantification of ageing and survival in orthodox seeds. *Seed Science & Technology* 9, 373-409.
- El-Temsah, Y.S., Joner, E.J., 2012. Impact of Fe and Ag nanoparticles on seed germination and differences in bioavailability during exposure in aqueous suspension and soil. *Environmental Toxicology* 27, 42-49. <https://doi.org/10.1002/tox.20610>

- Fahad, S., Bajwa, A.A., Nazir, U., Anjum, S.A., Farooq, A., Zohaib, A., Sadia, S., Nasim, W., Adkins, S., Saud, S., Ihsan, M.Z., 2017. Crop production under drought and heat stress: plant responses and management options. *Frontiers in Plant Science* 8, 1147. <https://doi.org/10.3389/fpls.2017.01147>
- Faizan, M., Faraz, A., Mir, A.R., Hayat, S., 2021. Role of zinc oxide nanoparticles in countering negative effects generated by cadmium in *Lycopersicon esculentum*. *Journal of Plant Growth Regulation* 40, 101-115. <https://doi.org/10.1007/s00344-019-10059-2>.
- FAO, 2016. Food and Agriculture: Key to Achieving the 2030 Agenda for Sustainable Development. Job No. I5499, Food and Agriculture Organization of the United Nations, Rome. Available online at: <http://www.fao.org/3/a-i5499e.pdf>.
- Faraji, J., Sepehri, A., 2019. Ameliorative effects of TiO₂ nanoparticles and sodium nitroprusside on seed germination and seedling growth of wheat under PEG-stimulated drought stress. *Journal of Seed Science* 41, 309-317. <https://doi.org/10.1590/2317-1545v41n3213139>.
- Farmahini Farahani, A., Tavili, A., Azarnivand, H., Jafari, A., 2022. Effect of priming and nano particles application on seedling emergence, establishment, growth and physiological characteristics of *Sanguisorba minor* Scop. and *Agropyron intermedium* (Host) P. Beauv forage species under drought stress in natural field. *Rangeland* 16(2), 236-255. <http://rangelandsrm.ir/article-1-683-en.html>
- Fercha, A., Capriotti, A.L., Caruso, G., Cavaliere, C., Gherroucha, H., Samperi, R., Stampachiacchiere, S., Lagana, A., 2013. Gel-free proteomics reveal potential biomarkers of priming-induced salt tolerance in durum wheat. *Journal of Proteomics* 91, 486-499. <https://doi.org/10.1016/j.jprot.2013.08.010>
- Fercha, A., Capriotti, A.L., Caruso, G., Cavaliere, C., Samperi, R., Stampachiacchiere, S., Laganà, A., 2014. Comparative analysis of metabolic proteome variation in ascorbate-primed and unprimed wheat seeds during germination under salt stress. *Journal of Proteomics* 108, 238-257. <https://doi.org/10.1016/j.jprot.2014.04.040>
- Fraceto, L.F., Grillo, R., de Medeiros, G.A., Scognamiglio, V., Rea, G., Bartolucci, C., 2016. Nanotechnology in agriculture: which innovation potential does it have?. *Frontiers in Environmental Science* 4, 186737. doi: 10.3389/fenvs.2016.00020
- Furmanczyk, E. M., Kozacki, D., Hyk, W., Muszyńska, M., Sekrecka, M., & Skwiercz, A. T. (2025). In Vitro Study on Nematicidal Effect of Silver Nanoparticles Against *Meloidogyne incognita*. *Molecules*, 30(5), 1132.

- Gallardo, K., Job, C., Groot, S.P., Puype, M., Demol, H., Vandekerckhove, J., Job, D., 2001. Proteomics of *Arabidopsis* seed germination and priming. *Plant Physiology* 126, 835–848. <https://doi.org/10.1079/9780851996530.01>
- Geisler-Lee, J., Wang, Q., Yao, Y., Zhang, W., Geisler, M., Li, K., Huang, Y., Chen, Y., Kolmakov, A., Ma, X., 2013. Phytotoxicity, accumulation and transport of silver nanoparticles by *Arabidopsis thaliana*. *Nanotoxicology* 7(3), 323–337. <https://doi.org/10.3109/17435390.2012.658094>
- Ghafari, H., Razmjoo, J., 2013. Effect of foliar application of nano-iron oxidase, iron chelate and iron sulphate rates on yield and quality of wheat. *International Journal of Agronomy and Plant Production* 4(11), 2997–3003.
- Gohari, G., Mohammadi, A., Akbari, A., Panahirad, S., Dadpour, M.R., Fotopoulos, V., Kimura, S., 2020. Titanium dioxide nanoparticles (TiO₂ NPs) promote growth and ameliorate salinity stress effects on essential oil profile and biochemical attributes of *Dracocephalum moldavica*. *Scientific Reports* 10(1), 912. <https://doi.org/10.1038/s41598-020-57794-1>.
- Han, M., Gao, X., Su J.Z., Nie S., 2001. Quantum-dot-tagged microbeads for multiplexed optical coding of biomolecules. *Nature Biotechnology* 19, 631–635. <https://doi.org/10.1038/90228>
- Haq, S.I.U., Wali, S., Sama, N.U., Kamran, K., Ullah, Z., & Mohamed, H.I., 2024. Environmentally friendly synthesis of silver nanoparticles (AgNPs) using *Mentha arvensis* plants modulates physiological and biochemical attributes and yield of sunflower (*Helianthus annuus* L.). *Journal of Soil Science and Plant Nutrition* 24(2), 3610–3630. <https://doi.org/10.1007/s42729-024-01781-2>
- He, M., He, C.-Q., Ding, N.-Z., 2018. Abiotic stresses: general defenses of land plants and chances for engineering multi stress tolerance. *Frontiers of Plant Science* 9, 1771. <https://doi.org/10.3389/fpls.2018.01771>
- Hojjat, S.S., Ganjali, A., 2016. The effect of silver nanoparticle on lentil seed germination under drought stress. *International Journal of Farming Allied Sciences* 5(3), 208–212.
- Hosainzadegan, H., Delfan, B., 2009. Evaluation of antibiofilm activity of dentol. *Acta Medica Iranica* 47, 35–40.
- Hullmann, A., 2007. Measuring and assessing the development of nano technology. *Scientometrics* 70, 739–758. <https://doi.org/10.1007/s11192-007-0310-6>
- Humera Razzaq, H.R., Tahir, M.H.N., Sadaqat, H.A., Bushra Sadia, B.S., 2017. Screening of sunflower (*Helianthus annuus* L.) accessions under drought stress conditions, an experimental assay. *Journal of Soil Science and Plant Nutrition* 17, 662–671.

- Imran, Q.M., Falak, N., Hussain, A., Mun, B.-G., Yun, B.-W., 2021. Abiotic stress in plants; stress molecular response and role of biotechnological tools in stress rese. *Agronomy* 11, 1579. <https://doi.org/10.3390/agronomy11081579>
- Ingle, A.P., Biswas, A., Vanlalveni, C., Lalfakzuala R., Gupta, I., Ingle, P., Rokhum L., Rai, M., 2020. Biogenic synthesis of nanoparticles and their role in the management of plant pathogenic fungi. *Microbial Nanotechnology*, 135–161. <https://doi.org/10.4324/9780429276330-8>.
- Islam, M.M., Kayesh, E., Zaman, E., Urmi, T.A., Haque, M.M., 2018. Evaluation of rice (*Oryza sativa* L.) genotypes for drought tolerance at germination and early seedling stage. *Agriculturists* 16, 44–54.
- Jadczak, P., Kulpa, D., Drozd, R., Przewodowski, W., Przewodowska, A., 2020. Effect of AuNPs and AgNPs on the antioxidant system and antioxidant activity of lavender (*Lavandula angustifolia* Mill.) from in vitro cultures. *Molecules* 25(23), 5511. <https://doi.org/10.3390/molecules25235511>
- Jiang, H., Li, M., Chang, F., Li, W., Yin, L., 2012. Physiological analysis of silver nanoparticles and AgNO₃ toxicity to *Spirodela polyrrhiza*. *Environmental Toxicology and Chemistry* 31(8), 1880–1996. <https://doi.org/10.1002/etc.1899>
- Jiang, M., Song, Y., Kanwar, M.K., Ahammed, G.J., Shao, S., Zhou, J., 2021. Phytonanotechnology applications in modern agriculture. *Journal of Nanobiotechnology* 19, 1-20. <https://doi.org/10.1186/s12951-021-01176-w>
- Johnson, R., Puthur, J.T., 2021. Seed priming as a cost-effective technique for developing plants with cross tolerance to salinity stress. *Plant Physiology and Biochemistry* 162, 247–257. <https://doi.org/10.1016/j.plaphy.2021.02.034>.
- Kamat, P.V., 2002. Photophysical, photochemical and photocatalytic aspects of metal nanoparticles. *The Journal of Physical Chemistry B* 106, 7729–7744. <https://doi.org/10.1021/jp0209289>
- Kamle, M., Mahato, D.K., Devi, S., Soni, R., Tripathi, V., Mishra, A.K., Kumar, P., 2020. Nanotechnological interventions for plant health improvement and sustainable agriculture. *3 Biotech* 10(4), 168. <https://doi.org/10.1007/s13205-020-2152-3>
- Kasim, W.A., Osman, M.E., Omar, M.N., El-Daim, I.A.A., Bejai, S., Meijer, J., 2013. Control of drought stress in wheat using plant growth promoting bacteria. *Journal of Plant Growth Regulation* 32, 122–130. <https://doi.org/10.1007/s00344-012-9283-7>
- Kaveh, R., Li, Y.S., Ranjbar, S., Tehrani, R., Brueck, C.L., Van Aken, B., 2013. Changes in *Arabidopsis thaliana* gene expression in response to silver nanoparticles and silver

- ions. Environmental Science & Technology 47(18), 10637-10644. <https://doi.org/10.1021/es402209w>
- Kaya, M.D., Okcub, G., Ataka, M., Cikilic, Y., Kolsaricia, O., 2006. Seed treatments to overcome salt and drought stress during germination in sunflower (*Helianthus annuus* L.). European Journal of Agronomy 24, 291–295. <https://doi.org/10.1016/j.eja.2005.08.001>
- Kędziora, A., Speruda, M., Krzyżewska, E., Rybka, J., Łukowiak, A., & Bugła-Płoskońska, G. (2018). Similarities and differences between silver ions and silver in nanoforms as antibacterial agents. International journal of molecular sciences 19(2), 444.
- Khaeim, H., Kende, Z., Balla, I., Gyuricza, C., Eser, A., Tarnawa, Á., 2022. The effect of temperature and water stresses on seed germination and seedling growth of wheat (*Triticum aestivum* L.). Sustainability 14(7), 3887. <https://doi.org/10.3390/su14073887>
- Khalaki, M.A., Moameri, M., Lajayer, B.A., Astatkie, T., 2021. Influence of nano-priming on seed germination and plant growth of forage and medicinal plants. Plant Growth Regulation 93(1), 13-28. <https://doi.org/10.1007/s10725-020-00670-9>
- Khalofah, A., Kilany, M., Migdadi, H., 2021. Phytostimulatory Influence of *Comamonas testosteroni* and Silver Nanoparticles on *Linum usitatissimum* L. under Salinity Stress. Plants 10(4), 790. <https://doi.org/10.3390/plants10040790>
- Khan, H., 2014. Medicinal plants in light of history: recognized therapeutic modality. Journal of evidence-based complementary & alternative medicine 19(3), 216-219. <https://doi.org/10.1177/2156587214533346>
- Khan, I., Raza, M.A., Awan, S.A., Shah, G.A., Rizwan, M., Ali, B., Tariq, R., Hassan, M.J., Alyemeni, M.N., Brestic, M., Zhang, X., Ali, S., Huang, L., 2020. Amelioration of salt induced toxicity in pearl millet by seed priming with silver nanoparticles (AgNPs): the oxidative damage, antioxidant enzymes and ions uptake are major determinants of salt tolerant capacity. Plant Physiology and Biochemistry 156, 221–232. <https://doi.org/10.1016/j.plaphy.2020.09.018>.
- Khan, S., Al-Qurainy, F., Al-hashimi, A., Nadeem, M., Tarroum, M., Salih, A.M., Shaikhaldein, O.H., Alfarraj, N.S., 2024. Promoting effect of biogenic AgNPs on *Echinops macrochaetus* for improving growth parameters and mitigating the toxic effect caused by salinity stress. Biotechnology & Biotechnological Equipment 38(1), 2354710. <https://doi.org/10.1080/13102818.2024.2354710>
- Khan, S., Zahoor, M., Khan, R.S., Ikram, M., Islam, N.U., 2023. The impact of silver nanoparticles on the growth of plants: The agriculture applications. Heliyon 9(6), e16928. DOI: 10.1016/j.heliyon.2023.e16928

- Kornarzyński, K., Sujak, A., Czernel, G., Wiącek, D., 2020. Effect of Fe₃O₄ nanoparticles on germination of seeds and concentration of elements in *Helianthus annuus* L. under constant magnetic field. *Scientific Reports* 10(1), 8068. <https://doi.org/10.1038/s41598-020-64849-w>.
- Krishnasamy, R., Natesh, R., Obbineni, J.M. 2024. Efficient ROS Scavenging Improves the Growth and Yield in Black Gram (*Vigna mungo* (L.) Hepper) after Seed Priming and Treatment using Biosynthesized Silver Nanoparticles with *Pongamia pinnata* (L.) Pierre Leaf Extract. *Journal of Plant Growth Regulation*, 1-17. <https://doi.org/10.1007/s00344-024-11276-0>
- Kumar, V.K., Muthukrishnan, S., Rajalakshmi, R., 2020. Phytostimulatory effect of phytochemical fabricated nanosilver (AgNPs) on *Psophocarpus tetragonolobus* (L.) DC. seed germination: An insight from antioxidative enzyme activities and genetic similarity studies. *Current Plant Biology* 23, 100158. <https://doi.org/10.1016/j.cpb.2020.100158>
- Liu, C., Zhou, H., Zhou, J., 2021. The applications of nanotechnology in crop production. *Molecules* 26(23), 7070. <https://doi.org/10.3390/molecules26237070>
- Lobell, D.B., Schlenker, W., Costa-Roberts, J., 2011. Climate trends and global crop production since 1980. *Science* 333, 616–620. [10.1126/science.1204531](https://doi.org/10.1126/science.1204531)
- Mahra, S., Tripathi, S., Tiwari, K., Sharma, S., Mathew, S., Kumar, V., Sharma, S., 2024. Harnessing nanotechnology for sustainable agriculture: From seed priming to encapsulation. *Plant Nano Biology* 11, 100124. <https://doi.org/10.1016/j.plana.2024.100124>
- Maier, S.A., Brongersma, M.L., Kik, P.G., Meltzer, S., Requicha, A.A., Atwater, H.A., 2001. Plasmonics—a route to nanoscale optical devices. *Advanced Materials* 13, 1501–1505.
- Maroufi, K., Farahani, H.A., Moradi, O., 2011. Evaluation of nano priming on germination percentage in green gram (*Vigna radiata* L.). *Advances in Environmental Biology*, 3659-3664.
- Marrelli, M., 2021. Medicinal plants. *Plants* 10(7), 1355. <https://doi.org/10.3390/plants10071355>
- Mathur, P., Jha, S., Ramteke, S., & Jain, N. K. (2018). Pharmaceutical aspects of silver nanoparticles. *Artificial cells, nanomedicine, and biotechnology*, 46, 115-126.
- Matras, E., Gorczyca, A., Przemieniecki, S.W., Cwieja, M.O., 2022. Surface properties-dependent antifungal activity of silver nanoparticles, *Scientific Reports* 12(1), 18046. <https://doi.org/10.1038/s41598-022-22659-2>

- Mawale, K., S., Giridhar, P., 2024. Green approaches for the synthesis of silver nanoparticle and its augmentation in Seed germination, growth, and antioxidant level in *Capsicum annuum* L. *Plant Nano Biology* 10, 100107. <https://doi.org/10.1016/j.plana.2024.100107>
- McDonald, M.B., 2000. Seed priming. In: Black M, Bewley JD (eds) *Seed technology and its biological basis*. Sheffield Academic Press, Sheffield, 287–325
- Mirkin, C.A., Letsinger, R.L, Mucic, R.C., Storhoff, J.J., 1996. A DNA-based method for rationally assembling nanoparticles into macroscopic materials. *Nature* 382, 607–609. doi:10.1038/382607a0
- Montanha, G.S., Rodrigues, E.S., Marques, J.P.R., de Almeida, E., Colzato, M., Pereira de Carvalho, H.W., 2020. Zinc nanocoated seeds: an alternative to boost soybean seed germination and seedling development. *SN Applied Sciences* 2(5), 857. <https://doi.org/10.1007/s42452-020-2630-6>.
- Muhammad, I., Kolla, M., Volker, R., Günter, N., 2015. Impact of nutrient seed priming on germination, seedling development, nutritional status and grain yield of maize. *Journal of Plant Nutrition* 38(12), 1803-1821. <https://doi.org/10.1080/01904167.2014.990094>
- Mustapha, T., Misni, N., Ithnin, N.R., Daskum, A.M., Unyah, N.Z., 2022. A review on plants and microorganisms mediated synthesis of silver nanoparticles, role of plants metabolites and applications. *International Journal of Environmental Research and Public Health* 19(2), 674.
- Nejatzadeh, F., 2021. Effect of silver nanoparticles on salt tolerance of *Satureja hortensis* L. during in vitro and in vivo germination tests. *Heliyon* 7(2). <https://doi.org/10.1016/j.heliyon.2021.e05981>
- Nile, S.H., Thiruvengadam, M., Wang, Y., Samynathan, R., Shariati, M.A., Rebezov, M., Nile, A., Sun, M., Venkidasamy, B., Xiao, J., Kai, G., 2022. Nano-priming as emerging seed priming technology for sustainable agriculture—recent developments and future perspectives. *Journal of Nanobiotechnology* 20(1), 254. <https://doi.org/10.1186/s12951-022-01423-8>
- Okçu, G., Kaya, M.D., Atak, M., 2005. Effects of salt and drought stresses on germination and seedling growth of pea (*Pisum sativum* L.). *Turkish Journal of Agriculture and Forestry* 29(4), 237-242. <https://journals.tubitak.gov.tr/agriculture/vol29/iss4/2>
- one 14(4), e0214480. <https://doi.org/10.1371/journal.pone.0214480>
- Oukarroum, A., Barhoumi, L., Pirastru L., Dewez, D., 2013. Silver nanoparticle toxicity effect on growth and cellular viability of the aquatic plant *Lemna gibba*. *Environmental Toxicology and Chemistry* 32(4), 902-7. <https://doi.org/10.1002/etc.2131>

- Parveen, A., Rao, S., 2014. Effect of nanosilver on seed germination and seedling growth in *Pennisetum glaucum*. *Journal of Cluster Science* 26, 693-701. DOI 10.1007/s10876-014-0728-y.
- Patidar, P., Kumar, P., Parmar, S.S., Khan, N., 2024. Influence of Silver Nanoparticles on Onion (*Allium cepa* L.) Seed Germination and Seedling Vigor. *Journal of Advances in Biology & Biotechnology* 27(8), 10-16. <https://doi.org/10.9734/jabb/2024/v27i81116>
- Pramanik, P., Krishnan, P., Maity, A., Mridha, N., Mukherjee, A., Rai, V., 2020. Application of nanotechnology in agriculture. *Environmental Nanotechnology* 4, 317-348. https://doi.org/10.1007/978-3-030-26668-4_9
- Prasad, R., Bhattacharyya, A., Nguyen, Q.D., 2017. Nanotechnology in sustainable agriculture: recent developments, challenges, and perspectives. *Frontiers in Microbiology* 8, 1014. <https://doi.org/10.3389/fmicb.2017.01014>
- Pulizzi, F., 2019. Nano in the future of crops. *Nature Nanotechnology* 14(6), 507-507. <https://doi.org/10.1038/s41565-019-0475-1>
- Qian HaiFeng, Q.H., Peng XiaoFeng, P.X., Han Xiao, H.X., Ren Jie, R.J., Sun LiWei, S.L., Fu ZhengWei, F.Z., 2013. Comparison of the toxicity of silver nanoparticles and silver ions on the growth of terrestrial plant model *Arabidopsis thaliana*. *Journal of Environmental Sciences* 25, 1947- 1955. [https://doi.org/10.1016/S1001-0742\(12\)60301-5](https://doi.org/10.1016/S1001-0742(12)60301-5)
- Rafique, M., Sadaf, I., Rafique, M. S., Tahir, M. B., 2016. A review on green synthesis of silver nanoparticles and their applications. *Artificial Cells, Nanomedicine, and Biotechnology* 45(7), 1272–1291. <https://doi.org/10.1080/21691401.2016.1241792>
- Rahimi, D., Kartoolinejad, D., Nourmohammadi, K., Naghdi, R., 2016. Increasing drought resistance of *Alnus subcordata* C.A. Mey. seeds using a nano priming technique with multi-walled carbon nanotubes. *Journal of Forest Science* 62(6), 269–78. <https://doi.org/10.17221/15/2016-JFS>.
- Rai, M., Yadav, A., Gade, A., 2009. Silver nanoparticles as a new generation of antimicrobials. *Biotechnology Advances* 27(1), 76–83. <https://doi.org/10.1016/j.biotechadv.2008.09.002>
- Raj, A.B., Raj, S.K., 2019. Seed priming: An approach towards agricultural sustainability. *Journal of Applied and Natural Science* 11(1), 227-234. <https://doi.org/10.31018/jans.v11i1.2010>
- Rajjou, L., Belghazi, M., Huguet, R., Robin, C., Morea, A., Job, C., Job, D., 2006. Proteomic investigation of the effect of salicylic acid on *Arabidopsis* seed germination and

- establishment of early defense mechanisms. *Plant Physiology* 141, 910–923. <https://doi.org/10.1104/pp.106.082057>
- Rajwade, J.M., Chikte, R.G., Paknikar, K.M., 2020. Nanomaterials: new weapons in a crusade against phytopathogens. *Applied microbiology and biotechnology* 104(4), 1437-1461. <https://doi.org/10.1007/s00253-019-10334-y>
- Rashid, A., Harris, D., Hollington, P., Ali, S., 2004. On-farm seed priming reduces yield losses of mungbean (*Vigna radiata*) associated with mungbean yellow mosaic virus in the North West Frontier Province of Pakistan. *Crop Protection* 23, 1119-1124. <https://doi.org/10.1016/j.cropro.2004.04.002>
- Rhaman, M.S., Rauf, F., Tania, S.S., Khatun, M., 2020. Seed priming methods: Application in field crops and future perspectives. *Asian Journal of Research in Crop Science* 5(2), 8-19. DOI: 10.9734/AJRCS/2020/v5i230091
- Rizzardi, M.A., Luiz, A.R., Roman, E.S., Vargas, L., 2009. Effect of cardinal temperature and water potential on morning glory (*Ipomoea triloba*) seed germination. *Planta Daninha* 27, 13-21. <https://doi.org/10.1590/S0100-83582009000100003>
- Salama, H.M.H., 2012. Effects of silver nanoparticles in some crop plants, common bean (*Phaseolus vulgaris* L.) and corn (*Zea mays* L.). *International Resesarch Journal of Biotechnology* 3(10), 190-197.
- Savage, N., Diallo, M.S., 2005. Nanomaterials and water purification: opportunities and challenges. *Journal of Nanoparticle Research* 7, 331–342. DOI 10.1007/s11051-005-7523-5
- Savithramma, N., Ankanna, S., Bhumi, G., 2012. Effect of nanoparticles on seed germination and seedling growth of *Boswellia ovalifoliolata* – an endemic and endangered medicinal tree taxon. *Nano Vision* 2(1), 61-68.
- Savithramma, N., Rao, M.L., Rukmini, K., Devi, P.S., 2011. Antimicrobial activity of silver nanoparticles synthesized by using medicinal plants. *International Journal of ChemTech Research* 3, 1394–1402.
- Schilling, K.E., 2016. Comment on “Climate and agricultural land use change impacts on streamflow in the upper midwestern United States” by Satish C. Gupta et al. *Water Resources Research* 52(7), 5694-5696. <https://doi.org/10.1002/2015WR018482>
- Sencan, A., Kilic, S., Kaya, H. 2024., Stimulating effect of biogenic nanoparticles on the germination of basil (*Ocimum basilicum* L.) seeds. *Scientific Reports* 14(1), 1715. <https://doi.org/10.1038/s41598-023-50654-8>
- Shafiq, F., Iqbal, M., Ali, M., Ashraf, M.A., 2019. Seed pre-treatment with polyhydroxy fullerene nanoparticles confer salt tolerance in wheat through upregulation of H₂O₂

neutralizing enzymes and phosphorus uptake. *Journal of Soil Science and Plant Nutrition* 19, 734-742. <https://doi.org/10.1007/s42729-019-00073-4>.

Shan, G., Surampalli, R.Y., Tyagi, R.D., Zhang, T.C., 2009. Nanomaterials for environmental burden reduction, waste treatment, and nonpoint source pollution control: a review. *Frontiers of Environmental Science & Engineering in China* 3, 249–264. <https://doi.org/10.1007/s11783-009-0029-0>

Shang, Y., Hasan, M.K., Ahammed, G.J., Li, M., Yin, H., Zhou, J., 2019. Applications of nanotechnology in plant growth and crop protection: a review. *Molecules* 24(14), 2558. <https://doi.org/10.3390/molecules24142558>

Sharma, P., Bhatt, D., Zaidi, M.G.H., Paradha Saradhi, P., Khanna P.K., Arora, S., 2012. Silver nanoparticle-mediated enhancement in growth and antioxidant status of *Brassica juncea*. *Applied Biochemistry and Biotechnology* 167, 2225–2233. <https://doi.org/10.1007/s12010-012-9759-8>

Shelar, A., Nile, S.H., Singh, A.V., Rothenstein, D., Bill, J., Xiao, J., Chaskar, M., Kai, G., Patil, R., 2023. Recent advances in nano-enabled seed treatment strategies for sustainable agriculture: challenges, risk assessment, and future perspectives. *Nano-Micro Letters* 15(1), 54. <https://doi.org/10.1007/s40820-023-01025-5>

Shelar, A., Singh, A.V., Maharjan, R.S., Laux, P., Luch, A., Gemmati, D., Tisato, V., Singh, S.P., Santilli, M.F., Shelar, A., Chaskar, M., 2021. Sustainable agriculture through multidisciplinary seed nanoprimer: prospects of opportunities and challenges. *Cells* 10(9), 2428. <https://doi.org/10.3390/cells10092428>

Sonawane, H., Arya, S., Math, S., Shelke, D., 2021. Myco-synthesized silver and titanium oxide nanoparticles as seed priming agents to promote seed germination and seedling growth of *Solanum lycopersicum*: A comparative study. *International Nano Letters* 11(4), 371-379. <https://doi.org/10.1007/s40089-021-00346-w>

Song, K., Zhao, D., Sun, H., Gao, J., Li, S., Hu, T., He, X., 2022. Green nanoprimer: responses of alfalfa (*Medicago sativa* L.) seedlings to alfalfa extracts capped and light-induced silver nanoparticles. *BMC Plant Biology* 22(1), 323. <https://doi.org/10.1186/s12870-022-03692-9>

Srivastava, G., Das, C.K., Das, A., Singh, S.K., Roy, M., Kim, H., Sethy, N., Kumar, A., Sharma, R.K., Singh, S.K., Philip, D., 2014. Seed treatment with iron pyrite (FeS₂) nanoparticles increases the production of spinach. *RSC Advances* 4(102), 58495-58504. <https://doi.org/10.1039/c4ra06861k>.

- Stampoulis, D., Sinha, S.K., White, J.C., 2009. Assay-dependent phytotoxicity of nanoparticles to plants. *Environmental Science & Technology* 43, 9473–9479. <https://doi.org/10.1021/es901695c>
- Szopińska, D., Politycka, B., 2016. The effects of hydro-and osmopriming on the germination, vigour, and hydrolytic enzymes activity of common zinnia (*Zinnia elegans* Jacq.) seeds. *Folia Horticulture* 28, 3–11. <https://doi.org/10.1515/fhort-2016-0001>
- Taran, N., Storozhenko, V., Svetlova, N., Batsmanova, L., Shvartau, V., Kovalenko, M., 2017. Effect of zinc and copper nanoparticles on drought resistance of wheat seedlings. *Nanoscale Research Letters* 12, 1-6. <https://doi.org/10.1186/s11671-017-1839-9>.
- Thiruvengadam, M., Gurunathan, S., Chung, I.M., 2015. Physiological, metabolic, and transcriptional effects of biologically-synthesized silver nanoparticles in turnip (*Brassica rapa* ssp. *rapa* L.). *Protoplasma* 252, 1031-1046. <https://doi.org/10.1007/s00709-014-0738-5>.
- Trenberth, K.E., Dai, A., Van Der Schrier, G., Jones, P.D., Barichivich, J., Briffa, K.R., Sheffield, J., 2014. Global warming and changes in drought. *Nature Climate Change* 4, 17–22. <https://doi.org/10.1038/nclimate2067>
- Upadhyaya, H., Begum, L., Dey, B., Nath, P.K., Panda, S.K., 2017. Impact of calcium phosphate nanoparticles on rice plant. *Journal of Plant Science Phytopathology* 1, 1–10. <https://dx.doi.org/10.29328/journal.jpssp.1001001>
- Usman, M., Farooq, M., Wakeel, A., Nawaz, A., Cheema, S.A., ur Rehman, H., Ashraf, I., Sanaullah, M., 2020. Nanotechnology in agriculture: Current-status, challenges and future opportunities. *Science of The Total Environment* 721, 137778. <https://doi.org/10.1016/j.scitotenv.2020.137778>
- Valivand, M., Amooaghaie, R., Ahadi, A., 2019. Seed priming with H₂S and Ca²⁺ trigger signal memory that induces cross-adaptation against nickel stress in zucchini seedlings. *Plant Physiology and Biochemistry* 143, 286-298. <https://doi.org/10.1016/j.plaphy.2019.09.016>
- Van Nguyen, D., Nguyen, H.M., Le, N.T., Nguyen, K.H., Nguyen, H.T., Le, H.M., Nguyen, A.T., Dinh, N.T.T., Hoang, S.A., Van Ha, C., 2022. Copper nanoparticle application enhances plant growth and grain yield in maize under drought stress conditions. *Journal of Plant Growth Regulation* 41(1), 364-375. <https://doi.org/10.1007/s00344-021-10301-w>.
- Vanlalveni, C., Lallianrawna, S., Biswas, A., Selvaraj, M., Changmai, B., Rokhum, S.L., 2021. Green synthesis of silver nanoparticles using plant extracts and their antimicrobial activities: A review of recent literature. *RSC advances* 11(5), 2804-2837.

- Vishal, B., Kumar, P.P., 2018. Regulation of seed germination and abiotic stresses by gibberellins and abscisic acid. *Frontiers in Plant Science* 9, 838. <https://doi.org/10.3389/fpls.2018.00838>
- Vishwanath, R., Negi, B., 2021. Conventional and green methods of synthesis of silver nanoparticles and their antimicrobial properties. *Current Research in Green and Sustainable Chemistry* 4, 100205. <https://doi.org/10.1016/j.crgsc.2021.100205>
- Wang, W.Q., Liu, S.J., Song, S.Q., Møller, I.M., 2015. Proteomics of seed development, desiccation tolerance, germination, and vigor. *Plant Physiology and Biochemistry* 86, 1-15. <https://doi.org/10.1016/j.plaphy.2014.11.003>
- Waqas, M., Korres, N.E., Khan, M.D., Nizami, A.S., Deeba, F., Ali, I., Hussain, H., 2019. Advances in the concept and methods of seed priming. In: Hasanuzzaman, M., Fotopoulos, V. (eds) *Priming and Pretreatment of Seeds and Seedlings*. Springer, Singapore, 11-41. https://doi.org/10.1007/978-981-13-8625-1_2.
- Waqas, M.A., Kaya, C., Riaz, A., Farooq, M., Nawaz, I., Wilkes, A., Li, Y., 2019. Potential mechanisms of abiotic stress tolerance in crop plants induced by thiourea. *Frontiers in Plant Science* 10, 1336. <https://doi.org/10.3389/fpls.2019.01336>.
- Wu, L.M., Fang, Y., Yang, H.N., Bai, L.Y., 2019. Effects of drought-stress on seed germination and growth physiology of quinclorac-resistant *Echinochloa crusgalli*. *PloS*
- Yacoubi, R., Job, C., Belghazi, M., Chaibi, W., Job, D., 2013. Proteomic analysis of the enhancement of seed vigour in osmoprimed alfalfa seeds germinated under salinity stress. *Seed Science Research* 23(2), 99-110. doi:10.1017/S0960258513000093
- Yadav, S., Modi, P., Dave, A., Vijapura, A., Patel, D., Patel, M., 2020. Effect of abiotic stress on crops, in *Sustainable Crop Production*. ed. by M. Hasanuzzaman, M.C.M.T. Filho, M. Fujita, T.A.R. Nogueira (IntechOpen, London, 2020)
- Yan, A., Chen, Z., 2019. Impacts of silver nanoparticles on plants: a focus on the phytotoxicity and underlying mechanism. *International Journal of Molecular Sciences* 20(5) 23–25. <https://doi.org/10.3390/ijms20051003>.
- Yasur, J., Rani, P.U., 2013. Environmental effects of nanosilver: impact on castor seed germination, seedling growth, and plant physiology. *Environmental Science and Pollution Research* 20, 8636-48.
- Yin, L., Colman, B.P., McGill, B.M., Wright, J.P., Bernhardt, E.S., 2012. Effects of silver nanoparticle exposure on germination and early growth of eleven wetland plants. *Public Library of Sciences* 7(10), e47674, 2012. doi:10.1371/journal.pone.0047674

- Younis, M.E., Abdel-Aziz, H.M.M., Heikal, Y.M., 2019. Nanopriming technology enhances vigor and mitotic index of aged *Vicia faba* seeds using chemically synthesized silver nanoparticles. *South African Journal of Botany* 125, 393-401. <https://doi.org/10.1016/j.sajb.2019.08.018>
- Z€ach, M., H€agglund, C., Chakarov, D., Kasemo, B., 2006. Nanoscience and nanotechnology for advanced energy systems. *Current Opinion in Solid State and Materials Science* 10,132–143. <https://doi.org/10.1016/j.cossms.2007.04.004>
- Zare, A.R., Solouki, M., Omid, M., Irvani, N., Oladzad Abasabadi, A., Mahdi Nezhad, N., 2011. Effect of various treatments on seed germination and dormancy breaking in *ferula assa foetida*. *Trakia Journal of Sciences* 9, 57-61.
- Zhao, Y., Anjum, M.N., Pu, L., Wang, G., Luo, Y., 2018. Optimal on demand delay-constrained fair distribution for self-coexistence WRAN. *Computer Networks* 134, 260-271. <https://doi.org/10.1016/j.comnet.2018.02.001>
- Zulfiqar, F., Ashraf, M., 2021. Nanoparticles potentially mediate salt stress tolerance in plants. *Plant Physiology and Biochemistry* 160, 257–268. <https://doi.org/10.1016/j.plaphy.2021.01.028>.