

MASTER OF SCIENCE (BIOTECHNOLOGY)

[ DIVYA YADAV ]

2022

**IN SILICO ANALYSIS OF ANTIHISTAMINE  
DRUGS AS NEUROPROTECTANTS TARGETING  
DOPAMINE D2-LIKE RECEPTORS IN  
PARKINSON'S DISEASE**

A DISSERTATION

SUBMITTED IN PARTIAL FULFILLMENT OF THE  
REQUIREMENTS FOR THE AWARD OF THE DEGREE  
OF

MASTER OF SCIENCE  
IN  
**BIOTECHNOLOGY**

Submitted by:

**Divya Yadav**

**2K20/MSCBIO/06**

Under the supervision of:

**PROF. PRAVIR KUMAR**



**DEPARTMENT OF BIOTECHNOLOGY  
DELHI TECHNOLOGICAL UNIVERSITY  
(Formerly Delhi College of Engineering)**

Bawana Road, Delhi – 110042

**MAY, 2022**

DELHI TECHNOLOGICAL UNIVERSITY  
(Formerly Delhi College of Engineering)  
Bawana Road, Delhi – 110042

**CANDIDATE’S DECLARATION**

I, **Divya Yadav**, (Roll No.: 2K20/MSCBIO/06) hereby certify that the work which is presented in the Major Project entitled “**In silico analysis of antihistamine drugs as neuroprotectants targeting dopamine D2-like receptors in Parkinson’s disease**” in fulfilment of the requirement for the award of Degree of Masters of Science in Biotechnology and submitted to the Department of Biotechnology, Delhi Technological University, Delhi is an authentic record of my own work, carried during a period from 10-June-2021 to 01-May-2022, under the supervision of **Prof. Pravir Kumar**.

The matter presented in this thesis has not been submitted by me for the award of any other degree of this or any other university. This work has been communicated in Scopus indexed journal and conference with the following details:

**1. Title of Paper:** “Restoration and targeting of aberrant neurotransmitters in Parkinson's disease therapeutics.”

**Author’s Name:** **Divya Yadav** and Pravir Kumar

**Name of the Journal:** Neurochemistry International

**Conference date with venue (if applicable):** NA

**Have you registered for the conference (yes/no):** NA

**Status of the paper (Accepted/published/communicated):** Accepted

**Date of paper communication:** 14<sup>th</sup> January 2022

**Date of paper acceptance:** 17<sup>th</sup> March 2022

**Date of paper publication:** -

**2. Title of Paper:** “Parkinson’s Disease: An Overview and Role of Glutamate and its Receptors.”

**Author’s Name:** **Divya Yadav** and Pravir Kumar

**Name of Conference:** 5th International Conference on Information Systems and Computer Networks (ISCON), 2021

**Conference date with venue (if applicable):** October 22-23, 2021 (Virtual)

**Have you registered for the conference (yes/no):** Yes

**Status of paper:** Published

**Date of paper communicated:** 30<sup>th</sup> July, 2021

**Date of paper acceptance:** 9<sup>th</sup> October, 2021

**Date of paper publication:** 15<sup>th</sup> February, 2022

Place: New Delhi

Date:

**Divya Yadav**  
**(2K20/MSCBIO/06)**

**DEPARTMENT OF BIOTECHNOLOGY**  
**DELHI TECHNOLOGICAL UNIVERSITY**  
(Formerly Delhi College of Engineering)  
Bawana Road, Delhi – 110042

**CERTIFICATE**

To the best of my knowledge, the above work entitled **“In silico analysis of antihistamine drugs targeting Dopamine D2-like receptors in Parkinson’s disease”** has not been submitted in part or full for any Degree or Diploma to this University or elsewhere. I further certify that the publication and indexing information given by the student is correct.

Place: DTU, New Delhi

Date:

**Prof. Pravir Kumar**  
(SUPERVISOR)  
Head of Department  
Delhi Technological University

# PROOF OF PUBLICATION

## 1. Title of Paper: “Restoration and targeting of aberrant neurotransmitters in Parkinson's disease therapeutics.”


Neurochemistry International 156 (2022) 105327



Contents lists available at [ScienceDirect](#)

### Neurochemistry International

journal homepage: [www.elsevier.com/locate/neuint](http://www.elsevier.com/locate/neuint)



## Restoration and targeting of aberrant neurotransmitters in Parkinson's disease therapeutics



Divya Yadav<sup>a,b</sup> Pravir Kumar<sup>a,b,1,\*</sup>

<sup>a</sup> Molecular Neuroscience and Functional Genomics Laboratory, Department of Biotechnology, Delhi, India

<sup>b</sup> Delhi Technological University (Formerly Delhi College of Engineering), Delhi, 110042, India


### ARTICLE INFO



#### Keywords:

Neurotransmitter  
Neurotransmission  
Parkinson's disease  
Neurodegeneration


### ABSTRACT

Neurotransmitters are considered as a fundamental regulator in the process of neuronal growth, differentiation and survival. Parkinson's Disease (PD) occurs due to extensive damage of dopamine-producing neurons; this causes dopamine deficits in the midbrain, followed by the alternation of various other neurotransmitters (glutamate, GABA, serotonin, etc.). It has been observed that fluctuation of neurotransmission in the basal ganglia exhibits a great impact on the pathophysiology of PD. Dopamine replacement therapy, such as the use of L-DOPA, can increase the dopamine level, but it majorly ameliorates the motor symptoms and is also associated with long-term complications (for e.g., LID). While the non-dopaminergic system can efficiently target non-motor



About Elsevier Products & Solutions Services Shop & Discover   

Home > Journals > Neurochemistry International



### Neurochemistry International


The Journal of Cellular and Molecular Neuroscience

Publishing options: [OA Open Access](#) [Subscription](#)

[Guide for authors](#) [Track your paper](#) [Order journal](#)

ISSN: 0197-0186

**Editor-in-Chief** [Editorial board](#)

 Keiji Wada

Neurochemistry International is devoted to the rapid publication of outstanding original articles and timely reviews in **neurochemistry**. Manuscripts on a broad

[Submit your paper](#)

The Impact Factor of this journal is 3.921, ranking it 121 out of 273 in *Neurosciences*

With this journal indexed in 5 international databases, your published article can be read and cited by researchers worldwide

[View articles](#)

Home > Journals > Neurochemistry International > Abstracting & Indexing



### Neurochemistry International

[Submit your Paper](#) [View Articles](#)

[Guide for authors](#) [Track your paper](#) [Order journal](#)

ISSN: 0197-0186

### Abstracting & Indexing

- Science Citation Index
- Web of Science
- PubMed
- PubMed/Medline
- Scopus

## 2. Title of Paper: “Parkinson’s Disease: An Overview and Role of Glutamate and its Receptors.”

2021 5th International Conference on Information Systems and Computer Networks (ISCON)  
GLA University, Mathura, India. Oct 22-23, 2021

# Parkinson’s Disease: An Overview and Role of Glutamate and its Receptors

Glutamate receptor based therapy as an alternate way to treat Parkinson’s

Divya Yadav

Molecular Neurosciences and Functional Genomics Laboratory,  
Dept. of Biotechnology  
Delhi Technological University  
New Delhi – 110042, India  
divya\_2k20mscbio06@dtu.ac.in

Pravir Kumar

Molecular Neurosciences and Functional Genomics Laboratory,  
Dept. of Biotechnology  
Delhi Technological University  
New Delhi – 110042, India  
pravirkumar@dtu.ac.in

© 2021 IEEE | DOI: 10.1109/ISCON52037.2021.9702338

**Abstract**—Among several neurodegenerative disorders, Parkinson’s is one of the utmost widespread diseases, occur due to the degeneration of the dopaminergic neurons in the region of midbrain which is identified by motor signs like tremor & bradykinesia; non-motor features (like impaired vision, depression, sleeping disorders). Currently, there is no treatment available that can cure PD. It is possible that ligands of glutamate receptor may decelerate the progression of PD via delaying progressive degeneration of dopamine neuron as glutamate is one of the key regulators of basal ganglia functioning & its level increases in PD patients. In addition, alteration of glutamate receptor during the disease and the anti-parkinsonian process has been reported. Reversal of

Parkinson Disease Association (APDA), around 1 million people in the U.S. are having PD and more than 10 million individuals worldwide are suffering from PD. Still there is no cure available for PD but some drugs are there in the market to treat the symptoms & slow down the progression of disease. Selection of drug must be done after estimating both the short-term benefits and long-term consequences. Another factor while selecting the drug is to consider patient-specific factors, for example level of disability, functional & chronologic age, drug tolerance and expected efficacy [3]. Before the motor symptoms turn up, around 50-70% neurons in SNc are degenerated. Current drugs used by PD patients

The screenshot shows the IEEE Xplore search results page. At the top, there are navigation links for IEEE.org, IEEE Xplore, IEEE SA, IEEE Spectrum, and More Sites. The main header includes 'IEEE Xplore' logo, 'Browse', 'My Settings', 'Help', and 'Institutional Sign In' buttons. A search bar is present with a dropdown menu set to 'All' and a search icon. Below the search bar, there are options for 'Search within Publication' and 'ADVANCED SEARCH'. The main content area displays the conference title 'International Conference on Information Systems and Computer Networks (ISCON)' and provides links for 'Copy Persistent Link', 'Browse Title List', and 'Sign up for Conference Alerts'. A navigation bar shows 'Proceedings', 'All Proceedings', and 'Popular'. The search results section shows the conference details: '2021 5th International Conference on Information Systems and Computer Networks (ISCON)' and '22-23 Oct. 2021'. A search bar within the results area is also visible. The results list shows one entry: 'Parkinson's Disease: An Overview and Role of Glutamate and its Receptors: Glutamate receptor based therapy as an alternate way to treat Parkinson's' by Divya Yadav and Pravir Kumar, published in 2021. The authors' names are highlighted with red boxes. There are options to 'Select All on Page' and 'Sort By: Sequence'. A 'Quick Links' section provides links to 'Abstract', 'HTML', and other document formats. A promotional banner for 'Need Full-Text' access to IEEE Xplore is visible on the right side of the page.

## **ACKNOWLEDGEMENT**

The completion of this study could not have been possible without the expertise of my guide, **Prof. Pravir Kumar**, Head of the Department, Department of Biotechnology, Delhi Technological University, who gave me this opportunity to work under his guidance. It was his constant encouragement that enabled me to complete this work successfully. I would like to express my earnest gratitude to him.

I'm highly thankful to Mr. Rohan Gupta, for providing such an incredible support, guidance and valuable suggestions during my entire work. I'm equally gratified to all the respected lab seniors, Ms. Smita Kumari, Mr. Rahul Tripathi, Mr. Sudhanshu Sharma, Mrs. Dia Advani, and Ms. Mehar Sahu for offering beneficial ideas and support.

I would like to mention a special thanks to staff members and Mr. C.B. Singh and Mr. Jitender Singh for providing helping hand whenever required. At last, I would like to express my gratitude and affection to my family and friends who have supported me through the entire tenure of study.

**DIVYA YADAV**

2K20/MSCBIO/06

## ABSTRACT

Among several neurodegenerative disorders, Parkinson's is one of the utmost widespread diseases, occur due to the degeneration of the dopaminergic neurons in the region of midbrain which is identified by motor signs like tremor & bradykinesia; non-motor features (like impaired vision, depression, sleeping disorders). Currently, there is no treatment available that can cure PD. However, there is an imbalance between signaling pathway in the basal ganglia circuit. The direct pathway is downregulated and mediated via dopamine D1-like receptors. In contrast, the indirect pathway is upregulated and this is mediated via dopamine D2-like receptors. Using dopamine D2-like receptors antagonists, PD symptoms can be ameliorated. Increased histaminergic signaling has been observed in PD.

Herein, we want to analyze the interaction between approved antihistamine drugs with dopamine D2-like receptors. We curated a list of 58 approved antihistamine drugs to analyze their affinity with dopamine D2-like receptors as an inhibitor. Literature indicates a substantial correlation between the antihistamines and dopamine D2-like receptors. Thus, targeting these drugs might regulate the disrupted indirect pathway in the basal ganglia circuit.

Through molecular docking analysis, we identified that bilastine, a highly potent and selective inhibitor of histamine H1 receptor shows high affinity with dopamine D2-like receptors and might inhibit the aberrant signaling cascade of indirect pathway and upregulated histaminergic signaling simultaneously. Further studies are required to determine the action.



## CONTENTS

|   |      |
|---|------|
| <b>Candidate's Declaration</b>                                  | ii   |
| <b>Certificate</b>  | iv   |
| <b>Acknowledgement</b>  | vii  |
| <b>Abstract</b>   | viii |
| <b>Contents</b>   | ix   |
| <b>List of Figures</b>  | xi   |
| <b>List of Tables</b>   | xii  |
| <b>List of Abbreviations</b>                                    | xiii |
| <b>CHAPTER 1 INTRODUCTION</b>                                   | 1-3  |
| 1.1 Background of Study   | 1    |
| 1.2 Problem Statement   | 3    |
| 1.3 Objective of Study  | 3    |
| <b>CHAPTER 2 LITERATURE REVIEW</b>                              | 4-19 |
| 2.1 Parkinson's Disease   | 4    |
| 2.1.1 Symptoms and Diagnosis of PD                              | 6    |
| 2.1.2 Current treatment of PD                                   | 7    |
| 2.2 Basal ganglia structure in the brain                        | 9    |
| 2.3 Aberrant neurotransmission in PD                            | 10   |
| 2.3.1 Implication of different neurotransmitter receptors in PD | 14   |
| 2.3.2 Dopamine receptors  | 16   |

|   |       |
|---|-------|
| 2.3.3 Histamine receptors   | 18    |
| 2.4 Therapeutic effects of dopamine and histamine receptors in PD | 18    |
| <b>CHAPTER 3 MATERIAL AND METHODS</b>                             | 20-23 |
| 3.1 Material used   | 20    |
| 3.2 Workflow  | 20    |
| 3.3 Methods for prediction of potential compounds                 | 21    |
| <b>CHAPTER 4 RESULT AND DISCUSSION</b>                            | 24-30 |
| 4.1 Sequence similarity analysis                                  | 24    |
| 4.2 Structures of target receptors                                | 26    |
| 4.3 Dopamine D2-like receptors antagonists                        | 27    |
| 4.4 BBB permeability analysis                                     | 28    |
| 4.5 Analysis of receptor-ligand interaction                       | 28    |
| 4.6 Drug-likeness of the selected ligand                          | 30    |
| <b>CHAPTER 5 CONCLUSION AND FUTURE PERSPECTIVE</b>                | 31    |
| <b>References</b>   | 32-44 |
| <b>LIST OF PUBLICATION</b>  | 45    |

## LIST OF FIGURES

| <b>Fig. No.</b> | <b>Figure Caption</b>  | <b>Page No.</b> |
|-----------------|--|-----------------|
| Figure 1.1      | Alternation in basal ganglia circuit and development of Parkinson's Disease                              | 2               |
| Figure 2.1      | Role of different neurotransmitter receptors in Parkinson's Disease                                      | 5               |
| Figure 2.2      | Depiction of dopaminergic signaling  | 11              |
| Figure 2.3      | Aberrant neurotransmission contributing to the pathophysiology of PD                                     | 13              |
| Figure 2.4      | Different types of neurotransmitters associated with the pathophysiology of PD                           | 15              |
| Figure 3.1      | Flowchart of protocol followed   | 22              |
| Figure 4.1      | Dopamine D2-like receptor (D2, D3, and D4) sequences aligned with histamine receptors (H1, H2, H3 and H4 | 24-25           |
| Figure 4.2      | The 3-D structure of Dopamine D2-like receptors were obtained from PDB                                   | 26-27           |
| Figure 4.3      | Interaction between bilastine and Dopamine D2-like receptors   | 29              |

## LIST OF TABLES

| <b>S. No.</b> | <b>Table Caption</b>  | <b>Page No.</b> |
|---------------|---|-----------------|
| Table I       | Existing drugs used to treat PD symptoms and associated adverse effects | 8-9             |
| Table II      | Types of dopamine receptors   | 16-17           |
| Table III     | Binding energy of known Dopamine D2-like receptor antagonists           | 27              |
| Table IV      | Binding energy (in kcal/mol) of selected ligands                        | 28              |
| Table V       | Physiochemical properties of selected ligands                           | 30              |

## LIST OF ABBREVIATIONS

|               |  |
|---------------|--|
| <b>PD</b>     | Parkinson's Disease  |
| <b>PET</b>    | Positron Emission Tomography   |
| <b>LHb</b>    | Lateral Habenula   |
| <b>APDA</b>   | American Parkinson Disease Association                                 |
| <b>LID</b>    | Levodopa-Induced Dyskinesia  |
| <b>GABA</b>   | Gamma-Aminobutyric Acid  |
| <b>AMPA</b>   | $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole Propionic Acid Receptor |
| <b>SPECT</b>  | Single Photon Emission Computed Tomography                             |
| <b>FDA</b>    | Food and Drug Administration   |
| <b>SNc</b>    | Substantia Nigra Pars Compacta   |
| <b>MAO-B</b>  | Monoamine Oxidase B  |
| <b>L-Dopa</b> | Levodopa   |
| <b>RLS</b>    | Restless Legs Syndrome   |
| <b>Str</b>    | Striatum   |
| <b>BDNF</b>   | Brain-derived Neurotrophic Factor                                      |
| <b>GPe</b>    | Globus Pallidus Externa  |
| <b>MRI</b>    | Magnetic Resonance Imaging   |
| <b>LGP</b>    | Lateral Globus Pallidus  |
| <b>CREB</b>   | cAMP-Response Element Binding Protein                                  |
| <b>NMDAR</b>  | N-methyl-D-aspartate Receptor  |
| <b>CT</b>     | Computed Tomography  |
| <b>ERK</b>    | Extracellular Signal-Regulated Kinase                                  |
| <b>Gpi</b>    | Internal Globus Pallidus   |
| <b>6-OHDA</b> | 6-hydroxydopamine  |

|                                |  |
|--------------------------------|--|
| <b>CNS</b>                     | Central Nervous System                               |
| <b>LDAEP</b>                   | Loudness Dependence of the Auditory Evoked Potential |
| <b>5-HT</b>                    | Serotonin  |
| <b>CINs</b>                    | Cholinergic Interneurons                             |
| <b>cAMP</b>                    | Cyclic Adenosine Monophosphate                       |
| <b>PDD</b>                     | Parkinson's Disease Dementia                         |
| <b>HA</b>                      | Histamine  |
| <b>PI3K</b>                    | Phosphoinositide 3-Kinase                            |
| <b>MAPK</b>                    | Mitogen-Activated Protein Kinase                     |
| <b>A<math>\beta</math></b>     | Amyloid beta   |
| <b>eEF2</b>                    | Eukaryotic Elongation Factor 2                       |
| <b>nAChR</b>                   | Alpha-5 Nicotinic Acetylcholine Receptor             |
| <b>TNF-<math>\alpha</math></b> | Tumour Necrosis Factor Alpha                         |
| <b>COMT</b>                    | Catechol-O-Methyltransferase                         |
| <b>MDD</b>                     | Major Depressive Disorder                            |
| <b>SNr</b>                     | Substantia Nigra Pars Reticulata                     |
| <b>PDB</b>                     | Protein Data Bank                                    |
| <b>AD</b>                      | Alzheimer's Disease                                  |
| <b>HD</b>                      | Huntington's Disease                                 |
| <b>STN</b>                     | Subthalamic Nucleus                                  |
| <b>ALS</b>                     | Amyotrophic Lateral Sclerosis                        |

# CHAPTER – 1

## INTRODUCTION

### 1.1 BACKGROUND

Among several neurodegenerative disorders, Parkinson's is one of the utmost widespread diseases, with a high prevalence rate in the male gender. PD is a neurological disorder with early prominent death of pigmented dopaminergic neurons in the substantia nigra pars compacta (SNc), region of the midbrain which leads to dopamine deficit along with accumulation of intraneural inclusion, known as Lewy bodies that are composed of a protein called  $\alpha$ -synuclein aggregates [1], [2]. And the movement disorder is caused by mainly due to the deficiency of dopamine within the basal ganglia region [2] as shown in **Fig. 1.1** [3]. Furthermore, the striatum collects synaptic input from all the cortical regions, whereas the excitatory glutamatergic signal is regulated by the thalamic region [4]. And in the striatal region, the efficacy of synaptic transmission is regulated by different neurotransmitter systems, including glutamatergic, GABAergic, cholinergic, serotonergic, noradrenergic [5]–[7].

For instance, PD symptoms can be ameliorated upon administration of anticholinergic and anti-histamine drugs, and this indicates that apart from the dopaminergic deficit, other neurotransmitter systems are also affected [8], [9]. The pre-synaptic D4 receptors in the lateral habenula (LHb) exhibit a vital role in the modulation of PD-associated depression [10], agonist A412997 and antagonist L741742 of D4 receptors were injected in SNc sham-lesioned and SNc lesioned rats with different doses, and it was determined that at high concentration firing rate of LHb neurons and neurotransmission emittance are changed in both animal models whereas at low concentration changes occurred in the firing rate and release of neurotransmitters (Dopamine, Glutamate, and GABA in the LHb) through the GABAergic rostromedial tegmental nucleus only in the SNc sham-lesioned.

In PD, increased histaminergic innervation is present at the superior colliculus of SN with altered morphology and release of HA has many effects, such as activation of microglia and astrocytes [11]. During the evolution, it has been observed that the central HA system is well conserved [12]. Histaminergic neurons are present in the hypothalamus at the tuberomammillary nucleus, and they are involved in various functions such as memory

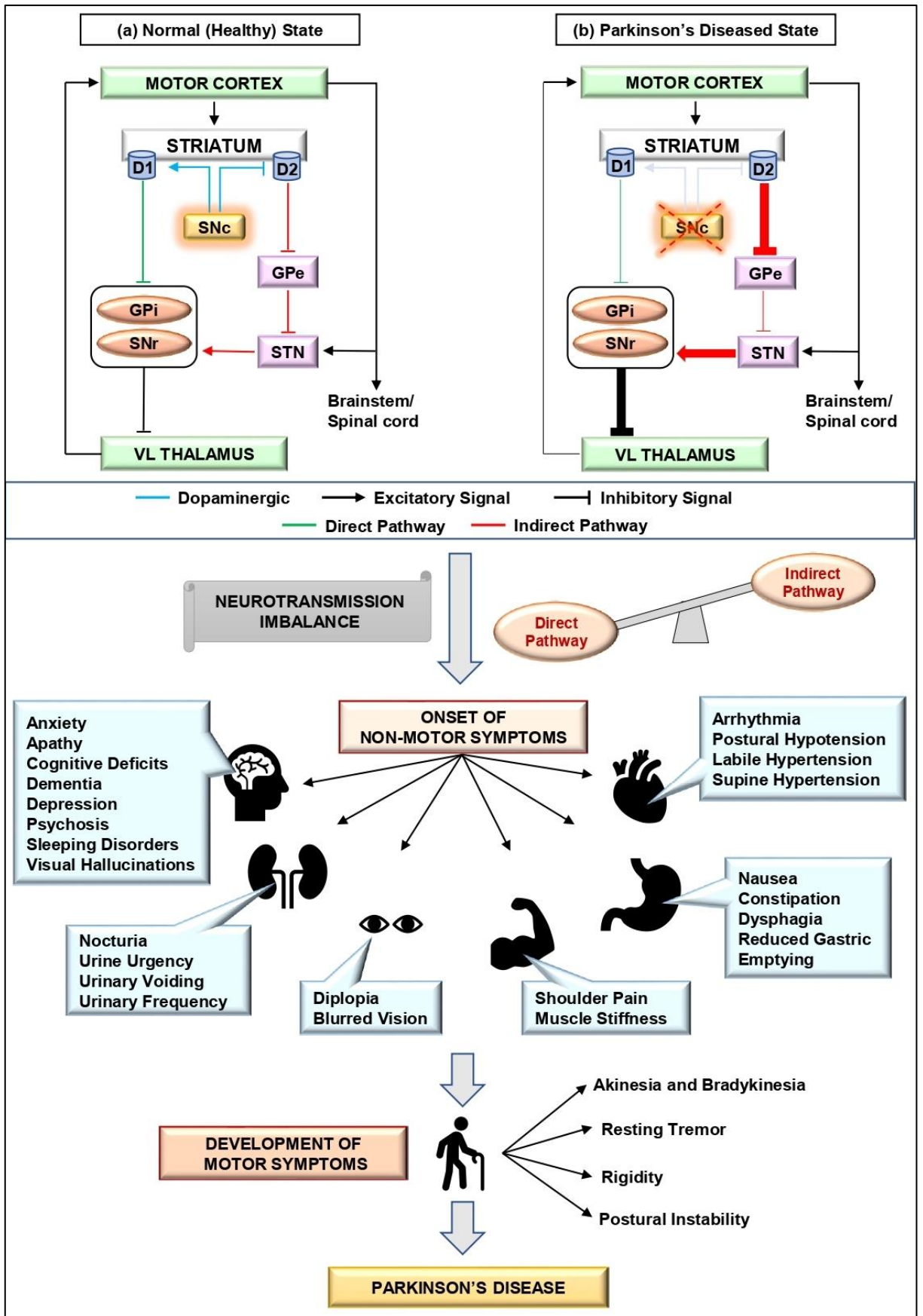


Figure 1.1: Alteration in basal ganglia circuit and development of Parkinson's Disease.



process, cognition and control of the sleep-wake cycle [13]. Thus, antihistamine might play dual function by targeting indirect pathway and upregulated histaminergic signaling in PD.

## **1.2 PROBLEM STATEMENT**

Around 6.1 million people had diagnosed with PD in 2016, among which 47.5% were women and 52.5% were men & according to American Parkinson Disease Association (APDA), around 1 million people in the U.S. are having PD and more than 10 million individuals worldwide are suffering from PD. Still there is no cure available for PD but some drugs are there in the market to treat the symptoms & slow down the progression of disease.

Before the motor symptoms turn up, around 50-70% neurons in SNc are degenerated. In order to reduce the adverse effects, researchers can work on either of two approaches. Modification of currently used levodopa drugs with better efficacy and minimal side effects or to look for an alternative medication which would have good results on patients with least possible adverse effects.

Thus, the treatment with the ability to reinstate conventional basal ganglia functions might act as an analeptic intervention by reverting the motor symptoms that arise due to the adverse effects of current therapies used for PD. Also, traditional treatment neglects non-motor symptoms in PD as it is mainly implemented to improve motor dysfunction and therefore, novel pharmacological targets are required to treat both motor & non-motor signs and to enhance the current analeptic strategies used to treat PD.

## **1.3 OBJECTIVE OF STUDY**

- To identify antagonists of dopamine D2-like receptors via drug repurposing approach.
- To screen approved antihistamine drugs as potential inhibitor of dopamine D2-like receptors.
- To compare the screening result and identify if repurposing approved drugs can be used instead of current medication.

## CHAPTER – 2

### LITERATURE REVIEW

#### 2.1 PARKINSON'S DISEASE

Parkinson's Disease (PD) is a well-known, multifactorial [14], second most common neurodegenerative [15] and age-related disorder [16] with a high risk in the male gender [17]. It occurred due to the loss of dopamine-producing neurons in the region of the midbrain [18], [19], which leads to dopamine deficits and alteration in the basal ganglia circuit. The presence of Lewy bodies ( $\alpha$ -synuclein aggregates) is the hallmark of PD [20]. Symptoms of PD are broadly classified into 2 types [21]: (i) Motor symptoms and (ii) Nonmotor symptoms that lead to functional damage to the body. Motor symptoms and signs include bradykinesia, postural instability, tremor, akinesia, rigidity with several other attributes like precision grip impairment, speech problems and gait disturbance, while the non-motor symptoms are comprised of several events including hyposmia, impaired colour vision, anxiety, depression, constipation, early cognitive dysfunction, dementia and sleep disorders which significantly accountable for the disability of patient and also escalate the risk of mortality [22]. It has been reported that the non-motor symptoms are developed before the emergence of motor signs of PD. Thus, they are also known as pre-motor symptoms [23]. They can also be used as biomarkers in the diagnosis of PD [24]. But the foremost thing is to identify the correct pathophysiology of the disease. In the pathophysiology of PD, studies revealed that the changes occur in the striatal dopamine receptors; for example, increased number of dopamine receptors are linked with dyskinesias, psychotic trauma, on-off phenomena, while the decreased number of dopamine receptors is associated with the disability of the patient and loss of response to levodopa [25]. To date, curation of PD is not possible, but some drugs are there in the market to treat the symptoms and slow down the progression of the disease. Among these drugs, levodopa has been considered as the most effective medication and gold therapy for PD as it can reverse disabilities and also improve quality of life by acting as a dopamine replacement agent. However, long-term administration of levodopa results in levodopa-induced dyskinesia (LID) [26], and a recent study [27] has reported that the

early onset of LID is related to the rapid dose increase of dopaminergic drug. This highlights the need for another effective drug to treat PD with the presence of minimal adverse effects. Besides dopamine, other neurotransmitters levels are also found

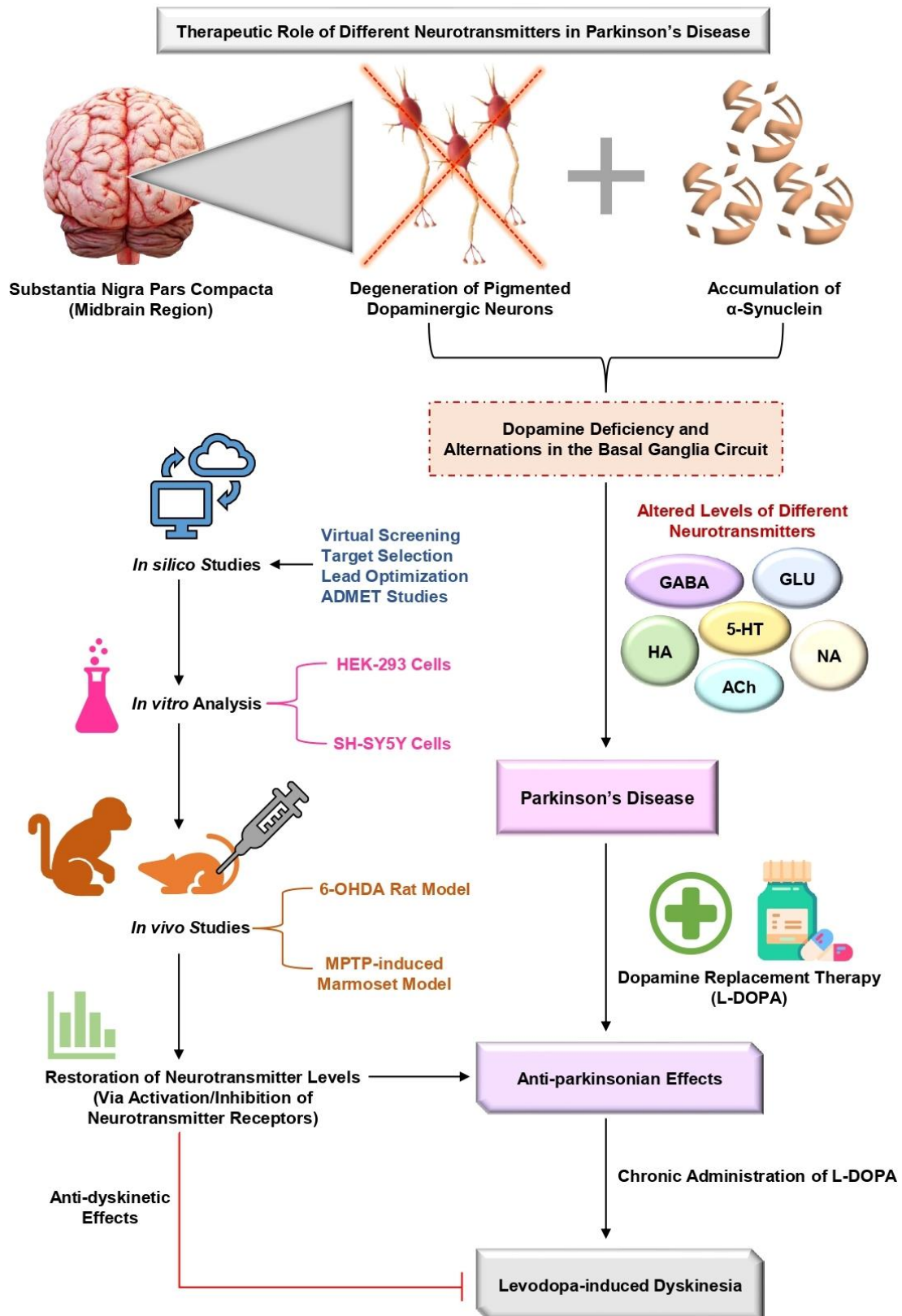


Figure 2.1: Role of different neurotransmitter receptors in Parkinson's Disease

to be altered; thus, these neurotransmitters, together with their different and distinct receptors, can be targeted against PD [3] (**Fig 2.1**). Altered striatal neurotransmitter levels have been observed in PD; for instance, levels of GABA and serotonin are found to be decreased while glutamate, acetylcholine and histamine levels are upregulated [28]. PD involves complex changes in several pathways, which include specific changes in the expression of neurotransmitter receptors [29]. Thus, these non-dopaminergic systems, including glutamatergic, serotonergic, GABAergic, histaminergic, adrenergic and cholinergic pathways within the cortex, brainstem and basal ganglia, are also very crucial for the therapeutic aspect [30]. Few drugs are already accessible in the public domain, and several drugs are currently under clinical trials, which are focused on the non-dopaminergic system. For example, pimavanserin acts on serotonin receptors, and it is used to treat non-motor symptoms like visual hallucinations and psychosis in PD [31], [32]. It has been observed that the non-dopaminergic system can target non-motor symptoms more efficiently as compared to the motor system of PD [33].

### **2.1.1 Symptoms and Diagnosis of PD**

Symptoms are majorly of 2 types: (a) Motor symptoms & (b) Non motor symptoms, that leads to functional impairment. Motor symptoms and signs include bradykinesia, postural instability, tremor, akinesia, rigidity with several other features like precision grip impairment, micrographia, and speech problems and gait disturbance [34]. Non motor symptoms include hyposmia, impaired color vision, anxiety, depression, constipation, early cognitive dysfunction, dementia and sleep disorders which significantly responsible for disability of patient and even increase the risk of mortality [35], [36]. It is also suggested that non-motor symptoms may present even earlier than the emergence of motor signs of PD [37]. Thus, they can be serves as possible biomarker for the detection purpose with higher significance and greater reliability. And also, it will be cost effective method as compared to currently available methods.

Primary step in the diagnosis of PD is based on history and physical examination of the individual but majorly relies on the motor symptoms and initially it requires the presence of bradykinesia along with the minimal existence of any one of the features including: Muscle rigidity, or 4- to 6-Hz rest tremor, or postural fluctuations that are not due to the primary cerebellar, vestibular, visual or proprioceptive dysfunction [38]. Certain signs

and symptoms should not be present, for example neuroleptic therapy at emergence of symptoms, babinski signs, cerebellar signs, initial extreme dementia associated to disruption of memory, language and praxis, existence of a cerebral tumor or communicating hydrocephalus on CT scan report, oculogyric crises, history of repeated head injuries/strokes with stepwise development of parkinsonian features [39]. Due to lack of accuracy in other diagnostic method functional imaging is required for authentication of the clinical diagnostic method and to identify the underlying cause of the disease. Imaging technique for PD includes metabolic positron emission tomography (PET), dopaminergic PET/ SPECT diffusion-weighted MRI, hybrid PET–MRI, and amyloid- $\beta$  PET [40]. Among in these imaging techniques discrimination of PD from atypical parkinsonism (which is a frequent diagnostic issue in the clinic) such as multiple system atrophy or the progressive supranuclear palsy can be done accurately by using structural/ diffusion-weighted MRI & metabolic PET while to look into dementia which commonly complicates the degree of PD, some useful and efficient tools are amyloid- $\beta$  PET, metabolic PET, and cholinergic PET [41].

### 2.1.2 Current treatment of PD

Existing drugs used by PD patients majorly includes [42]:

- a) Dopamine agonists,
- b) MAO-B (Monoamine oxidase-B) inhibitors,
- c) Levodopa (also called L-Dopa),
- d) COMT (Catechol-O-methyl transferase) inhibitors, &
- e) Some other drugs like anticholinergic drugs.

Among these drugs (**Table I**), levodopa has been considered as the most effective medication for PD as it could reverse the disabilities and also improves quality of life by acting as dopamine replacement agent [43]. Despite the effectiveness of levodopa, it also has several adverse effects which are associated with the emergence of erratic absorption & fluctuating plasma level and prolonged levodopa therapy results in chronic treatment complications for example levodopa-induced dyskinesia (LID) & motor fluctuations [44]. Recent modification for enhanced levodopa effect includes formulation for inhalation uptake through the lungs, gastric retentive formulations, and also levodopa prodrug associated to nutritional molecules, solubilized levodopa for continuous subcutaneous infusion, extended-release microspheres controlling drug release, all these products are

currently under development [44].

**Table I: Existing drugs used to treat PD symptoms and associated adverse effects**

| Drug Class                                     | Generic Name (Brand Name)                                    | Adverse Effects  | References |
|--|--|--|------------|
| <b>1. Levodopa</b>                             | Carbidopa-levodopa (Sinemet)<br>[Immediate-release drug]     | Nausea, vomiting, postural hypotension, dyskinesias  | [45]       |
|  | Carbidopa-levodopa (Parcopa)<br>[Orally disintegrating drug] | Low blood pressure, nausea, confusion, dyskinesia  | [46]       |
|  | Carbidopa-levodopa (Duopa)<br>[Enteral suspension]           | Headache, dizziness, nausea, difficulty in sleeping, vomiting  | [47]       |
|  | Carbidopa-levodopa (Rytary)<br>[Extended-release drug]       | Dyskinesia, insomnia, headache, sweating, salivation   | [48]       |
| <b>2. MAO-B Inhibitors</b>                     | Selegiline (l-deprenyl, Eldepryl)                            | Hallucinations, cardiac related adverse effect, postural hypotension, nausea, dizziness, insomnia, confusion   | [49]       |
|  | Safinamide (Xadago)  | Nausea, dizziness, somnolence, headache, back pain, transient mild dyskinesia  | [50]       |
|  | Rasagiline (Azilect®)  | Hypertension, postural hypotension, headache, dizziness, nausea, vomiting, joint pain, hallucinations, sleeping issues, anxiety, rashes, impaired liver function, weight increases/decreases   | [51]       |
| <b>3. Dopamine Agonists (a) Ergoline class</b> | Bromocriptine (Parlodel, Cycloset)                           | Constipation, nausea, vomiting, asthenia, dizziness, headache, rhinitis  | [52]       |
|  | Cabergoline (Caberlin, Dostinex, Cabaser)                    | Constipation, nausea, dizziness, headache, fatigue   | [53]       |
|  | Pergolide (Permax)   | Constipation, diarrhea, sedation, postural hypotension, nausea, dizziness, arrhythmia, dyspnea, hallucinations, confusion, psychosis, visual disorders   | [54]       |
|  | Lisuride (Dopergin, Proclacam, Revanil)                      | Nausea, orthostatic hypotension, fatigue, dizziness, dyskinesia, headache, vertigo, peripheral edema, Mitchell's disease, sweating, dyspnea  | [53]       |
| <b>(b) Non-ergoline class</b>                  | Pramipexole (Mirapex)  | Dyskinesia, postural hypotension, constipation, hallucinations, nausea, asthenia, sleeping issues, dizziness, extrapyramidal movement, headache, edema of lower extremities, confusion in mind | [55]       |
|  | Apomorphine hydrochloride (KYNMOBI™)                         | Reactions at the site of injection, contusion, discharge through nose, nausea, vomiting, confusion, dizziness, peripheral edema, dyskinesia, somnolence,                                       | [56]       |

|                                 |   |   |      |
|---------------------------------|---|---|------|
|                                 |   | hallucinations  |      |
|                                 | Rotigotine Transdermal System (Neupro®)           | Dyskinesia, reaction at the site of application, postural hypotension, vomiting, headache, difficulty in sleep, drowsiness, tiredness, edema of lower extremities, diaphoresis, nausea, dizziness | [57] |
|                                 | Ropinirole (Requip®)                              | Dyskinesia, excess sleepiness, edema of lower extremities, constipation, fatigue, vomiting, hypotension, dizziness, postural hypotension, nausea, impulse control disorders                       | [58] |
|                                 | Piribedil (Clarium, Pronoran, Trastal, Trivastal) | Vomiting, dizziness, hypotension, postural hypotension, syncope, confusion, nausea, agitation   | [59] |
| <b>4. COMT Inhibitors</b>       | Opicapone (Ongentys®)                             | Dyskinesia, constipation, insomnia, urinary tract infection, dry mouth, dizziness, somnolence, weight loss, hallucinations, creatine phosphokinase level increases                                | [60] |
|                                 | Entacapone (Comtan®)                              | Dyskinesia, diarrhea, nausea, urine discoloration   | [61] |
|                                 | Tolcapone (Tasmar®)                               | Dyskinesia, nausea, vomiting, anorexia, insomnia, orthostatic symptoms, hallucination, urine discoloration  | [62] |
| <b>5. Anticholinergic Drugs</b> | Benztropine (Cogentin®)                           | Dry mouth, confusion, blurred vision, memory dysfunction, light headedness, orthostasis, dysuria  | [63] |
|                                 | Trihexyphenidyl HCL (formerly Artane®)            | Dry mouth, constipation, dizziness, vertigo, headache, confusion, blurred vision, nausea, anxiety, nervousness  | [64] |
| <b>6. Adenosine antagonists</b> | Istradefylline (NOURIANZ™)                        | Dyskinesia, nausea, constipation, hallucination, insomnia, somnolence   | [65] |
| <b>7. Others</b>                | Zonisamide  | Decreased appetite, dyskinesia, nasopharyngitis, somnolence   | [66] |
|                                 | Amantadine  | Difficulty in sleeping, dizziness and lightheadedness, hallucinations, dry mouth, nausea, orthostatic hypotension, constipation, peripheral edema   | [67] |

## 2.2 Basal ganglia structure in the brain

The basal ganglia depict a multifarious circuit and its foremost function is to process sensory-motor information accurately [68]. The basal ganglia circuit is an imperative part of the management of behavior such as locomotion, reward learning, eye movement, etc., and elementary traits of the basal ganglia are conserved throughout vertebrate phylogeny [69]. In the basal ganglia functional organization, Striatum (Str) the major input nucleus of the circuit which collects projections from virtually all regions of cortex and then via

the input nuclei information passes to the two regions; MGP & SNr that stands for medial globus pallidus and substantia nigra pars reticulata, which are the output nuclei; then the information finally moves to ventrolateral thalamus which ultimately projects back to the cortex that results in closing of the loop [70]. Signals from Str to the output nuclei can be reach directly as well as indirectly, in case of direct pathway there is a monosynaptic link among the neurons that express the dopamine receptor, D1 and gamma amino butyric acid-ergic (GABAergic) neurons in region of SNr and the globus pallidus internus (Gpi) whereas in the case of indirect pathway which emerges from another group of neurons that express dopamine receptor, D2 that project to region of lateral globus pallidus (LGP) & then moves to Gpi from the region of subthalamic nucleus (STN) as a glutamatergic relay and LGP sends GABAergic projections to the STN which further send it to the MGP & SNr and also send it back to LGP; also, few scientists identified that LGP sends GABAergic projections to the MGP along with STN , schematic depiction of basal ganglia circuit is shown in **Fig. 1a**. The gradual loss of dopaminergic neurons in SNc region and the associated dopaminergic denervation of the Str lead to a cascade of complicated alternations in the regulation of basal ganglia nuclei as shown in **Fig. 1b**; in this process glutamate plays a vital role specifically at two regions of the circuit: Str and STN [71]. Functional re-organization of the basal ganglia circuit develops due to the neurodegeneration of neurons in PD [72]. This ensues in the augmented activity of output nuclei in the basal ganglia, which would be the consequence of intensified glutamatergic neurotransmission from the STN [73]. Studies suggest that in the expansion of PD, escalated activity of an indirect pathway is the foremost contributor [74]–[76].

### **2.3 ABERRANT NEUROTRANSMISSION IN PD**

Targeting and modulation of various neurotransmitters may have the potential to terminate the dopamine deficiency. Dopamine is a catecholamine neurotransmitter, having several functions in the brain like cognition, locomotor activity, food ingestion, endocrine regulation, etc. [77], and the dopaminergic system has been studied with a lot of emphasis in the past five decades because of the association of dysregulation of dopaminergic neurotransmission (dopaminergic signaling cascade is illustrated in **Fig. 2.2**) with various diseases such as PD [78], Schizophrenia [79], Tourette's syndrome [80], thus agonist and antagonist of dopamine receptor play a key role in the treatment of these



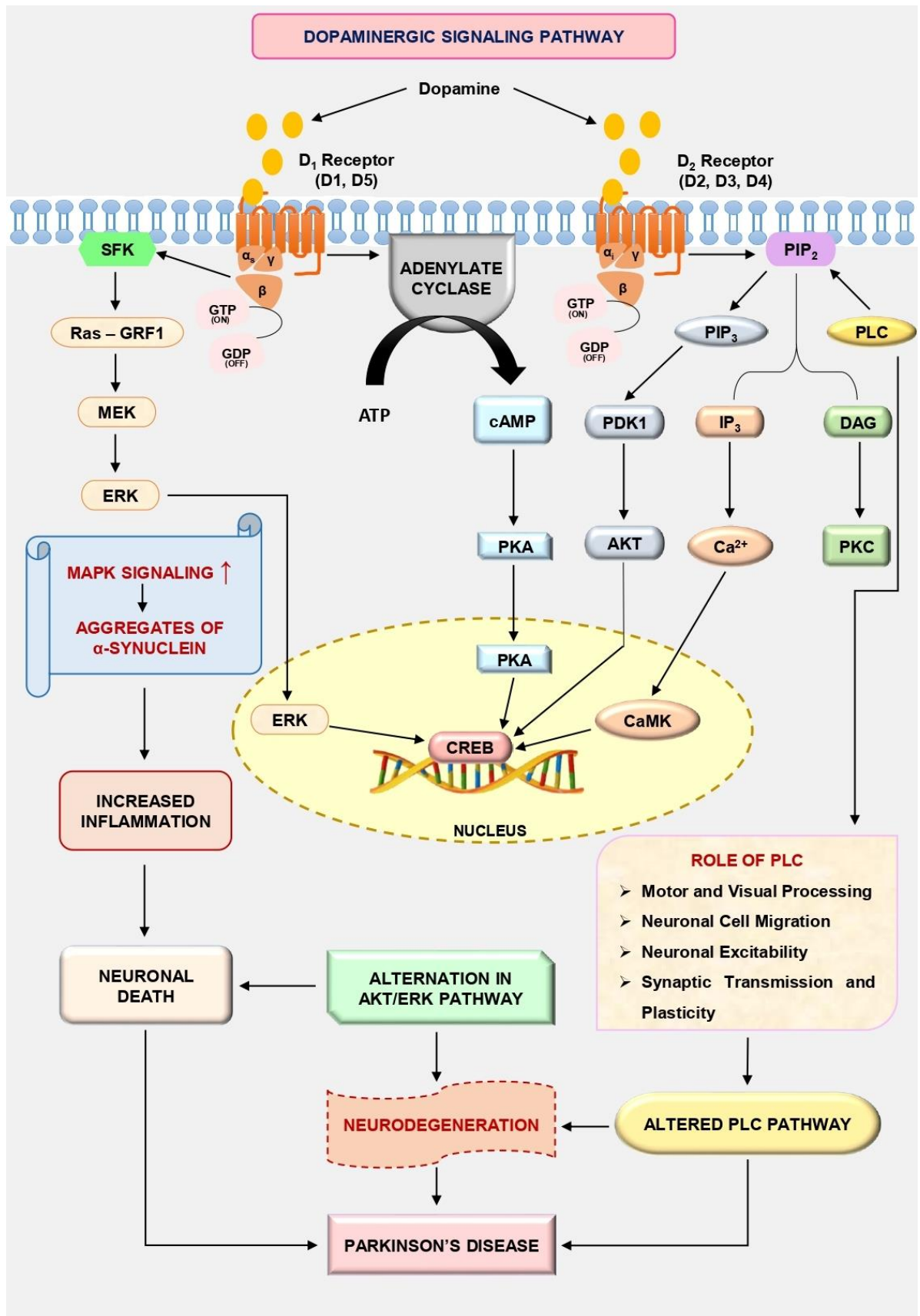


Figure 2.2: Depiction of dopaminergic signaling

diseases, for example, to increase hypokinesia in PD patients, agonists of dopamine receptors [81], [82] are used while antagonists have been developed for the blockade of hallucinations and delusions occurring in the patients of schizophrenia [83]. Degeneration of dopaminergic neurons in PD leads to upregulated glutamatergic neurotransmission, which ultimately contributes to motor dysfunction [84]. 19 distinct receptor binding sites of glutamate were analyzed in Pitx3ak mice mimicking PD condition, among which 11 out of them were found to be altered, including various GABAergic, cholinergic, dopaminergic and serotonin receptors [85]. In 6-OHDA mice, AMPAR levels remained unaffected while NMDAR expression was upregulated in the Str, causing overactivity of glutamatergic neurotransmission [86]. Upregulation of mGluR5 was observed in PD patients when compared with healthy individuals in the striatal region during PET/CT study [87]. Within the basal ganglia region, mGluRs are extensively expressed, and modulation of these receptors is able to regulate various functions such as neuronal excitability [88], [89]. In PD patients, overexpression of ERK-pathway is associated with dyskinesia which might further be associated with aberrant glutamatergic and dopaminergic signaling in the Str [90]. The serotonergic system is involved in various CNS diseases, including PD [91]. Serotonin is a monoamine neurotransmitter that regulates the function of several peripheral organs, and drugs that modulate serotonin receptors are extensively used in psychiatric and neurological disorders [92]. Serotonin is also known as 5-hydroxytryptamine (5-HT). Dysfunction of the 5-HT system performs a vital role in symptoms of PD, both motor and non-motor symptoms like LID, insomnia, depression, etc., and progressive and non-linear degeneration of serotonergic neurons has been observed in PD [93] confirmed by PET and other molecular imaging studies [94]. Central serotonergic function can be evaluated using loudness dependence of the auditory evoked potentials (LDAEP), so this can serve as a potential biomarker in early-onset PD patients to assess serotonergic neurotransmission [95]. It has been shown that altered GABA levels are present in the basal ganglia of PD patients, and these altered levels are associated with axial symptoms (specifically with the degree of gait disturbance); thus, it may be possible that the GABAergic neurotransmission has a significant role in the development of some axial symptoms of PD [96]. In a study including 60 PD patients using functional magnetic resonance imaging (fMRI), it has been observed that the severity of disease and levels of GABA can be correlated inversely in the motor cortex [97]. Alteration of GABAergic transmission has been found with chronic depletion

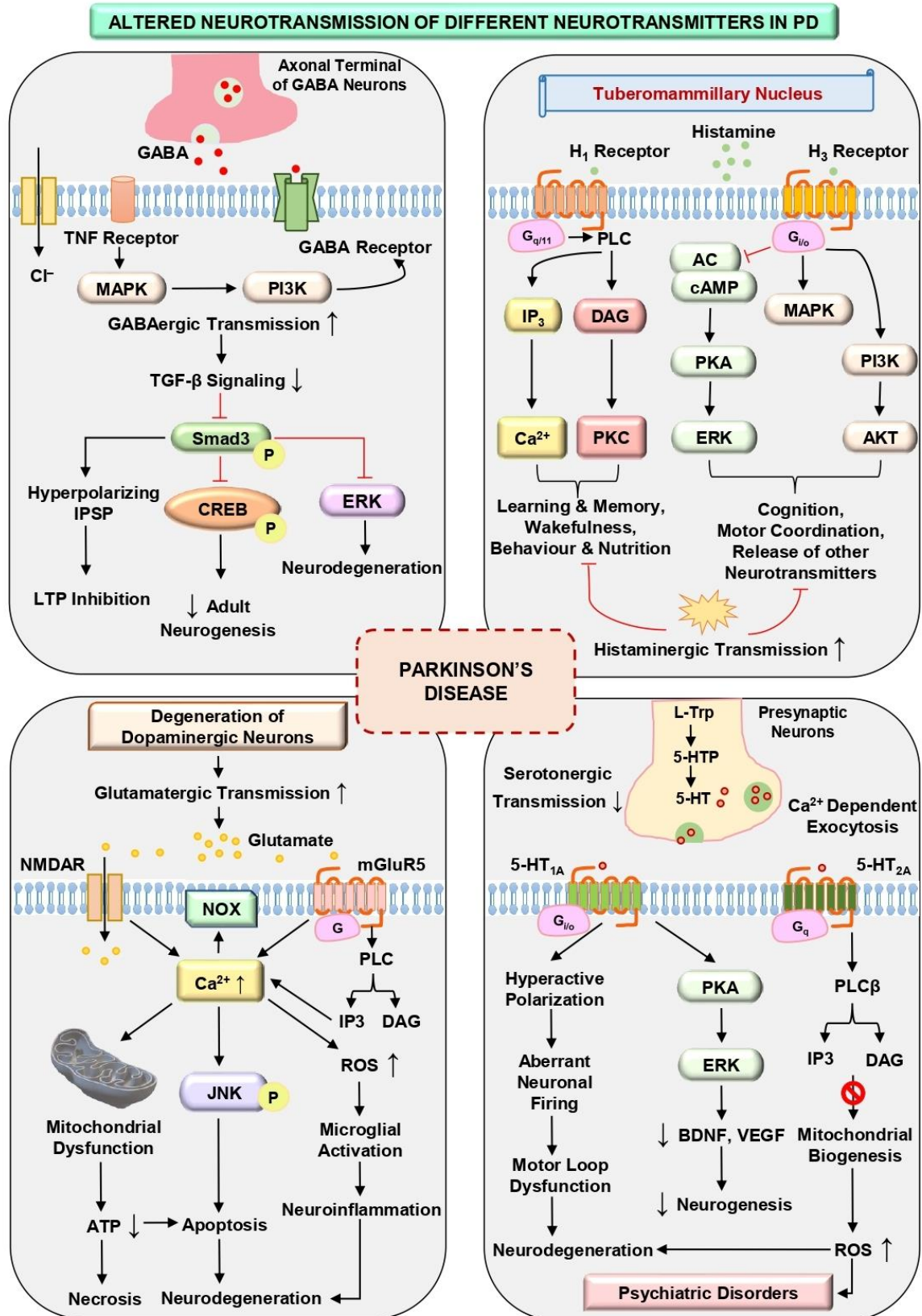


Figure 2.3: Aberrant neurotransmission contributing to the pathophysiology of PD

dopamine in the striatum [98]. Increased choline levels are found in PD [99]. Dopamine and cholinergic system have a perplex anatomical association, and the cholinergic interneurons (CINs) play a key role here in the regulation [100]. Studies suggested that in PDD, because of less A $\beta$  load, cholinergic replacement therapies are efficacious when compared with other neurodegenerative dementias with high A $\beta$  concentration [101]. In an SPECT study, scientists have found that disturbed limbic-paralimbic and prominence cholinergic networks are linked with PDD [102]. They are responsible for controlling locomotion and coordination [103] and can provide a potential therapeutic target against cognitive deficits and dementia in PD [104]. The noradrenergic system is structurally positioned at the nucleus of the brainstem called locus coeruleus [105], the prime source of synthesis of noradrenaline and noradrenergic projections virtually reaches all regions of the brain. The noradrenergic system performs a vital in regulating the synthesis of BDNF as astrocytes express several subtypes of adrenergic receptors [3] (mentioned in **Fig. 2.4**), which have the capability to get activated by noradrenaline & coupled with cAMP production and result in CREB activation via PI3K and MAPK pathway [106]. In rodents, substantial noradrenergic cortical depletion suggests a clear sign of coeruleo-cortical noradrenergic system degeneration, and in PD patients, 50% decline in cortical noradrenaline levels were reported [107]. In PD, increased histaminergic innervation is present at the superior colliculus of SN with altered morphology and release of HA has many effects, such as activation of microglia and astrocytes [11].

### **2.3.1 Implication of different neurotransmitter receptors in PD**

As neurotransmitters play a fundamental role in the functioning of the brain, proteins and neurotransmitter receptors which are associated with the synthesis and activation/inactivation of neurotransmitters, are potent targets for therapeutic drug development with greater impact on the neurological disorder [108]. Neurotransmitters are also essential in controlling the endocrine system. Different types & sub-types of neurotransmitters are illustrated in (**Fig. 2.4**). Neurotransmitter receptors are not only limited to the dendrites; they are also found on the cell bodies as well as on presynaptic terminals. Different neurotransmitter receptors, along with their implications in PD, are summed up in (see **Fig. 2.3**).

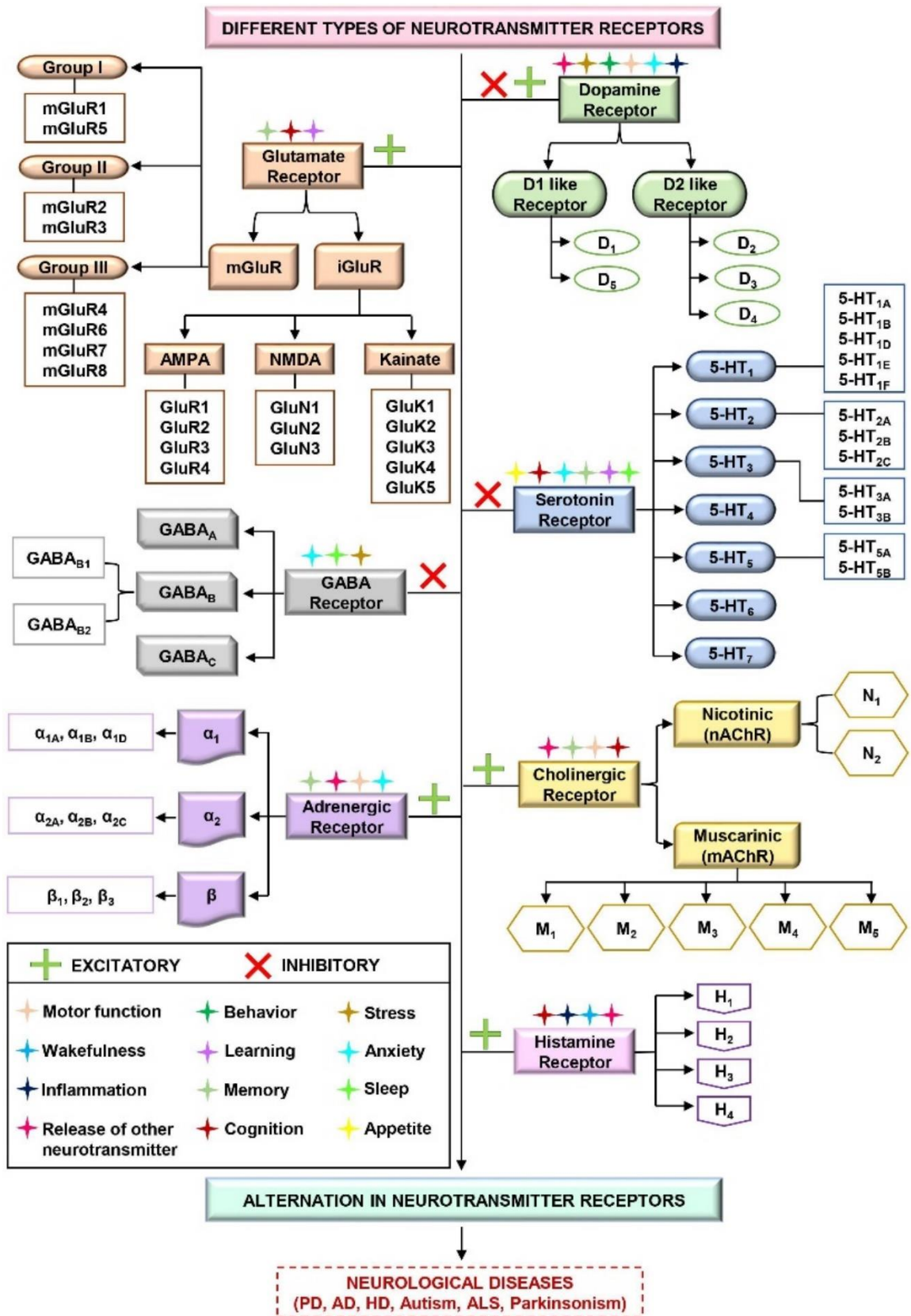


Figure 2.4: Different types of neurotransmitters associated with the pathophysiology of PD

### 2.3.2 Dopamine receptors

Dopamine receptors are mainly of two types [109]: D1 like dopamine receptors (D1, D5) and D2 like dopamine receptors (D2, D3, D4). Different properties of dopamine receptors are mentioned in (Table II) [3]. In neurons, activation of D1 receptor resulted in eEF2 dephosphorylation and this caused an upregulation of the Brain-derived neurotrophic factors (BDNF), and synapsin2b expression, which was accompanied by the least but remarkable elevation in the general protein synthesis and this translation pathway provides a therapeutic target for the treatment of depression and psychiatric disorders [110].

**Table II: Types of dopamine receptors**

| Receptor Class                         | D1  | D5  | D2  | D4  | D5  |
|--|---|---|---|---|---|
| <b>Chromosome location</b>             | 5q 35.1   | 4p 15.1-16.1  | 11q 22-23   | 3q 13.3   | 11p 15.5  |
| <b>Gene name</b>                       | DRD1  | DRD5  | DRD2  | DRD3  | DRD5  |
| <b>Abundance in CNS</b>                | Most abundant   | Less abundant   | Second most abundant  | Third most abundant   | Least abundant  |
| <b>Localization in brain</b>           | Substantia nigra, hypothalamus, amygdala hippocampus, frontal cortex, striatum, caudate nucleus, olfactory tubercle   | Substantia nigra, hypothalamus, pre frontal cortex, putamen, caudate nucleus, nucleus accumbens, amygdala   | Hypothalamus, Putamen, Cerebral cortex, ventral tegmental area, olfactory bulb, striatum  | Islands of Calleja, cortex, striatum, putamen, caudate nucleus, nucleus accumbens   | Hypothalamus, amygdala, frontal cortex, nucleus accumbens   |
| <b>Isoforms</b>                        | -   | -   | 3   | 2   | -   |
| <b>Mass (in KDa)</b>                   | 49.29   | 52.95   | 50.61   | 44.22   | 43.90   |
| <b>Polypeptide length</b>              | 446   | 477   | 443   | 400   | 419   |
| <b>Amino acid in COOH terminal</b>     | 113   | 116   | 16  | 16  | 18  |
| <b>Post-translational modification</b> | Glycosylation, disulfide bond, lipidation   | Glycosylation, disulfide bond, lipidation   | Glycosylation, disulfide bond, lipidation, ubiquitination   | Glycosylation, disulfide bond   | Glycosylation, disulfide bond, lipidation, ubiquitination   |
| <b>Chemical binder</b>                 | Cinnarizine, imipramine, mirtazapine, trimipramine  | Cinnarizine, imipramine, mirtazapine, trimipramine  | Amphetamine, desipramine, imipramine, maprotiline, mirtazapine  | -   | Chlorpromazine  |
| <b>Interactors (Small molecules)</b>   | Acepromazine, amoxapine, bromocriptine, cinnarizine, clozapine, ergotamine, iloperidone, imipramine, loxapine, methotrimeprazine, pipotiazine, pramipexole, propericiazine, quetiapine, ropinirole, rotigotine, thioproperazine, thioridazine, trimipramine, ziprasidone, ergoloid mesylate, dopamine, cabergoline, methylergometrine, olanzapine, apomorphine, | Apomorphine, aripiprazole, dopamine, zuclopenthixol, bromocriptine, chlorpromazine, cinnarizine, cabergoline, fenoldopam, olanzapine, carphenazine, levodopa, mirtazapine, ergotamine, imipramine, lisuride, methotrimeprazine, pergolide, pramipexole, quetiapine, ropinirole, rotigotine, trimipramine, ziprasidone | Olanzapine, aripiprazole, apomorphine, bromocriptine, cabergoline, chlorpromazine, haloperidol, pergolide, pimozide, flupentixol, levodopa, loxapine, metoclopramide, paliperidone pipotiazine, prochlorperazine, risperidone, thiothixene, acepromazine, clozapine, perphenazine, ropinirole, thioproperazine, trifluoperazine, ziprasidone, acetophenazine, buspirone, domperidone, fluspirilene, molindone, quetiapine, sertindole, thioridazine, alizapride, amantadine, carphenazine, chlorprothixene, droperidol, | Aripiprazole, bromocriptine, cariprazine, olanzapine, haloperidol, amisulpride, asenapine, cabergoline, chlorprothixene, levodopa, pramipexole, pergolide, ropinirole, paliperidone, risperidone, sulpiride, ziprasidone, amoxapine, quetiapine, apomorphine, as-8112, captodiame, chlorpromazine, clozapine, | Apomorphine, ropinirole, clozapine, chlorpromazine, dopamine, iloperidone, lisuride, methotrimeprazine, pergolide, quetiapine, remoxipride, rotigotine, asenapine, flibanserin, levodopa, aripiprazole, cabergoline, olanzapine, paliperidone, pramipexole, promazine, propiomazine, risperidone, thiethylperazine, ziprasidone, amoxapine, bromocriptine |

|   |  |   |  |   |   |
|---|--|---|--|---|---|
|   | flupentixol, fluphenazine, carphenazine, chlorprothixene, fenoldopam, levodopa, lisuride, mirtazapine, pergolide, acetophenazine, aripiprazole, asenapine, chlorpromazine, haloperidol, minaprine, paliperidone, perphenazine, promazine, propiomazine, risperidone, thiethylperazine, thiothixene, triflupromazine, zuclopenthixol, |   | fluphenazine, zuclopenthixol, amisulpride, asenapine, mesoridazine, methotrimeprazine, mirtazapine, pramipexole, remoxipride, sulpiride, thiethylperazine, amoxapine, bifeprunox, brexpiprazole, cariprazine, cinnarizine, dopamine, doxepin, ergoloid mesylate, ergotamine, iloperidone, itopride, ketamine, lisuride, minaprine, promazine, propiomazine, triflupromazine, amphetamine, aniracetam, as-8112, bicifadine, brasofensine, bromopride, desipramine, imipramine, lurasidone, maprotiline, memantine, mianserin, nortriptyline, promethazine, rolicyclidine, rotigotine, trimipramine, yohimbine | domperidone, dopamine, iloperidone, lisuride, methotrimeprazine, pimoziide, quetiapine, remoxipride, rotigotine, yohimbine                    |   |
| <b>Accession no. (UniprotKB)</b>                | P21728   | P21918  | P14416   | P35462  | P21917  |
| <b>Type</b>                                     | Stimulatory receptor (Gs-coupled)  | Stimulatory receptor (Gs-coupled)   | Inhibitory receptor (Gi-coupled)   | Inhibitory receptor (Gi-coupled)  | Inhibitory receptor (Gi-coupled)  |
| <b>Function</b>                                 | Memory, locomotion, renal control, impulse control, sleep  | Vasodilatation, stimulation of renin secretion, cognition, attention, decision making, motor learning | Sleep, sexual activity, attention, memory & learning   | Cognition, regulation of food ingestion, impulse control, sleep, attentiveness  | Attention, cognition, sleep, sexual activity, impulse control   |
| <b>Mechanism</b>                                | Via activating adenylyl cyclase activity   | Via activating adenylyl cyclase activity  | Via inhibiting of adenylyl cyclase activity  | Via inhibiting of adenylyl cyclase activity   | Via inhibiting of adenylyl cyclase activity   |
| <b>Signaling affected</b>                       | PLC signaling, PKA and PKC pathways  | PLC signaling, PKA and PKC pathways   | PLC signaling, PKA, PKC pathways, $\beta$ -arrestin pathway, EGFR, GIRK signaling pathway  | PLC signaling, PKA, PKC pathways, $\beta$ -arrestin pathway, EGFR, GIRK signaling pathway   | PLC signaling, PKA, PKC pathways, $\beta$ -arrestin pathway, EGFR, GIRK signaling pathway   |
| <b>Examples of associated neuronal diseases</b> | PD, schizoaffective disorder, psychosis, TS, ALS, unipolar depression, schizophrenia   | PD, schizoaffective disorder, psychosis, depressive disorder, unipolar depression, schizophrenia,     | PD, AD, HD, bipolar disorder, schizoaffective disorder, psychosis, TS, fibromyalgia, hyperprolactinemia, RLS, post-traumatic stress disorder, OCD, autism  | PD, MDD, AD, HD, bipolar disorder, TS, schizoaffective disorder, psychosis, hyperprolactinemia, RLS, schizophrenia, DAN, delirium, acromegaly | PD, MDD, AD, HD bipolar disorder, schizoaffective disorder, psychosis, TS, RLS, treatment resistant depression hyperprolactinemia, schizophrenia, DAN |

In a study, anxiolytic and antidepressant-like effects along with increased neural stem cell proliferation, neuronal differentiation and long-term persistence have been shown by agonists of D1 receptor in rats via positive regulation of a signaling pathway known as the Wnt/ $\beta$ -catenin pathway in the hippocampus [111]. Earlier, one study revealed that D2 receptor on astrocytes suppresses neuroinflammation, and now using the PD mouse model, it has been suggested that agonist of D2 receptor inhibits activation of Nod-like receptor protein 3 (NLRP3) inflammasome via a  $\beta$ -arrestin2-mediated mechanism, as NLRP plays a vital in the pathogenesis of PD thereby D2 receptor agonist might provide novel pharmacological relevance associated with PD [112]. It has been discovered that

D3 receptors also participate in the regulation of neuronal development and contribute to structural plasticity, along with the ability to trigger neuroprotective mechanisms via intracellular pathways [113]. Data has also proven that D3 receptors form active and functional heteromers by executing direct interaction with the other receptors and these heteromers have distinctive pharmacological properties like for instance, nicotinic acetylcholine receptors (nAChR) found on dopaminergic neurons form a heteromer with D3 receptors, which act as a molecular modulator of neurotrophic effects mediated via nicotine thus, it can provide pharmacological targets for PD [113].

### **2.3.3 Histamine receptors**

Histamine receptors have been classified into four types: H1, H2, H3, and H4; are G protein-coupled receptors. Several studies indicate the negative impact of the histaminergic system on the progression of PD [114], as HA levels increase in PD shown in PD animal models, and this could be associated with an accelerated rate of degeneration of pigmented dopaminergic neurons in the SNc [115]. In PD, elevated HA levels might link with neovascularization [114]. Antagonists of HA receptors improves motor and other symptoms in PD condition, thereby highlighting its significance in the clinical response [116]. TNF- $\alpha$  upregulation has been found in several neurodegenerative diseases, including PD, AD and ALS [117]. It has been demonstrated that famotidine, an antagonist of H2, receptor improves motor functioning in primates PD model [118]. But oral administration of famotidine showed no anti-dyskinetic effects in a human study, including 43 PD patients [119], no adverse events were reported. While another study using a different antagonist had some contradictory results. Chronic administration of imzepip, an agonist of H3R, results in reduced LIDs in a rodent model by blocking the release of two neurotransmitters (Glutamate and GABA) and preventing ERK1/2 phosphorylation [120]. In the pathophysiology of PD, activation of microglial cells plays a vital role [121], and its activity is modulated by H4R. An antagonist of H4R, JNJ7777120, blocks the mRNA expression of CD68, IL-1 $\beta$  and TNF- $\alpha$ ; along with this, it has been shown that JNJ7777210 does not have a direct effect on dopaminergic neurons [122].

## **2.4 THERAPEUTIC EFFECT OF DOPAMINE AND HISTAMINE RECEPTORS IN PD.**



A recent study [123], has shown that as compared to non-dyskinetic control mice, dyskinetic mice have a remarkably higher expression level of D5 receptors, upon injection of SCH- 23390 (antagonist of D1 like dopamine receptor), dyskinetic behavior is inhibited after levodopa administration while injection of SKF-83959 (a partial agonist of D5 receptor) has remarkable dyskinetic movements without levodopa indicating a significant role of D5 receptors towards the pathophysiology of PD and associated dyskinetic effects. Upon activation via an agonist, the D3 receptors increase dopamine levels and as well as reduces  $\alpha$ -synuclein aggregation. Activated D3 receptors also enhance secretion of the brain-derived neurotrophic factors (BDNF), thus, increases oxidative stress, improves neuroinflammation, and in addition, age of onset can be predicted by a mutation in the D3 receptors [124]. Yang et al., found that in the striatum, the D1 plus D3 combined level is even better correlated than the D1 and D3 alone to the clinical manifestations of Lewy bodies disease patients [125].

HA is a key regulator of microglial phagocytosis and production of reactive oxygen species (ROS), which are involved in neuronal death of dopaminergic neurons in the SNc as shown in in vivo model, and by using H1 antihistamines, HA-induced microglia activation can be prevented along with increased neuronal survival [126]. Alone antagonists of H1 or H4 are not able to affect the production of TNF- $\alpha$  and IL-1 $\beta$  [127] while using both receptor antagonists together inhibit the HA-induced microglia activation and also blocks the liberation of TNF- $\alpha$  and IL-1 $\beta$  from the brain of rat. In the hemiparkinsonian rodent models of PD, ranitidine which is also an antagonist of the H2 receptor, suppresses LID via regulating the accumulation of Delta FosB protein and ERK pathway, simultaneously preserving the therapeutic effects of DOPA in easing akinesia [128]. Upregulation of H3R has been found in the post-mortem study of PD patients, thereby indicating the possible role of H3R in the treatment of PD, for example, administration of inverse agonist of H3R (BF 2649) or partial agonist of H4R (clobenpropit) with antihistamine monoclonal antibodies remarkably prevent accumulation of  $\alpha$ -synuclein in the striatum [129]. H4R is recognized as a new potential target for various neurodegenerative diseases like PD, ALS, etc.[130]. In a recent study, increased expression of H4R mRNA have been observed in the post-mortem study of PD patients [131]. And this study also demonstrated that JNJ prevents the decrease in dopamine in the striatum region of PD rats and improves TH-positive fiber degeneration along with the reduction in Lewy bodies count [131].

## CHAPTER – 3

### MATERIAL AND METHODS

#### 3.1 MATERIAL USED

**Database used:** Drugbank, PubMed, ChEMBL, Pubchem, UniProt, Protein Data Bank (PDB), BioGRID.

**Software used:** Clustal Omega, SwissDock, DockThor, Swiss PDB Viewer, SwissADME, Biovia Discovery Studio Visualizer.

#### 3.2 WORKFLOW

Dopamine D2-like receptors were selected as the target molecules from the literature survey. The aim was to screen antihistamine drugs against dopamine D2-like receptors to repurpose an antagonist that might target the aberrant indirect dopaminergic and histaminergic signaling in PD. List of FDA approved antihistamine drugs were identified by Drugbank. Further, the BBB (Blood Brain Barrier) permeable drugs were selected as ligands for the study. Flowchart of protocol has been depicted in **Fig. 3.1**.

#### 3.3 METHODS FOR PREDICTION OF POTENTIAL COMPOUNDS

##### 3.3.1 Retrieval of Receptor Sequences for Similarity Analysis

For the sequence similarity analysis, sequences of dopamine receptors, DRD2 (P14416), DRD3 (P35462), DRD4 (P21917) and histamine receptors, HRH1 (P3567), HRH2 (P25021), HRH3 (Q9Y5N1), HRH4 (Q9H3N8) were obtained in fasta file format from UniProt database (<https://www.uniprot.org/uniprot/>) [132]. Multiple sequence alignment was performed with the help of a freely available tool, Clustal Omega [133].

##### 3.3.2 Data Extraction

3-D structure of dopamine receptors, D2 receptor, D3 receptor, D4 receptor were extracted from PDB (<https://www.rcsb.org/>) [134]. A total of 58 FDA approved

antihistamine drugs were mined from Drugbank [135]. Structures of these drugs were collected from Pubchem. Quetiapine, Eticlopride and Clozapine structures were also extracted. These are well-known antagonists of different D2-like receptors identified with the help of BioGRID database (<https://thebiogrid.org/>). These 3 drugs were used as control drugs for the respective receptor. ChEMBL database was used to assess different properties of ligands like molecular weight, a clinical phase of drug, IC<sub>50</sub> value, rotatable bonds, number of hydrogen bond donor/acceptor [136]. And out of these 46 drugs were selected for further molecular docking studies.

### **3.3.3 BBB Permeability Analysis**

BBB permeability analysis were done using online BBB permeability predictor tool (<https://www.cbligand.org/BBB/mainpage.php>).

### **3.3.4 Molecular Docking**

The molecular docking was carried out using SwissDock [137] and DockThor [138] web server to understand the receptor-ligand interaction. The steps mentioned below were performed further:

#### **a. Preparation of the Target Receptors**

The 3-D structures of dopamine receptor 2, 3 and 4 were extracted from RCBS PDB in .pdb file format. Then, this .pdb file was opened in Biovia Discovery Studio Visualizer. The structures were modified and redefined by deleting unwanted molecules of water and heteroatoms. Further, to compensate the loss the structures were opened in Swiss PDB Viewer for energy minimization. Finally, the modified files were saved in .pdb format.

#### **b. Preparation of the Ligand Molecules**

The 3-D structures of ligands were retrieved in .sdf file format. Then, this .sdf files were converted in .mol2 file format using Biovia Discovery Studio Visualizer.

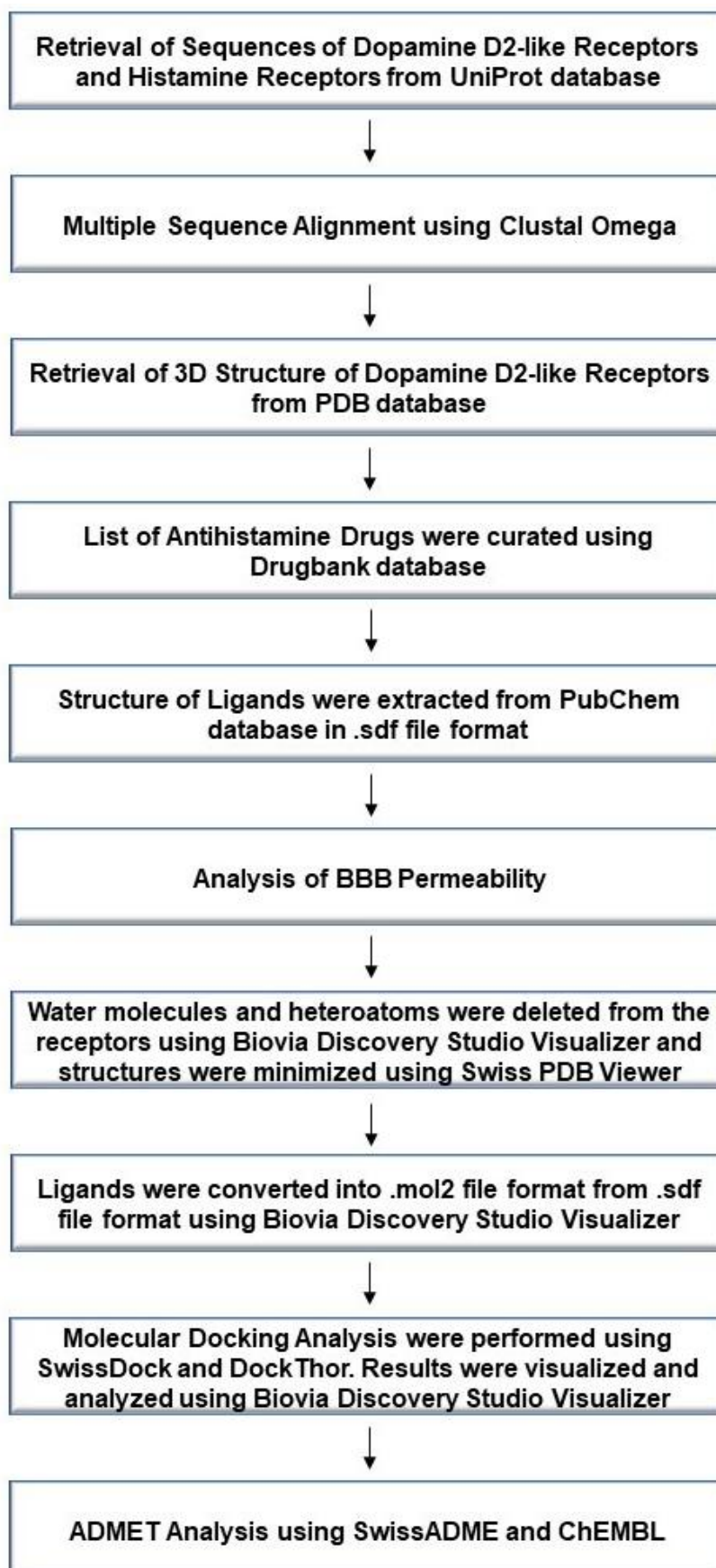


Figure 3.1: Flowchart of protocol followed

### **c. Molecular Docking Studies**

After the successful preparation of the receptors and ligand structures, molecular docking analysis was performed by DockThor and SwissDock web servers. Interactions were visualized and evaluated with the help of Biovia Discovery Studio Visualizer.

#### **3.3.5 ADMET Analysis**

At last, ADMET analysis and pharmacokinetic properties were calculated using SwissADME and ChEMBL. It is a crucial step while identifying the lead molecules as it describes about the pharmacokinetics and drug-likeness of the compounds.

# CHAPTER – 4

## RESULT AND DISCUSSION

### 4.1 SEQUENCE SIMILARITY ANALYSIS

Sequence of dopamine (D2, D3, D4) and histamine receptors (H1, H2, H3, H4) were aligned. Similarities were present between dopamine and histamine receptor sequences. However, variations were observed in the length of the sequences. Regions of common amino acid were highlighted with black rectangles in the **Fig 4.1**. Different symbols indicate the match (\*) and similarity with few mismatch (“:”), (“.”).

Different color indicates different type of amino acid. For instance, blue color represents negatively charged amino acid that are Glutamate (D) and Aspartate (E) while pink color represents positively charged amino acid including Arginine, Lysine and Histidine symbolized with one letter code, R, K, and H respectively. Polar and non-polar amino acid are represented with red and green colors respectively.

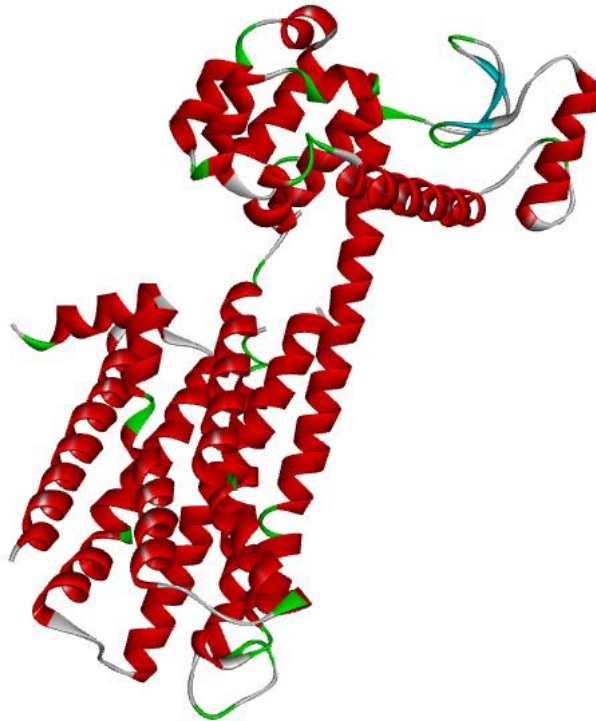


|                            |   |     |
|----------------------------|---|-----|
| HRH3_Histamine_H3_receptor | PA-ILSWEYLSGGSS-IPEGHCYAEFFYNWYFLITASTLEFFTPFLSVTFNLSIYLNIO   | 226 |
| HRH4_Histamine_H4_receptor | PM-ILVSESWKD-----EGSECEPGFFSEWYILAITSFLEFVIVLVAIFNMNIYWSLW    | 202 |
| HRH1_Histamine_H1_receptor | PI--LGWNHFMQQTSVRRREKCEIDFYDVTWFKVMTAIIINFYLPDLLMLWFYAKIYKAVR | 218 |
| HRH2_Histamine_H2_receptor | LSIHLGWNSRNET-SKGNHTTSKCKVQVNEVYGLVDGLVTFYLPDLLIMCITYYRIKVAR  | 210 |
| DRD4_dopamine_receptor     | PV-LCGLNDVRG----R--DPAVCRL-EDRDYVYVSSVCSFFLPCPLMLLLYWATFRGLQ  | 220 |
| DRD2_dopamine_receptor     | PL-LFGLNNA-----DQNECII-ANPAFVYVSSIVSFYVPFIVTLLVYIKIYIVLR      | 217 |
| DRD3_dopamine_receptor     | PL-LFGFNNT-----G--DPTVCSI-SNPDFVIYVSSVVSFYLPFGVTLLVYARIYVVLK  | 216 |
|                            | : . * *   | :   |
| HRH3_Histamine_H3_receptor | RRTRLRLDGAREA-----AGPEPPEAQSPPPPGCWGCWQKGHGEAMPLHRYG---       | 276 |
| HRH4_Histamine_H4_receptor | KRDHLRCQSHP-----GL---TAV---SSNICGHSF-----                     | 228 |
| HRH1_Histamine_H1_receptor | QHCQHRELINRSLPSFSEIKLRPENPKGDAKKP-GKESPEVL-K---RKPKDAGGGSV    | 272 |
| HRH2_Histamine_H2_receptor | DQAKRINH-----   | 219 |
| DRD4_dopamine_receptor     | RWEVARRAK-----LHGRAPRRPSGP--GP-PSPTTPAP-----RLPQDPCGPD        | 262 |
| DRD2_dopamine_receptor     | RRRKRVTNR--SSRAFRAHLRAPLKGNCT--HP-EDMKLCTVIMKSNG-SFPVNRRRVEA  | 272 |
| DRD3_dopamine_receptor     | QRRRKRILTRQNSQCNSVRPQTPQLSP--DP-AHLEL-----                    | 252 |
| HRH3_Histamine_H3_receptor | -----VGEAAVGA-----EAGEATLG                                    | 292 |
| HRH4_Histamine_H4_receptor | -----RGR-----LSSRRSLS   | 239 |
| HRH1_Histamine_H1_receptor | LKSPSQTPKEMKSPVVSQEDDREVDKLYCFPLDIVHMQAAAEGSSRDYVAVNRSHGQLK   | 332 |
| HRH2_Histamine_H2_receptor | -----   | 219 |
| DRD4_dopamine_receptor     | APPAPGLPRG-----PCGPDCAAPSLPQDPCGPDCAAPPAG-----LP              | 302 |
| DRD2_dopamine_receptor     | ARRAQELEMESSTSP-PERTRYSPIPPS-----HHQLTLPDPSHHG-LHSTPDSPAK     | 324 |
| DRD3_dopamine_receptor     | -----KRYYSICQ-----DTALGGPG---FQERGGE-LK                       | 277 |
| HRH3_Histamine_H3_receptor | GGGGGGSVASPTSSSGSSRG---TERPRSLKR-----GSKPSASSAS-LEKRMKM--     | 339 |
| HRH4_Histamine_H4_receptor | -----ASTEVPASF---HSER---QRRKSSLMF-----SSRTKMNSNT-IASKMGSFS    | 280 |
| HRH1_Histamine_H1_receptor | TDEQGLNTHGASEISEDQMLGDSQSFSRTDSDTTTETAPGKGLRSGSNTGLDYIKFTWK   | 392 |
| HRH2_Histamine_H2_receptor | -----   | 219 |
| DRD4_dopamine_receptor     | PDPCGSNCAPPDAV-----RAAALPPQTPPQ-----T-----                    | 329 |
| DRD2_dopamine_receptor     | PEKNGHAKDHPKIA-----KIFEIQTMPNGKTRTSLKT-----                   | 357 |
| DRD3_dopamine_receptor     | REEKTRNSLSPTIA-----PKLSLEVRKLSNGRLSTSLKL-----                 | 312 |
| HRH3_Histamine_H3_receptor | ----VSQSFTQRFRLSRDRKVAKSLAVIVSIFGLCWAPYTLMMIIRAACHG-HCVDPYWY  | 394 |
| HRH4_Histamine_H4_receptor | QSDSVALHQREHVELLRARLAKSLAILLGVFAVCWAPYSLFTIVLSFYSSATGPKSVWY   | 340 |
| HRH1_Histamine_H1_receptor | RLRSHSRQYVSLHMRERKAQQLGFIMAAFILCWIPYFIFFMVIAFCNK-CC-NEHLH     | 450 |
| HRH2_Histamine_H2_receptor | -----SSWKAATIREHKATVTLAAVMGAFIICWFPYFATFVYRGLRGD-DAINEVLE     | 270 |
| DRD4_dopamine_receptor     | -----RRRRRAKITGRERKAMRVLVVVGFLLCWTPFFVHITQALCPA-CSVPPRLV      | 382 |
| DRD2_dopamine_receptor     | -----MSR-RKLSQQKKEKATQMLAIVLGVFIICWLPFFITHILNHCN--CNIPPLY     | 408 |
| DRD3_dopamine_receptor     | -----GPLQPRGVPLREKKATQMAIVLGAFTVCWLPFFLTHVLNTHCQT-CHVSPELY    | 365 |
|                            | : :: : :: * ; * * * :   |     |
| HRH3_Histamine_H3_receptor | ETSFLLWANSVNPVLYPLCHHSFRAFTKLLCPQKLIKIQPHSSLEHCWK-----        | 445 |
| HRH4_Histamine_H4_receptor | RIAFWLQWFNSFVNPLLYPLCHKRFQKAFKIFCIKKQPLPSQHSRSVSS-----        | 390 |
| HRH1_Histamine_H1_receptor | MFTIWLGYINSTLNPLIYPLCNENFKKTFKRILHIRS-----                    | 487 |
| HRH2_Histamine_H2_receptor | AIVLWLGYANSALNPILYAALNRDVRTGYQQLFCCRLANRNSHKTSLSRNASQLSRQSR   | 330 |
| DRD4_dopamine_receptor     | SAVTWLGYVNSALNPVIYTFVNAEFRNVFRKALRACC-----                    | 419 |
| DRD2_dopamine_receptor     | SAFTWLGYVNSAVNPIIYTFNIEFRKAFKILHC-----                        | 443 |
| DRD3_dopamine_receptor     | SATVWLGYVNSALNPVIYTFNIEFRKAFKILSC-----                        | 400 |
|                            | ** : * * : * * : * :  |     |
| HRH3_Histamine_H3_receptor | -----   | 445 |
| HRH4_Histamine_H4_receptor | -----   | 390 |
| HRH1_Histamine_H1_receptor | -----   | 487 |
| HRH2_Histamine_H2_receptor | EPRQQEEKPLKLQVWSGTEVTAPQGATDR                                 | 359 |
| DRD4_dopamine_receptor     | -----   | 419 |
| DRD2_dopamine_receptor     | -----   | 443 |
| DRD3_dopamine_receptor     | -----   | 400 |

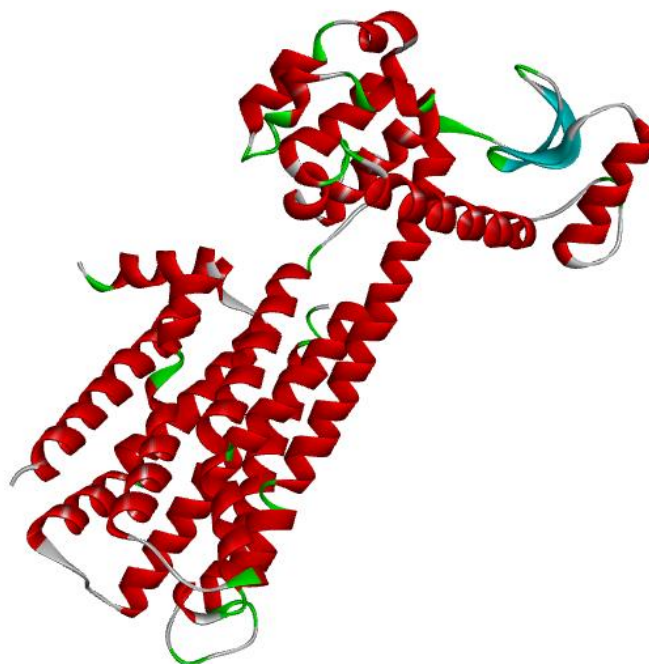
**Figure 4.1: Dopamine D2-like receptor (D2, D3, and D4) sequences aligned with histamine receptors (H1, H2, H3 and H4).**

## 4.2 STRUCTURES OF TARGET RECEPTORS

The energy minimized 3-D structures of dopamine receptors, D2, D3, D4 receptors were showed in **Fig. 4.2.**

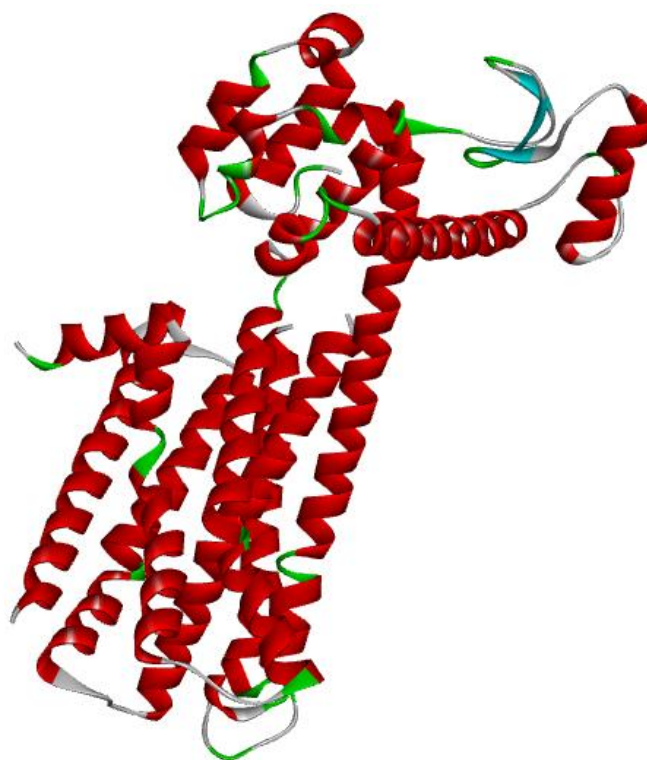


**(A) 3-D structure of dopamine 2 receptor**



**(B) 3-D structure of dopamine 3 receptor**





(C) 3-D structure of dopamine 4 receptor

**Figure 4.2: The 3-D structure of Dopamine D2-like receptors were obtained from PDB. (A) Dopamine 2 receptor, (B) Dopamine 3 receptor, (C) Dopamine 4 receptor. (Water molecules and heteroatoms were removed)**

### 4.3 DOPAMINE D2-LIKE RECEPTORS ANTAGONISTS

With the help of BioGRID database, Quetiapine, Eticlopride and Clozapine were selected as the control drugs for D2, D3, and D4 receptor respectively. Each drug was docked with its respective receptor and docking scores were obtained. These values were used as cut-off scores against antihistamine drugs.

**Table III: Binding energy of known Dopamine D2-like receptor antagonists**

| S.No. | Receptor Name       | Drug Name   | Estimated $\Delta G$ (kcal/mol) |
|-------|---------------------|-------------|---------------------------------|
| 1.    | Dopamine receptor 2 | Quetiapine  | -8.4                            |
| 2.    | Dopamine receptor 3 | Eticlopride | -8.22                           |
| 3.    | Dopamine receptor 4 | Clozapine   | -7.56                           |

#### 4.4 BBB PERMEABILITY ANALYSIS

The list of antihistamine drugs were mined from DrugBank database and their structures were obtained from PubChem. Further, these drugs were subjected to BBB permeability analysis. Out of 58 drugs, 9 were non-BBB permeable.

#### 4.5 ANALYSIS OF RECEPTOR-LIGAND INTERACTION

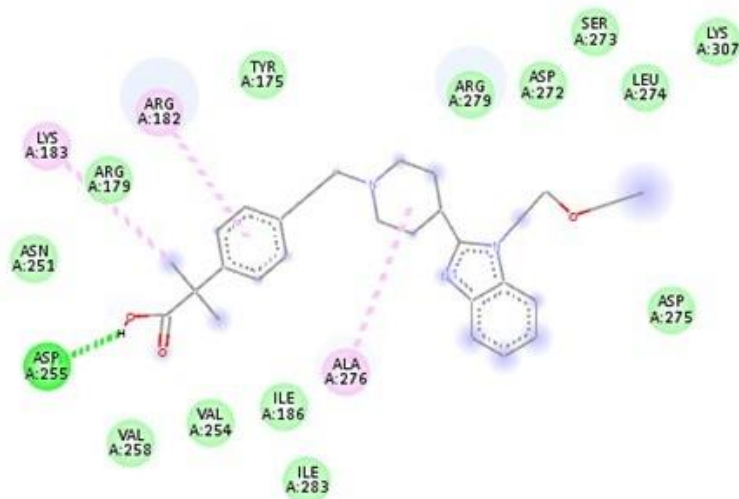
Out of 46 drugs, Bilastine was the only drug that met the required cutoff against all the three dopamine D2-like receptors. 7 compounds met the required cutoff of -7.56 kcal/mol in case of Dopamine D4 receptor whereas 5 compounds met the required cutoff of -8.22 kcal/mol in case of Dopamine D3 receptor. In addition, only 1 compound i.e., Bilastine met the required cutoff of -8.4 kcal/mol in case of Dopamine D2 receptor.

**Table IV: Binding energy ( in kcal/mol) of selected ligands**

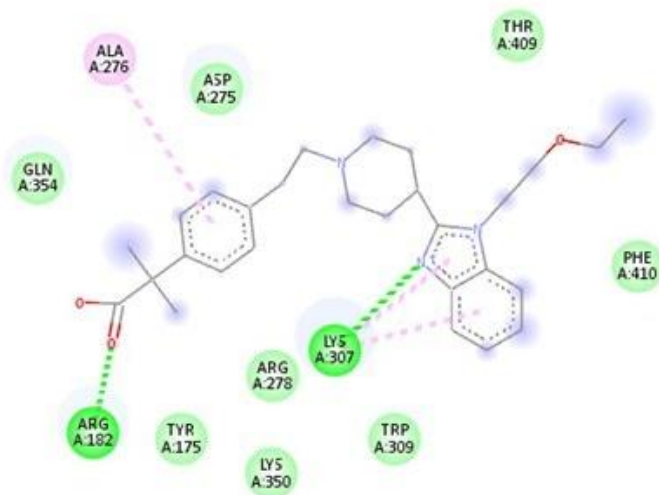
| S. No. | Ligands     | PubChem ID | D2 Receptor  | D3 Receptor  | D4 Receptor  |
|--------|-------------|------------|--------------|--------------|--------------|
| 1.     | Bilastine   | 185460     | <b>-8.84</b> | <b>-8.59</b> | <b>-8.7</b>  |
| 2.     | Acrivastine | 5284514    | -8.33        | <b>-8.48</b> | <b>-8.73</b> |
| 3.     | Hydroxyzine | 3658       | -8.15        | -7.99        | <b>-8.08</b> |
| 4.     | Buclizine   | 6729       | -8.29        | <b>-8.27</b> | <b>-8.44</b> |
| 5.     | Azelastine  | 2267       | -8.13        | -8.02        | <b>-8.09</b> |
| 6.     | Cinnarizine | 1547484    | -7.89        | <b>-8.44</b> | <b>-7.96</b> |
| 7.     | Olopatadine | 5281071    | -7.83        | <b>-8.71</b> | <b>-8.13</b> |

Bilastine was approved in 2007 and it is used for the treatment of urticaria and allergic rhinoconjunctivitis. It is characterized as a highly potent and specific inhibitor of histamine H1 receptor [139].

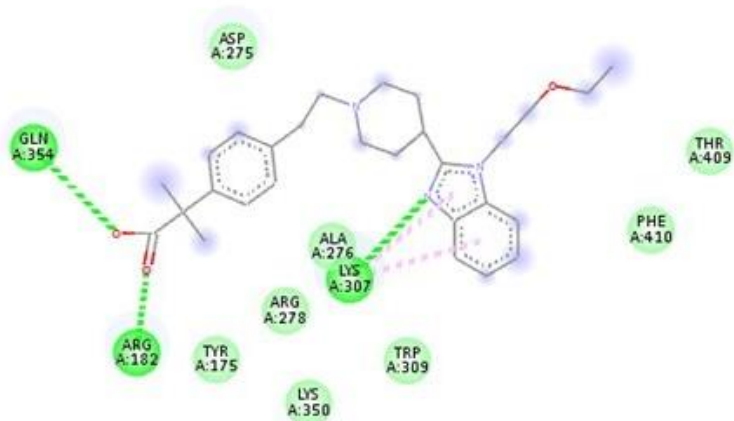
After analyzing the results, we observed that bilastine interacts with the Aspartate-255 residue of dopamine D2 receptor, Arginine-182 and Lysine-307 residues of dopamine D3 receptor via H-bond interaction. Similarly, bilastine interacts with the Arginine-182, Lysine-307 and Glutamine-354 residues of dopamine D4 receptor as shown in **Fig 4.3**. Van der waal and alkyl-alkyl interactions were also observed.



(a) Bilastine interaction with D2 receptor



(b) Bilastine interaction with D3 receptor



(c) Bilastine interaction with D4 receptor

**Interactions**

- |   |                            |   |          |
|---|----------------------------|---|----------|
|  | van der Waals              |  | Alkyl    |
|  | Conventional Hydrogen Bond |  | Pi-Alkyl |

**Figure 4.3: Interaction between bilastine and Dopamine D2-like receptors**

#### 4.6 DRUG-LIKENESS OF THE SELECTED LIGANDS

The ADMET analysis and pharmacokinetic study are extremely imperative phases in the process of drug discovery and development. It was carried out with help of ChEMBL and SwissADME.

**Table V: Physicochemical properties of selected ligands**

| Properties  | Drug Name      |                    |                |                    |                    |                    |                    |
|---|----------------|--------------------|----------------|--------------------|--------------------|--------------------|--------------------|
|   | Bilastine      | Acrivastine        | Hydroxyzine    | Buclizine          | Azelastine         | Cinnarizine        | Olopatadine        |
| <b>Molecular weight (g/mol)</b>                       | 463.62         | 348.44             | 374.90         | 433.03             | 381.90             | 368.51             | 337.41             |
| <b>H-bond donors</b>                                  | 1              | 1                  | 1              | 0                  | 0                  | 0                  | 1                  |
| <b>H-bond acceptors</b>                               | 5              | 4                  | 4              | 2                  | 3                  | 2                  | 4                  |
| <b>Molar refractivity</b>                             | 140.53         | 109.18             | 113.12         | 140.20             | 115.27             | 125.86             | 99.23              |
| <b>Topological polar surface area (Å<sup>2</sup>)</b> | 67.59          | 53.43              | 35.94          | 6.48               | 38.13              | 6.48               | 49.77              |
| <b>Lipinski's rule of five</b>                        | ✓              | ✓                  | ✓              | ✓<br>one violation | ✓<br>one violation | ✓<br>one violation | ✓                  |
| <b>Log P</b>  | 4.03           | 3.43               | 3.94           | 5.01               | 3.85               | 4.32               | 2.97               |
| <b>Gastro Intestinal absorption</b>                   | High           | High               | High           | Low                | High               | High               | High               |
| <b>Bioavailability score</b>                          | 0.55           | 0.55               | 0.55           | 0.55               | 0.55               | 0.55               | 0.55               |
| <b>BBB permeant</b>                                   | Yes            | Yes                | Yes            | No                 | Yes                | Yes                | Yes                |
| <b>Solubility</b>                                     | Poorly soluble | Moderately soluble | Poorly soluble | Poorly soluble     | Poorly soluble     | Poorly soluble     | Moderately soluble |

Selection of drug must be done after estimating both the short-term benefits and long-term consequences. Another factor while selecting the drug is to consider patient-specific factors, for example level of disability, functional & chronologic age, drug tolerance and expected efficacy [140].

## CHAPTER – 5

### CONCLUSION AND FUTURE PERSPECTIVE

Current drugs used for the treatment of PD have several adverse effects to overcome these side effects alternate medication is required. The goal is to achieve a drug or combination of drugs that could have maximum efficacy with minimal adverse effects. Aberrant indirect pathway signaling mediated via dopamine D2-like receptors and upregulated histaminergic signaling have been observed as a major contributor in the pathophysiology of PD. Thus, antihistamine drugs might show therapeutic effects against this aberrant signaling cascade.

The docking of the ligands was carefully performed and their interactions were observed. From this study, it can be considered that Bilastine, an inhibitor of histamine H1 receptor possess high binding affinity for dopamine D2-like receptors. Thus, the information gained on the basis of docking score postulates a piece of preliminary evidence for its potential as a dopamine D2-like receptors inhibitor and anti-parkinsonian medication. It may be implemented in designing effective therapeutics for PD in the future.

In addition, structures similar to bilastine might show similar results. Thus, they can also be screened against dopamine D2-like receptors. Moreover, the selected compounds can be checked by exploration in cell culture assay. Also, the effect of compounds can be studied by cell viability and LDH assay. These experiments will adjunct the above data pertaining to the therapeutic competence of these chosen compounds to be used for PD.

## References

- [1] D. W. Dickson, “Neuropathology of Parkinson Disease,” *Parkinsonism Relat. Disord.*, vol. 46, no. Suppl 1, p. S30, Jan. 2018, doi: 10.1016/J.PARKRELDIS.2017.07.033.
- [2] L. V. Kalia and A. E. Lang, “Parkinson’s disease,” *Lancet*, vol. 386, no. 9996, pp. 896–912, Aug. 2015, doi: 10.1016/S0140-6736(14)61393-3.
- [3] D. Yadav and P. Kumar, “Restoration and targeting of aberrant neurotransmitters in Parkinson’s disease therapeutics,” *Neurochem. Int.*, vol. 156, p. 105327, Jun. 2022, doi: 10.1016/J.NEUINT.2022.105327.
- [4] S. Zhai, W. Shen, S. M. Graves, and D. J. Surmeier, “Dopaminergic modulation of striatal function and Parkinson’s disease,” *Journal of Neural Transmission*. Springer-Verlag Wien, 2019, doi: 10.1007/s00702-019-01997-y.
- [5] A. C. Kreitzer and R. C. Malenka, “Striatal Plasticity and Basal Ganglia Circuit Function,” *Neuron*, vol. 60, no. 4, pp. 543–554, Nov. 26, 2008, doi: 10.1016/j.neuron.2008.11.005.
- [6] H. S. Moghaddam, A. Zare-Shahabadi, F. Rahmani, and N. Rezaei, “Neurotransmission systems in Parkinson’s disease,” *Rev. Neurosci.*, vol. 28, no. 5, pp. 509–536, Jul. 2017, doi: 10.1515/revneuro-2016-0068.
- [7] P. Wichit, S. Thanprasertsuk, O. Phokaewvarangkul, R. Bhidayasiri, and S. Bongsebandhu-phubhakdi, “Monoamine Levels and Parkinson’s Disease Progression: Evidence From a High-Performance Liquid Chromatography Study,” *Front. Neurosci.*, vol. 15, Jul. 2021, doi: 10.3389/fnins.2021.605887.
- [8] T. H. Johnston *et al.*, “Pridopidine, a clinic-ready compound, reduces 3,4-dihydroxyphenylalanine-induced dyskinesia in Parkinsonian macaques,” *Mov. Disord.*, vol. 34, no. 5, pp. 708–716, May 2019, doi: 10.1002/mds.27565.
- [9] S. Ztaou and M. Amalric, “Contribution of cholinergic interneurons to striatal pathophysiology in Parkinson’s disease,” *Neurochemistry International*, vol. 126. Elsevier Ltd, pp. 1–10, Jun. 01, 2019, doi: 10.1016/j.neuint.2019.02.019.
- [10] Y. Hui, C. Du, T. Xu, Q. Zhang, H. Tan, and J. Liu, “Dopamine D4 receptors in the lateral habenula regulate depression-related behaviors via a pre-synaptic mechanism in experimental Parkinson’s disease,” *Neurochem. Int.*, vol. 140, Nov. 2020, doi: 10.1016/j.neuint.2020.104844.
- [11] O. V. Anichtchik, J. O. Rinne, H. Kalimo, and P. Panula, “An altered histaminergic innervation of the substantia nigra in Parkinson’s disease,” *Exp. Neurol.*, vol. 163, no. 1, pp. 20–30, 2000, doi: 10.1006/exnr.2000.7362.

- [12] R. E. Brown, D. R. Stevens, and H. L. Haas, "The physiology of brain histamine," 2001. [Online]. Available: [www.elsevier.com/locate/pneurobio](http://www.elsevier.com/locate/pneurobio).
- [13] M. E. Parsons and C. R. Ganellin, "Histamine and its receptors," *British Journal of Pharmacology*, vol. 147, no. SUPPL. 1. Jan. 2006, doi: 10.1038/sj.bjp.0706440.
- [14] N. Titova, C. Padmakumar, S. J. G. Lewis, and K. R. Chaudhuri, "Parkinson's: a syndrome rather than a disease?," *Journal of Neural Transmission*, vol. 124, no. 8. Springer-Verlag Wien, pp. 907–914, Aug. 01, 2017, doi: 10.1007/s00702-016-1667-6.
- [15] T. R. Mhyre, J. T. Boyd, R. W. Hamill, and K. A. Maguire-Zeiss, "Parkinson's disease," *Subcell. Biochem.*, vol. 65, pp. 389–455, May 2012, doi: 10.1007/978-94-007-5416-4\_16.
- [16] G. J. A. Macphee and D. A. Stewart, "Parkinson's disease - Pathology, aetiology and diagnosis," *Reviews in Clinical Gerontology*, vol. 22, no. 3. pp. 165–178, Aug. 2012, doi: 10.1017/S095925981200007X.
- [17] S. Cerri, L. Mus, and F. Blandini, "Parkinson's Disease in Women and Men: What's the Difference?," *Journal of Parkinson's Disease*, vol. 9, no. 3. IOS Press, pp. 501–515, 2019, doi: 10.3233/JPD-191683.
- [18] H. Braak, K. Del Tredici-Braak, and T. Gasser, "Special issue 'Parkinson's disease,'" *Cell and Tissue Research*, vol. 373, no. 1. Springer Verlag, Jul. 01, 2018, doi: 10.1007/s00441-018-2863-5.
- [19] L. V. Kalia and A. E. Lang, "Parkinson's disease," *The Lancet*, vol. 386, no. 9996. Lancet Publishing Group, pp. 896–912, Aug. 29, 2015, doi: 10.1016/S0140-6736(14)61393-3.
- [20] A. H. Schapira and P. Jenner, "Etiology and pathogenesis of Parkinson's disease," *Movement Disorders*, vol. 26, no. 6. pp. 1049–1055, May 2011, doi: 10.1002/mds.23732.
- [21] J. Jankovic, "Parkinson's disease: Clinical features and diagnosis," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 79, no. 4. BMJ Publishing Group, pp. 368–376, 2008, doi: 10.1136/jnnp.2007.131045.
- [22] S. G. Reich and J. M. Savitt, "Parkinson's Disease," *Medical Clinics of North America*, vol. 103, no. 2. W.B. Saunders, pp. 337–350, Mar. 01, 2019, doi: 10.1016/j.mcna.2018.10.014.
- [23] H. Reichmann, "Premotor Diagnosis of Parkinson's Disease," *Neuroscience Bulletin*, vol. 33, no. 5. Springer Singapore, pp. 526–534, Oct. 01, 2017, doi: 10.1007/s12264-017-0159-5.

- [24] P. Mahlknecht, K. Seppi, and W. Poewe, “The concept of prodromal Parkinson’s disease,” *Journal of Parkinson’s Disease*, vol. 5, no. 4. IOS Press, pp. 681–697, Nov. 21, 2015, doi: 10.3233/JPD-150685.
- [25] U. K. Rinne, “Brain neurotransmitter receptors in Parkinson’s disease,” in *Movement Disorders*, Elsevier, 1981, pp. 59–74.
- [26] A. Manson, P. Stirpe, and A. Schrag, “Levodopa-induced-dyskinesias clinical features, incidence, risk factors, management and impact on quality of life,” *Journal of Parkinson’s Disease*, vol. 2, no. 3. pp. 189–198, 2012, doi: 10.3233/JPD-2012-120103.
- [27] J. Y. Hong *et al.*, “Rapid drug increase and early onset of levodopa-induced dyskinesia in Parkinson’s disease,” *PLoS One*, vol. 15, no. 8 August, Aug. 2020, doi: 10.1371/journal.pone.0237472.
- [28] S. Jamwal and P. Kumar, “Insight Into the Emerging Role of Striatal Neurotransmitters in the Pathophysiology of Parkinson’s Disease and Huntington’s Disease: A Review,” *Curr. Neuropharmacol.*, vol. 17, no. 2, pp. 165–175, Mar. 2018, doi: 10.2174/1570159x16666180302115032.
- [29] L. Brichta, P. Greengard, and M. Flajolet, “Advances in the pharmacological treatment of Parkinson’s disease: Targeting neurotransmitter systems,” *Trends in Neurosciences*, vol. 36, no. 9. pp. 543–554, Sep. 2013, doi: 10.1016/j.tins.2013.06.003.
- [30] M. E. Freitas and S. H. Fox, “Nondopaminergic treatments for Parkinson’s disease: current and future prospects,” *Neurodegenerative disease management*, vol. 6, no. 3. pp. 249–268, Jun. 01, 2016, doi: 10.2217/nmt-2016-0005.
- [31] J. Cummings *et al.*, “Pimavanserin for patients with Parkinson’s disease psychosis: A randomised, placebo-controlled phase 3 trial,” *Lancet*, vol. 383, no. 9916, pp. 533–540, 2014, doi: 10.1016/S0140-6736(13)62106-6.
- [32] S. H. Isaacson, B. Coate, J. Norton, and S. Stankovic, “Blinded SAPS-PD Assessment after 10 Weeks of Pimavanserin Treatment for Parkinson’s Disease Psychosis,” *J. Parkinsons. Dis.*, vol. 10, no. 4, pp. 1389–1396, 2020, doi: 10.3233/JPD-202047.
- [33] P. Huot and S. H. Fox, “Nondopaminergic treatments for Parkinson’s disease,” *Neurodegener. Dis. Manag.*, vol. 1, no. 6, pp. 491–512, Dec. 2011, doi: 10.2217/nmt.11.62.
- [34] S. Sveinbjornsdottir, “The clinical symptoms of Parkinson’s disease,” *J. Neurochem.*, pp. 318–324, Oct. 2016, doi: 10.1111/jnc.13691.



- [35] R. F. Pfeiffer, “Non-motor symptoms in Parkinson’s disease,” *Park. Relat. Disord.*, vol. 22, pp. S119–S122, Jan. 2016, doi: 10.1016/j.parkreldis.2015.09.004.
- [36] A. H. V. Schapira, K. R. Chaudhuri, and P. Jenner, “Non-motor features of Parkinson disease,” *Nat. Rev. Neurosci.*, vol. 18, no. 7, pp. 435–450, Jul. 2017, doi: 10.1038/nrn.2017.62.
- [37] M. Rodríguez-Violante, R. Zerón-Martínez, A. Cervantes-Arriaga, and T. Corona, “Who Can Diagnose Parkinson’s Disease First? Role of Pre-motor Symptoms,” *Arch. Med. Res.*, vol. 48, no. 3, pp. 221–227, Apr. 2017, doi: 10.1016/j.arcmed.2017.08.005.
- [38] A. Berardelli *et al.*, “EFNS/MDS-ES/ENS [corrected] recommendations for the diagnosis of Parkinson’s disease,” *Eur. J. Neurol.*, vol. 20, no. 1, pp. 16–34, Jan. 2013, doi: 10.1111/ENE.12022.
- [39] M. Politis, “Neuroimaging in Parkinson disease: from research setting to clinical practice,” *Nat. Rev. Neurol.*, vol. 10, no. 12, pp. 708–722, Dec. 2014, doi: 10.1038/NRNEUROL.2014.205.
- [40] G. Pagano, F. Niccolini, and M. Politis, “Imaging in Parkinson’s disease,” *Clin. Med. J. R. Coll. Physicians London*, vol. 16, no. 4, pp. 371–375, Aug. 2016, doi: 10.7861/clinmedicine.16-4-371.
- [41] C. P. Weingarten, M. H. Sundman, P. Hickey, and N. kwei Chen, “Neuroimaging of Parkinson’s disease: Expanding views,” *Neurosci. Biobehav. Rev.*, vol. 59, pp. 16–52, Dec. 2015, doi: 10.1016/j.neubiorev.2015.09.007.
- [42] D. Yadav and P. Kumar, “Parkinson’s Disease: An Overview and Role of Glutamate and its Receptors: Glutamate receptor based therapy as an alternate way to treat Parkinson’s,” in *2021 5th International Conference on Information Systems and Computer Networks (ISCON)*, Oct. 2021, pp. 1–5, doi: 10.1109/ISCON52037.2021.9702338.
- [43] P. A. L. Witt and S. Fahn, “Levodopa therapy for Parkinson disease: A look backward and forward,” *Neurology*, vol. 86, no. 14 Suppl 1, pp. S3–S12, Apr. 2016, doi: 10.1212/WNL.0000000000002509.
- [44] M. E. Freitas, M. Ruiz-Lopez, and S. H. Fox, “Novel Levodopa Formulations for Parkinson’s Disease,” *CNS Drugs*, vol. 30, no. 11, pp. 1079–1095, Nov. 2016, doi: 10.1007/S40263-016-0386-8.
- [45] A. Hsu, H. M. Yao, S. Gupta, and N. B. Modi, “Comparison of the pharmacokinetics of an oral extended-release capsule formulation of carbidopa-levodopa (IPX066) with immediate-release carbidopa-levodopa (Sinemet®), sustained-release carbidopa-levodopa (Sinemet® CR), and carbidopa-levodopa-

- entacapone (Stalevo®),” *J. Clin. Pharmacol.*, vol. 55, no. 9, pp. 995–1003, Sep. 2015, doi: 10.1002/JCPH.514.
- [46] W. G. Ondo, L. Shinawi, and S. Moore, “Comparison of orally dissolving carbidopa/levodopa (Parcopa) to conventional oral carbidopa/levodopa: A single-dose, double-blind, double-dummy, placebo-controlled, crossover trial,” *Mov. Disord.*, vol. 25, no. 16, pp. 2724–2727, Dec. 2010, doi: 10.1002/MDS.23158.
- [47] S. M. Hoy, “Levodopa/Carbidopa Enteral Suspension: A Review in Advanced Parkinson’s Disease,” *Drugs*, vol. 79, no. 15, pp. 1709–1718, Oct. 2019, doi: 10.1007/S40265-019-01201-1.
- [48] A. Mittur, S. Gupta, and N. B. Modi, “Pharmacokinetics of Rytary ®, An Extended-Release Capsule Formulation of Carbidopa-Levodopa,” *Clin. Pharmacokinet.*, vol. 56, no. 9, pp. 999–1014, Sep. 2017, doi: 10.1007/S40262-017-0511-Y.
- [49] M. Ahmari, A. Sharafi, J. Mahmoudi, I. Jafari-Anarkoli, M. Gharbavi, and M. J. Hosseini, “Selegiline (L-Deprenyl) Mitigated Oxidative Stress, Cognitive Abnormalities, and Histopathological Change in Rats: Alternative Therapy in Transient Global Ischemia,” *J. Mol. Neurosci.*, vol. 70, no. 10, pp. 1639–1648, Oct. 2020, doi: 10.1007/S12031-020-01544-5.
- [50] H. A. Blair and S. Dhillon, “Safinamide: A Review in Parkinson’s Disease,” *CNS Drugs*, vol. 31, no. 2, pp. 169–176, Feb. 2017, doi: 10.1007/S40263-017-0408-1.
- [51] Y. Chang, L. B. Wang, D. Li, K. Lei, and S. Y. Liu, “Efficacy of rasagiline for the treatment of Parkinson’s disease: an updated meta-analysis,” *Ann. Med.*, vol. 49, no. 5, pp. 421–434, Jul. 2017, doi: 10.1080/07853890.2017.1293285.
- [52] J. J. Van Hilten, C. C. Ramaker, R. L. Stowe, and N. J. Ives, “Bromocriptine versus levodopa in early Parkinson’s disease,” *Cochrane database Syst. Rev.*, vol. 2007, no. 4, 2007, doi: 10.1002/14651858.CD002258.PUB2.
- [53] C. E. Clarke and K. H. O. Deane, “Cabergoline for levodopa-induced complications in Parkinson’s disease,” *Cochrane database Syst. Rev.*, vol. 2001, no. 1, Jan. 2001, doi: 10.1002/14651858.CD001518.
- [54] A. Ciobica, Z. Olteanu, M. Padurariu, and L. Hritcu, “The effects of pergolide on memory and oxidative stress in a rat model of Parkinson’s disease,” *J. Physiol. Biochem.*, vol. 68, no. 1, pp. 59–69, Mar. 2012, doi: 10.1007/S13105-011-0119-X.
- [55] J. E. Frampton, “Pramipexole extended-release: a review of its use in patients with Parkinson’s disease,” *Drugs*, vol. 74, no. 18, pp. 2175–2190, 2014, doi: 10.1007/S40265-014-0322-5.

- [56] F. Carbone, A. Djamshidian, K. Seppi, and W. Poewe, “Apomorphine for Parkinson’s Disease: Efficacy and Safety of Current and New Formulations,” *CNS Drugs*, vol. 33, no. 9, pp. 905–918, Sep. 2019, doi: 10.1007/S40263-019-00661-Z.
- [57] D. Q. Pham and A. Nogid, “Rotigotine transdermal system for the treatment of Parkinson’s disease,” *Clin. Ther.*, vol. 30, no. 5, pp. 813–824, May 2008, doi: 10.1016/J.CLINTHERA.2008.05.007.
- [58] N. Hattori *et al.*, “Ropinirole Patch Versus Placebo, Ropinirole Extended-Release Tablet in Advanced Parkinson’s Disease,” *Mov. Disord.*, vol. 35, no. 9, pp. 1565–1573, Sep. 2020, doi: 10.1002/MDS.28071.
- [59] S. Perez-Lloret and O. Rascol, “Piribedil for the Treatment of Motor and Non-motor Symptoms of Parkinson Disease,” *CNS Drugs*, vol. 30, no. 8, pp. 703–717, Aug. 2016, doi: 10.1007/S40263-016-0360-5.
- [60] L. J. Scott, “Opicapone: A Review in Parkinson’s Disease,” *CNS Drugs*, vol. 35, no. 1, pp. 121–131, Jan. 2021, doi: 10.1007/S40263-020-00778-6.
- [61] X. Liao, N. Wu, D. Liu, B. Shuai, S. Li, and K. Li, “Levodopa/carbidopa/entacapone for the treatment of early Parkinson’s disease: a meta-analysis,” *Neurol. Sci.*, vol. 41, no. 8, pp. 2045–2054, Aug. 2020, doi: 10.1007/S10072-020-04303-X.
- [62] C. A. Artusi, L. Sarro, G. Imbalzano, M. Fabbri, and L. Lopiano, “Safety and efficacy of tolcapone in Parkinson’s disease: systematic review,” *Eur. J. Clin. Pharmacol.*, vol. 77, no. 6, pp. 817–829, Jun. 2021, doi: 10.1007/S00228-020-03081-X.
- [63] J. H. Friedman, W. C. Koller, M. C. Lannon, K. Busenbark, E. Swanson-Hyland, and D. Smith, “Benzotropine versus clozapine for the treatment of tremor in Parkinson’s disease,” *Neurology*, vol. 48, no. 4, pp. 1077–1081, 1997, doi: 10.1212/WNL.48.4.1077.
- [64] L. K. Sahoo *et al.*, “Comparison of effectiveness of trihexyphenidyl and levodopa on motor symptoms in Parkinson’s disease,” *J. Neural Transm.*, vol. 127, no. 12, pp. 1599–1606, Dec. 2020, doi: 10.1007/S00702-020-02257-0.
- [65] D. M. Paton, “Istradefylline: adenosine A2A receptor antagonist to reduce ‘OFF’ time in Parkinson’s disease,” *Drugs Today (Barc.)*, vol. 56, no. 2, pp. 125–134, 2020, doi: 10.1358/DOT.2020.56.2.3098156.
- [66] A. Goel, R. Sugumaran, and S. K. Narayan, “Zonisamide in Parkinson’s disease: a current update,” *Neurol. Sci.*, vol. 42, no. 10, pp. 4123–4129, Oct. 2021, doi: 10.1007/S10072-021-05550-2.

- [67] O. Rascol, M. Fabbri, and W. Poewe, “Amantadine in the treatment of Parkinson’s disease and other movement disorders,” *Lancet. Neurol.*, vol. 20, no. 12, pp. 1048–1056, Dec. 2021, doi: 10.1016/S1474-4422(21)00249-0.
- [68] N. Chuhma, K. F. Tanaka, and T. Nagai, “The physiology and pathophysiology of basal ganglia: From signal transduction to circuits,” *Neurochem. Int.*, vol. 131, Dec. 2019, doi: 10.1016/J.NEUINT.2019.104544.
- [69] S. Grillner and B. Robertson, “The Basal Ganglia Over 500 Million Years,” *Curr. Biol.*, vol. 26, no. 20, pp. R1088–R1100, Oct. 2016, doi: 10.1016/J.CUB.2016.06.041.
- [70] F. Blandini and J. T. Greenamyre, “Prospects of glutamate antagonists in the therapy of Parkinson’s disease,” *Fundam. Clin. Pharmacol.*, vol. 12, no. 1, pp. 4–12, 1998, doi: 10.1111/J.1472-8206.1998.TB00918.X.
- [71] F. Blandini, R. H. P. Porter, and J. T. Greenamyre, “Glutamate and Parkinson’s disease,” *Mol. Neurobiol.*, vol. 12, no. 1, pp. 73–94, 1996, doi: 10.1007/BF02740748.
- [72] T. Wu *et al.*, “Basal ganglia circuits changes in Parkinson’s disease patients,” *Neurosci. Lett.*, vol. 524, no. 1, pp. 55–59, Aug. 2012, doi: 10.1016/j.neulet.2012.07.012.
- [73] H. Bergman, T. Wichmann, and M. R. DeLong, “Reversal of experimental parkinsonism by lesions of the subthalamic nucleus,” *Science*, vol. 249, no. 4975, pp. 1436–1438, 1990, doi: 10.1126/science.2402638.
- [74] T. Wichmann, M. A. Kliem, and M. R. DeLong, “Antiparkinsonian and behavioral effects of inactivation of the substantia nigra pars reticulata in hemiparkinsonian primates,” *Exp. Neurol.*, vol. 167, no. 2, pp. 410–424, 2001, doi: 10.1006/exnr.2000.7572.
- [75] D. M. Lieberman, M. E. Corthesy, A. Cummins, and E. H. Oldfield, “Reversal of experimental parkinsonism by using selective chemical ablation of the medial globus pallidus,” *J. Neurosurg.*, vol. 90, no. 5, pp. 924–938, 1999, doi: 10.3171/jns.1999.90.5.0928.
- [76] J. A. Obeso *et al.*, “Functional organization of the basal ganglia: Therapeutic implications for Parkinson’s disease,” *Mov. Disord.*, vol. 23, no. SUPPL. 3, 2008, doi: 10.1002/mds.22062.
- [77] D. J. Kuhman, H. C. Walker, and C. P. Hurt, “Dopamine-mediated improvements in dynamic balance control in Parkinson’s disease,” *Gait Posture*, vol. 82, pp. 68–74, Oct. 2020, doi: 10.1016/j.gaitpost.2020.08.132.

- [78] J. O. Rinne, “Endogenous dopamine release in Parkinson’s disease,” *Lancet. Neurol.*, vol. 2, no. 8, pp. 460–461, 2003, doi: 10.1016/s1474-4422(03)00480-0.
- [79] J. Hietala’ and E. Syvalahti, “Special Section: Schizophrenia Dopamine in Schizophrenia,” 1996.
- [80] M. Hallett, “Tourette Syndrome: Update,” *Brain and Development*, vol. 37, no. 7. Elsevier B.V., pp. 651–655, Aug. 01, 2015, doi: 10.1016/j.braindev.2014.11.005.
- [81] U. Bonuccelli and N. Pavese, “Dopamine agonists in the treatment of Parkinson’s disease,” *Expert Review of Neurotherapeutics*, vol. 6, no. 1. pp. 81–89, Jan. 2006, doi: 10.1586/14737175.6.1.81.
- [82] P. Foley, M. Gerlach, K. L. Double, and P. Riederer, “Dopamine receptor agonists in the therapy of Parkinson’s disease,” *Journal of Neural Transmission*, vol. 111, no. 10–11. pp. 1375–1446, Oct. 2004, doi: 10.1007/s00702-003-0059-x.
- [83] S. M. Wang *et al.*, “Investigational dopamine antagonists for the treatment of schizophrenia,” *Expert Opinion on Investigational Drugs*, vol. 26, no. 6. Taylor and Francis Ltd, pp. 687–698, Jun. 03, 2017, doi: 10.1080/13543784.2017.1323870.
- [84] M. R. DeLong and T. Wichmann, “Basal ganglia circuits as targets for neuromodulation in Parkinson disease,” *JAMA Neurology*, vol. 72, no. 11. American Medical Association, pp. 1354–1360, Nov. 01, 2015, doi: 10.1001/jamaneurol.2015.2397.
- [85] J. N. Cremer, K. Amunts, J. Graw, M. Piel, F. Rösch, and K. Zilles, “Neurotransmitter receptor density changes in Pitx3ak mice - A model relevant to parkinson’s disease,” *Neuroscience*, vol. 285, pp. 11–23, Jan. 2015, doi: 10.1016/j.neuroscience.2014.10.050.
- [86] G. Sitzia, I. Mantas, X. Zhang, P. Svenningsson, and K. Chergui, “NMDA receptors are altered in the substantia nigra pars reticulata and their blockade ameliorates motor deficits in experimental parkinsonism,” *Neuropharmacology*, vol. 174, Sep. 2020, doi: 10.1016/j.neuropharm.2020.108136.
- [87] Y. Kang *et al.*, “18F-FPEB PET/CT Shows mGluR5 Upregulation in Parkinson’s Disease,” *J. Neuroimaging*, vol. 29, no. 1, pp. 97–103, Jan. 2019, doi: 10.1111/jon.12563.
- [88] K. Farmer *et al.*, “mGluR5 Allosteric Modulation Promotes Neurorecovery in a 6-OHDA-Toxicant Model of Parkinson’s Disease,” *Mol. Neurobiol.*, vol. 57, no. 3, pp. 1418–1431, Mar. 2020, doi: 10.1007/s12035-019-01818-z.
- [89] Z. Liu *et al.*, “Multifunctional memantine nitrate significantly protects against

- glutamate-induced excitotoxicity via inhibiting calcium influx and attenuating PI3K/Akt/GSK3beta pathway,” *Chem. Biol. Interact.*, vol. 325, Jul. 2020, doi: 10.1016/j.cbi.2020.109020.
- [90] L. L. Mariani, S. Longueville, J. A. Girault, D. Hervé, and N. Gervasi, “Differential enhancement of ERK, PKA and Ca<sup>2+</sup> signaling in direct and indirect striatal neurons of Parkinsonian mice,” *Neurobiol. Dis.*, vol. 130, Oct. 2019, doi: 10.1016/j.nbd.2019.104506.
- [91] P. De Deurwaerdère and G. Di Giovanni, “Serotonin in health and disease,” *International Journal of Molecular Sciences*, vol. 21, no. 10. MDPI AG, May 02, 2020, doi: 10.3390/ijms21103500.
- [92] M. Berger, J. A. Gray, and B. L. Roth, “The expanded biology of serotonin,” *Annual Review of Medicine*, vol. 60. pp. 355–366, 2009, doi: 10.1146/annurev.med.60.042307.110802.
- [93] M. Politis and F. Niccolini, “Serotonin in Parkinson’s disease,” *Behavioural Brain Research*, vol. 277. Elsevier, pp. 136–145, Jan. 05, 2015, doi: 10.1016/j.bbr.2014.07.037.
- [94] G. Pagano and M. Politis, “Molecular Imaging of the Serotonergic System in Parkinson’s Disease,” in *International Review of Neurobiology*, vol. 141, Academic Press Inc., 2018, pp. 173–210.
- [95] H. K. Park, J. J. Lee, and Y. M. Park, “Preserved Serotonergic Activity in Early-Onset Parkinson’s Disease,” *Can. J. Neurol. Sci.*, vol. 47, no. 3, pp. 344–349, May 2020, doi: 10.1017/cjn.2019.322.
- [96] R. L. O’Gorman Tuura, C. R. Baumann, and H. Baumann-Vogel, “Beyond dopamine: GABA, glutamate, and the axial symptoms of Parkinson disease,” *Front. Neurol.*, vol. 9, no. SEP, Sep. 2018, doi: 10.3389/fneur.2018.00806.
- [97] van Nuland A. *et al.*, “GABAergic changes in the thalamocortical circuit in Parkinson’s disease,” *Hum. Brain Mapp.*, vol. 41, no. 4, pp. 1017–1029, 2019, doi: 10.1002/hbm.24857.
- [98] I. L. Boccularo, C. Schwerdel, L. Cristiá-Lara, J. M. Fritschy, and L. Rubi, “Dopamine depletion induces neuron-specific alterations of GABAergic transmission in the mouse striatum,” *Eur. J. Neurosci.*, vol. 52, no. 5, pp. 3353–3374, Sep. 2020, doi: 10.1111/ejn.14886.
- [99] M. König *et al.*, “Increased Cholinergic Response in  $\alpha$ -Synuclein Transgenic Mice (h- $\alpha$ -synL62),” *ACS Chem. Neurosci.*, vol. 10, no. 4, pp. 1915–1922, Apr. 2019, doi: 10.1021/acchemneuro.8b00274.

- [100] S. Ztaou, J. Lhost, I. Watabe, G. Torromino, and M. Amalric, “Striatal cholinergic interneurons regulate cognitive and affective dysfunction in partially dopamine-depleted mice,” *Eur. J. Neurosci.*, vol. 48, no. 9, pp. 2988–3004, Nov. 2018, doi: 10.1111/ejn.14153.
- [101] J. H. Lee *et al.*, “Muscarinic M1 Receptor Coupling to G-protein is Intact in Parkinson’s Disease Dementia,” *J. Parkinsons. Dis.*, vol. 6, no. 4, pp. 733–739, 2016, doi: 10.3233/JPD-160932.
- [102] S. J. Colloby, I. G. Mckeith, D. J. Burn, D. J. Wyper, J. T. O’Brien, and J.-P. Taylor, “Cholinergic and perfusion brain networks in Parkinson disease dementia,” 2016. [Online]. Available: <http://www-01.ibm.com/software/analytics/spss/>.
- [103] C. Romeo, A. T. Raveendran, N. M. Sobha, and C. S. Paulose, “Cholinergic receptor alterations in the brain stem of spinal cord injured Rats,” *Neurochem. Res.*, vol. 38, no. 2, pp. 389–397, Feb. 2013, doi: 10.1007/s11064-012-0931-x.
- [104] A. Iarkov, C. Mendoza, and V. Echeverria, “Cholinergic Receptor Modulation as a Target for Preventing Dementia in Parkinson’s Disease,” *Frontiers in Neuroscience*, vol. 15. Frontiers Media S.A., Sep. 20, 2021, doi: 10.3389/fnins.2021.665820.
- [105] S. J. Sara, “The locus coeruleus and noradrenergic modulation of cognition,” *Nature Reviews Neuroscience*, vol. 10, no. 3, pp. 211–223, Mar. 2009, doi: 10.1038/nrn2573.
- [106] M. J. Chen, T. V. Nguyen, C. J. Pike, and A. A. Russo-Neustadt, “Norepinephrine induces BDNF and activates the PI-3K and MAPK cascades in embryonic hippocampal neurons,” *Cell. Signal.*, vol. 19, no. 1, pp. 114–128, Jan. 2007, doi: 10.1016/j.cellsig.2006.05.028.
- [107] R. Cash, R. Raisman, L. Lanfumey, A. Ploska, and Y. Agid, “Cellular Localization of Adrenergic Receptors in Rat and Human Brain,” 1986.
- [108] S. E. Hyman, “Neurotransmitters,” *Current biology*, pp. 154–158, 2005.
- [109] K. A. Neve, J. K. Seamans, and H. Trantham-Davidson, “Dopamine Receptor Signaling,” *J. Recept. Signal Transduct. Res.*, vol. 24, no. 3, pp. 165–205, 2004, doi: 10.1081/lrst-200029981.
- [110] O. David, I. Barrera, N. Gould, S. Gal-Ben-Ari, and K. Rosenblum, “D1 Dopamine Receptor Activation Induces Neuronal eEF2 Pathway-Dependent Protein Synthesis,” *Front. Mol. Neurosci.*, vol. 13, May 2020, doi: 10.3389/fnmol.2020.00067.

- [111] A. Mishra, S. Singh, V. Tiwari, Parul, and S. Shukla, “Dopamine D1 receptor activation improves adult hippocampal neurogenesis and exerts anxiolytic and antidepressant-like effect via activation of Wnt/ $\beta$ -catenin pathways in rat model of Parkinson’s disease,” *Neurochem. Int.*, vol. 122, pp. 170–186, Jan. 2019, doi: 10.1016/j.neuint.2018.11.020.
- [112] J. Zhu *et al.*, “Dopamine D2 receptor restricts astrocytic NLRP3 inflammasome activation via enhancing the interaction of  $\beta$ -arrestin2 and NLRP3,” *Cell Death Differ.*, vol. 25, no. 11, pp. 2037–2049, Nov. 2018, doi: 10.1038/s41418-018-0127-2.
- [113] V. Mutti, C. Fiorentini, C. Missale, and F. Bono, “Dopamine D3 receptor heteromerization: Implications for neuroplasticity and neuroprotection,” *Biomolecules*, vol. 10, no. 7. MDPI AG, pp. 1–15, Jul. 01, 2020, doi: 10.3390/biom10071016.
- [114] S. Nuutinen and P. Panula, “Histamine in Neurotransmission and Brain Diseases,” *Adv. Exp. Med. Biol.*, vol. 709, pp. 95–107, 2010, doi: 10.1007/978-1-4419-8056-4\_10.
- [115] L. Shan, A. M. Bao, and D. F. Swaab, “The human histaminergic system in neuropsychiatric disorders,” *Trends in Neurosciences*, vol. 38, no. 3. Elsevier Ltd, pp. 167–177, Mar. 01, 2015, doi: 10.1016/j.tins.2014.12.008.
- [116] E. García-Martín, P. Ayuso, A. Luengo, C. Martínez, and J. A. G. Agúndez, “Genetic variability of histamine receptors in patients with Parkinson’s disease,” *BMC Med. Genet.*, vol. 9, Mar. 2008, doi: 10.1186/1471-2350-9-15.
- [117] D. Tweedie, K. Sambamurti, and N. H. Greig, “TNF-Inhibition as a Treatment Strategy for Neurodegenerative Disorders: New Drug Candidates and Targets,” 2007.
- [118] T. H. Johnston, A. Van Der Meij, J. M. Brotchie, and S. H. Fox, “Effect of histamine H2 receptor antagonism on levodopa-induced dyskinesia in the MPTP-macaque model of Parkinson’s disease,” *Mov. Disord.*, vol. 25, no. 10, pp. 1379–1390, Jul. 2010, doi: 10.1002/mds.23069.
- [119] T. A. Mestre *et al.*, “Famotidine, a Histamine H2 Receptor Antagonist, Does Not Reduce Levodopa-Induced Dyskinesia in Parkinson’s Disease: A Proof-of-Concept Study,” *Mov. Disord. Clin. Pract.*, vol. 1, no. 3, pp. 219–224, Sep. 2014, doi: 10.1002/mdc3.12061.
- [120] A. Avila-Luna, C. Ríos, A. Gálvez-Rosas, S. Montes, J. A. Arias-Montaño, and A. Bueno-Nava, “Chronic administration of the histamine H3 receptor agonist immpip decreases l-Dopa-induced dyskinesias in 6-hydroxydopamine-lesioned rats,” *Psychopharmacology (Berl.)*, vol. 236, no. 6, pp. 1937–1948, Jun. 2019, doi:



10.1007/s00213-019-5182-y.

- [121] T. Bartels, S. De Schepper, and S. Hong, “Microglia modulate neurodegeneration in Alzheimer’s and Parkinson’s diseases,” *Sci. (New York, N.Y.)*, vol. 370, no. 6512, pp. 66–69, 2020, doi: 10.1126/science.abb8587.
- [122] P. Zhou *et al.*, “Histamine-4 receptor antagonist JNJ7777120 inhibits pro-inflammatory microglia and prevents the progression of Parkinson-like pathology and behaviour in a rat model,” *Brain. Behav. Immun.*, vol. 76, pp. 61–73, Feb. 2019, doi: 10.1016/j.bbi.2018.11.006.
- [123] Y. Wang *et al.*, “Inhibition of striatal dopamine D5 receptor attenuates levodopa-induced dyskinesia in a rat model of Parkinson’s disease,” *Brain Res.*, vol. 1754, Mar. 2021, doi: 10.1016/j.brainres.2020.147266.
- [124] P. Yang, J. S. Perlmutter, T. L. S. Benzinger, J. C. Morris, and J. Xu, “Dopamine D3 receptor: A neglected participant in Parkinson Disease pathogenesis and treatment?,” *Ageing Research Reviews*, vol. 57. Elsevier Ireland Ltd, Jan. 01, 2020, doi: 10.1016/j.arr.2019.100994.
- [125] P. Yang *et al.*, “Dopamine D1 + D3 receptor density may correlate with parkinson disease clinical features,” *Ann. Clin. Transl. Neurol.*, vol. 8, no. 1, pp. 224–237, Jan. 2021, doi: 10.1002/acn3.51274.
- [126] S. M. Rocha *et al.*, “Histamine induces microglia activation and dopaminergic neuronal toxicity via H1 receptor activation,” *J. Neuroinflammation*, vol. 13, no. 1, Jun. 2016, doi: 10.1186/s12974-016-0600-0.
- [127] W. Zhang, X. Zhang, Y. Zhang, C. Qu, X. Zhou, and S. Zhang, “Histamine Induces Microglia Activation and the Release of Proinflammatory Mediators in Rat Brain Via H1R or H4R,” *J. Neuroimmune Pharmacol.*, vol. 15, no. 2, pp. 280–291, Jun. 2020, doi: 10.1007/s11481-019-09887-6.
- [128] M. R. Ahmed *et al.*, “Pharmacological antagonism of histamine H2R ameliorated L-DOPA-induced dyskinesia via normalization of GRK3 and by suppressing FosB and ERK in PD,” *Neurobiol. Aging*, vol. 81, pp. 177–189, Sep. 2019, doi: 10.1016/j.neurobiolaging.2019.06.004.
- [129] A. Sharma *et al.*, “Histamine H3 and H4 receptors modulate Parkinson’s disease induced brain pathology. Neuroprotective effects of nanowired BF-2649 and clobenpropit with anti-histamine-antibody therapy,” *Prog. Brain Res.*, vol. 266, pp. 1–73, 2021, doi: 10.1016/bs.pbr.2021.06.003.
- [130] L. Shan, G. J. M. Martens, and D. F. Swaab, “Histamine-4 Receptor: Emerging Target for the Treatment of Neurological Diseases,” *Curr. Top. Behav. Neurosci.*, 2021, doi: 10.1007/7854\_2021\_237.

- [131] Q. Fang *et al.*, “Histamine-4 receptor antagonist ameliorates Parkinson-like pathology in the striatum,” *Brain. Behav. Immun.*, vol. 92, pp. 127–138, Feb. 2021, doi: 10.1016/j.bbi.2020.11.036.
- [132] A. Bateman *et al.*, “UniProt: the universal protein knowledgebase in 2021,” *Nucleic Acids Res.*, vol. 49, no. D1, pp. D480–D489, Jan. 2021, doi: 10.1093/NAR/GKAA1100.
- [133] F. Sievers and D. G. Higgins, “The Clustal Omega Multiple Alignment Package,” *Methods Mol. Biol.*, vol. 2231, pp. 3–16, 2021, doi: 10.1007/978-1-0716-1036-7\_1.
- [134] S. K. Burley, H. M. Berman, G. J. Kleywegt, J. L. Markley, H. Nakamura, and S. Velankar, “Protein Data Bank (PDB): The Single Global Macromolecular Structure Archive,” *Methods Mol. Biol.*, vol. 1607, pp. 627–641, 2017, doi: 10.1007/978-1-4939-7000-1\_26.
- [135] D. S. Wishart *et al.*, “DrugBank 5.0: a major update to the DrugBank database for 2018,” *Nucleic Acids Res.*, vol. 46, no. D1, pp. D1074–D1082, Jan. 2018, doi: 10.1093/NAR/GKX1037.
- [136] A. Gaulton *et al.*, “The ChEMBL database in 2017,” *Nucleic Acids Res.*, vol. 45, no. D1, pp. D945–D954, Jan. 2017, doi: 10.1093/NAR/GKW1074.
- [137] G. Bitencourt-Ferreira and W. F. de Azevedo, “Docking with SwissDock,” *Methods Mol. Biol.*, vol. 2053, pp. 189–202, 2019, doi: 10.1007/978-1-4939-9752-7\_12.
- [138] K. B. Santos, I. A. Guedes, A. L. M. Karl, and L. E. Dardenne, “Highly Flexible Ligand Docking: Benchmarking of the DockThor Program on the LEADS-PEP Protein-Peptide Data Set,” *J. Chem. Inf. Model.*, vol. 60, no. 2, pp. 667–683, Feb. 2020, doi: 10.1021/ACS.JCIM.9B00905.
- [139] M. K. Church, M. Tiongco-Recto, E. Ridolo, and Z. Novák, “Bilastine: a lifetime companion for the treatment of allergies,” *Curr. Med. Res. Opin.*, vol. 36, no. 3, pp. 445–454, Mar. 2020, doi: 10.1080/03007995.2019.1681134.
- [140] J. J. Chen and D. M. Swope, “Pharmacotherapy for Parkinson’s disease,” *Pharmacotherapy*, vol. 27, no. 12 Pt 2, 2007, doi: 10.1592/PHCO.27.12PART2.161S.

## LIST OF PUBLICATION

1. D. Yadav and P. Kumar, “Parkinson’s Disease: An Overview and Role of Glutamate and its Receptors: Glutamate receptor based therapy as an alternate way to treat Parkinson’s,” in *2021 5th International Conference on Information Systems and Computer Networks (ISCON)*, Oct. 2021, pp. 1–5, doi: 10.1109/ISCON52037.2021.9702338.
2. Yadav D, Kumar P. Restoration and targeting of aberrant neurotransmitters in Parkinson's disease therapeutics. *Neurochem Int.* 2022 Mar 21;156:105327. doi: 10.1016/j.neuint.2022.105327. Epub ahead of print. PMID: 35331828.

PAPER NAME

**New Microsoft Word Document.docx**

AUTHOR

**Divya Thesis**

WORD COUNT

**12422 Words**

CHARACTER COUNT

**75790 Characters**

PAGE COUNT

**59 Pages**

FILE SIZE

**4.8MB**

SUBMISSION DATE

**May 1, 2022 9:01 AM GMT+5:30**

REPORT DATE

**May 1, 2022 9:05 AM GMT+5:30**

### ● 55% Overall Similarity

The combined total of all matches, including overlapping sources, for each database.

- 6% Internet database
- 52% Publications database
- Crossref database
- Crossref Posted Content database
- 6% Submitted Works database

### ● Excluded from Similarity Report

- Bibliographic material
- Quoted material
- Cited material
- Small Matches (Less than 10 words)

● **55% Overall Similarity**

Top sources found in the following databases:

- 6% Internet database
- 52% Publications database
- Crossref database
- Crossref Posted Content database
- 6% Submitted Works database

TOP SOURCES

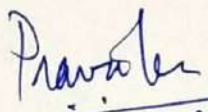
The sources with the highest number of matches within the submission. Overlapping sources will not be displayed.

|   |  |     |
|---|--|-----|
| 1 | <b>Divya Yadav, Pravir Kumar. "Restoration and targeting of aberrant neur...</b> | 34% |
|   | Crossref   |     |
| 2 | <b>Divya Yadav, Pravir Kumar. "Parkinson's Disease: An Overview and Rol...</b>   | 17% |
|   | Crossref   |     |
| 3 | <b>coursehero.com</b>  | 1%  |
|   | Internet   |     |
| 4 | <b>Kookmin University on 2020-06-02</b>  | <1% |
|   | Submitted works  |     |
| 5 | <b>dspace.dtu.ac.in:8080</b>   | <1% |
|   | Internet   |     |
| 6 | <b>Delhi Technological University on 2018-05-17</b>                              | <1% |
|   | Submitted works  |     |
| 7 | <b>Delhi Technological University on 2017-06-07</b>                              | <1% |
|   | Submitted works  |     |
| 8 | <b>University of Cincinnati on 2022-02-05</b>                                    | <1% |
|   | Submitted works  |     |

|    |  |     |
|----|--|-----|
| 9  | <b>ir.lib.uwo.ca</b><br>Internet                                       | <1% |
| 10 | <b>res.mdpi.com</b><br>Internet  | <1% |
| 11 | <b>Delhi Technological University on 2019-03-13</b><br>Submitted works | <1% |
| 12 | <b>Mahidol University on 2010-05-06</b><br>Submitted works             | <1% |
| 13 | <b>University of Malta on 2016-04-28</b><br>Submitted works            | <1% |
| 14 | <b>campus.uni-klu.ac.at</b><br>Internet                                | <1% |
| 15 | <b>documents.mx</b><br>Internet  | <1% |
| 16 | <b>biomedcentral.com</b><br>Internet                                   | <1% |
| 17 | <b>research.library.mun.ca</b><br>Internet                             | <1% |

To /  
✓ The Head Librarian,  
Delhi Technological University,  
Bawana Road, Shahbad Daultpur,  
New Delhi – 110042

Date: 02 May, 2022

  
02.05.2022  
Through ~~HoD~~, Biotechnology

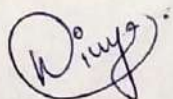
**Subject: Declaration of fulfilment of University's requirements for the award of M.Sc. Biotechnology**

Respected Sir/Madam,

I, **Divya Yadav**, final year student of **M.Sc. Biotechnology**, Roll no. **2K20/MSCBIO/06**, Department of Biotechnology, Delhi Technological University, hereby declare that the work which is presented in the Dissertation thesis entitled "**In silico analysis of antihistamine drugs as neuroprotectants targeting dopamine D2-like receptors in Parkinson's Disease**" in the fulfilment of the requirement for the award of Degree of Masters of Science in Biotechnology is an authentic record of my own work done under the supervision of **Prof. Pravit Kumar**.

The work attached to this declaration complies with the University's requirements and is my own work. The overall similarity is 55%, out of which 51% is from my own publications. The plagiarism report has been attached. Further, the work attached to this declaration has not been submitted in full or partial satisfaction of an academic award in another university.

Yours sincerely,



Divya Yadav  
M.Sc. Biotechnology – 2<sup>nd</sup> year  
2K20/MSCBIO/06