SYNTHESIS & CHARACTERIZATION OF BIO ACTIVE GLASS

AS

DOPANT IN GELATIN FILM

A Dissertation submitted in partial fulfillment of the requirement for the

Award of the degree of

MASTER OF TECHNOLOGY

IN

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By

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Certificate

This is to certify that Mr. SOURABH, M. Tech, student in the Department of Applied Chemistry And Polymer Technology has submitted a project report on "synthesis & characterization of bio active glass as dopant in gelatin film" in partial fulfillment of the requirement for award of degree of Master of Technology in Applied Chemistry and Polymer Technology, Delhi Technological University (formerly Delhi College of Engineering), Delhi, during the academic year 2014-16, is an authentic record. It is a record of the student's research work prepared under my supervision and guidance and has reached the requisite standards.

The work embodied in this major project has not been submitted for the award of any other degree to the best of our knowledge.

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ABSTRACT

Gelatin is a biodegradable, biocompatible and non immunogenic product, suitable for medical applications. To for various biomedical and other application of gelatin its mechanical, chemical and structural properties plays a vital role .So in the present study we are going to characterize various mechanical and structural properties of gelatin and bioglass (as a dopant) enriched gelatin films by various physical & mechanical characterization techniques like FTIR ,SEM ,XRD and UTM etc. With the help of various characterization techniques we can specify the various uses and application area of gelatin and gelatin doped materials.

CHAPTER 1

INTRODUCTION & LITERATURE REVIEW

1.1. INTRODUCTION:

Thermal denaturation or physical and chemical degradation of collagen involves the breaking of the triple-helix structure to give gelatin, a biodegradable, biocompatible and non immunogenic product, suitable for medical applications. At a temperature of about 403°C, gelatin aqueous solutions are in the sol state and form physical thermo reversible gels on cooling. During gelling, the chains undergo a conformational disorder order transition and tend to recover the collagen triple-helix structure. Gelatin is widely used in the pharmaceutical industry as well as in the biomedical: hard and soft capsules, microspheres, sealants for vascular prostheses, wound dressing and adsorbent pad for surgical use are among its most frequent applications. With respect to collagen, which is also known to have wide biomedical applications, gelatin does not express antigenicity in physiological conditions, and it is much cheaper and easier to obtain in concentrate solutions. On the other hand, gelatin exhibits poor mechanical properties which limit its possible applications as a biomaterial. Noticeable improvements of the mechanical properties of gelatin films in the direction of deformation have been obtained by inducing segmental orientation in gelatin "films through uniaxial stretching and successive air drying at constant elongation. The improvement of the mechanical properties of drawn gelatin films has been related to the renaturation level of the protein, evaluated through differential scanning calorimetry (d.s.c.). Since gelatin is soluble in aqueous solution, gelatin materials for long-term biomedical applications must be submitted to cross linking, which improves both the thermal and the mechanical stability of the biopolymer bioglass implanted gelatin film show good mechanical property [1].

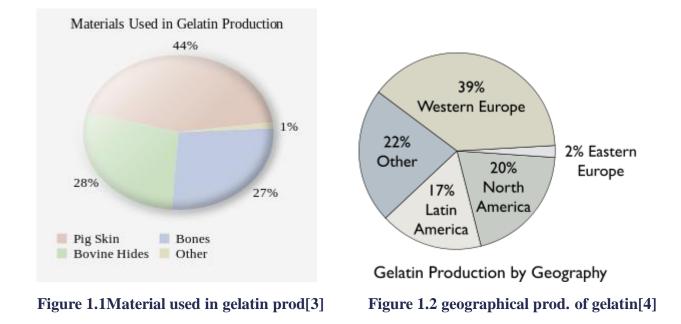
1.2. GELATIN:

1.2.1. Composition and properties

Gelatin is a mixture of peptides and proteins produced by partial hydrolysis of collagen extracted from the skin, bones, and connective tissues of animals such as domesticated cattle, chicken, pigs, horses ,and fish. During hydrolysis, the natural molecular bonds between individual collagen strands are broken down into a form that rearranges more easily. Its chemical composition is, in many respects, closely similar to that of its parent collagen. Photographic and pharmaceutical grades of gelatin are generally sourced from beef bones and pig skin. Gelatin readily dissolves in hot water, and sets to a gel on cooling. When added directly to cold water, it does not dissolve well. Gelatin is also soluble in most polar solvents. Gelatin solutions show visco elastic flow and streaming birefringence. Solubility is determined by the method of manufacture. Typically, gelatin can be dispersed in a relatively concentrated acid. Such dispersions are stable for 10–15 days with little or no chemical changes and are suitable for coating purposes or for extrusion into a precipitating bath.

The mechanical properties of gelatin gels are very sensitive to temperature variations, the previous thermal history of the gels, and time. These gels exist over only a small temperature range, the upper limit being the melting point of the gel, which depends on gelatin grade and concentration, but is typically less than 35 °C (95 °F) and the lower limit the freezing point at which ice crystallizes. The upper melting point is below human body temperature, a factor which is important for mouth feel of foods produced with gelatin. The viscosity of the gelatin/water mixture is greatest when the gelatin concentration is high and the mixture is kept cool at about 4 °C (39 °F). The gel strength is quantified using the Bloom test [2].

1.2.2. Production



The worldwide production amount of gelatin is about 375,000–400,000 tones per year $(830 \times 10^6 880 \times 10^{6}$ lb/a).On а commercial scale, gelatin is made from by-products of the meat and leather industries. Recently, fish by-products have also been considered because they eliminate some of the religious obstacles surrounding gelatin consumption. Gelatin is derived from pork skins, pork, horses, and cattle bones, or split cattle hides. The raw materials are prepared by different curing, acid, and alkali processes which are employed to extract the dried collagen hydrolysate. These processes may take several weeks, and differences in such processes have great effects on the properties of the final gelatin products. Gelatin can also be prepared in the home. Boiling certain cartilaginous cuts of meat or bones results in gelatin being dissolved into the water. Depending on the concentration, the resulting stock (when cooled) will naturally form a jelly or gel. This process is used for aspic.

While many processes exist whereby collagen can be converted to gelatin, they all have several factors in common. The intermolecular and intra molecular bonds which stabilize insoluble collagen must be broken, and the hydrogen bonds which stabilize the collagen helix must also be broken. The manufacturing processes of gelatin consists of three main stages:

1. Pretreatments to make the raw materials ready for the main extraction step and to remove impurities which may have negative effects on physiochemical properties of the final gelatin product

- 2. The main extraction step, which is usually done with hot water or dilute acid solutions as a multistage extraction to hydrolyze collagen into gelatin
- 3. The refining and recovering treatments including filtration, clarification, evaporation, sterilization, drying, rutting, grinding, and sifting to remove the water from the gelatin solution, to blend the gelatin extracted, and to obtain dried, blended and ground final product.

1.2.3. Pretreatments

If the raw material used in the production of the gelatin is derived from bones, dilute acid solutions are used to remove calcium and other salts. Hot water or several solvents may be used to reduce the fat content, which should not exceed 1% before the main extraction step. If the raw material consists of hides and skin; size reduction, washing, removal of hair from hides, and degreasing are necessary to prepare the hides and skins for the main extraction step.

Collagen hydrolysis is performed by one of three different methods: acid, alkali, and enzymatic hydrolysis. Acid treatment is especially suitable for less fully crosslinked materials such as pig skin collagen and normally requires 10 to 48 hours. Alkali treatment is suitable for more complex collagen such as that found in bovine hides and requires more time, normally several weeks. The purpose of the alkali treatment is to destroy certain chemical crosslinks still present in collagen. Within the gelatin industry, the gelatin obtained from acid-treated raw material has been called type-A gelatin and the gelatin obtained from alkali-treated raw material is referred to as type-B gelatin.

Enzymatic hydrolysis of collagen for gelatin extraction is relatively new. However, the treatment time is shorter than that required for alkali treatment, and results in almost complete conversion to the pure product. The physical properties of the final gelatin product are better.

1.2.4. Extraction

After preparation of the raw material, i.e., reducing cross-links between collagen components and removing some of the impurities such as fat and salts, partially purified collagen is converted into gelatin by extraction with either water or acid solutions at appropriate temperatures. All industrial processes are based on neutral or acid pH values because though alkali treatments speed up conversion, they also promote degradation processes. Acidic extraction conditions are extensively used in the industry, but the degree of acid varies with different processes. This extraction step is a multistage process, and the extraction temperature is usually increased in later extraction steps, which ensures minimum thermal degradation of the extracted gelatin.

1.2.5. Recovery

This process includes several steps such as filtration, evaporation, drying, grinding, and sifting. These operations are concentration-dependent and also dependent on the particular gelatin used. Gelatin degradation should be avoided and minimized, so the lowest temperature possible is used for the recovery process. Most recoveries are rapid, with all of the processes being done in several stages to avoid extensive deterioration of the peptide structure. A deteriorated peptide structure would result in a low gel strength, which is not generally desired [5].

1.2.6. Technical uses



- Certain professional and theatrical lighting equipment use color gels to change the beam color. These were historically made with gelatin, hence the term color gel.
- Gelatin typically constitutes the shells of pharmaceutical capsules to make them easier to swallow. Hypromellose is a vegetarian-acceptable alternative to gelatin, but is more expensive to produce.
- Some animal glues such as hide glue are essentially unrefined gelatin.
- It is used to hold silver halide crystals in an emulsion in virtually all photographic films and photographic papers. Despite some efforts, no suitable substitutes with the stability and low cost of gelatin have been found.
- Used as a carrier, coating, or separating agent for other substances, for example, it makes β-carotene water-soluble, thus imparting a yellow color to any soft drinks containing beta-carotene.

- Gelatin is closely related to bone glue and is used as a binder in match heads and sandpaper.
- Cosmetics may contain a nongelling variant of gelatin under the name hydrolyzed collagen.
- Gelatin was first used as an external surface sizing for paper in 1337 and continued as a dominant sizing agent of all European papers through the mid-19th century. In modern times, it is mostly found in watercolor paper, and occasionally in glossy printing papers, artistic papers, and playing cards, and it maintains the wrinkles in crêpe paper [6].

1.2.7. Other uses

- Blocks of ballistic gelatin simulate muscle tissue as a standardized medium for testing ammunition.
- Gelatin is used by synchronized swimmers to hold their hair in place during their routines, as it will not dissolve in the cold water of the pool. It is frequently referred to as "knoxing", a reference to Knox brand gelatin.
- When added to boiling water and cooled, unflavored gelatin can make a home-made hair styling gel that is cheaper than many commercial hair styling products, but by comparison has a shorter shelf life (about a week) when stored in this form (usually in a refrigerator). After being applied to scalp hair, it can be removed with rinsing and some shampoo.
- It is commonly used as a biological substrate to culture adherent cells.
- It may be used by those who are sensitive to tannins (which can irritate the stomach) in teas, soups or brews.
- It may be used as a medium with which to consume LSD. LSD in gelatin form is known as "windowpane" or "gel tabs".
- Gelatin is used to make the shells of paintballs, similar to the way pharmaceutical capsules are produced.
- It is used as an ingredient in implantable medical devices, such as in some bone void fillers.
- Leaf or sheet gelatin is used directly in food-based model-making, for example to make translucent, edible, diamond-paned windows in gingerbread houses.
- Gelatin can be used as a binding agent in India ink.
- It may be used as a technique within the process of fine-art printmaking. The prints are made by creating a block of gelatin and applying printing inks. The gelatin is made using twice the normal amount of gelatin granules to the usual amount of water. Once set –

printmaking ink (usually water-based) is applied to its surface. Other water-based media may also be applied. Items such as dried grass, leaves, and paper stencils are placed onto the inked surface [7].

1.3. BIOGLASS:

Current interest in biomaterials research focuses on advanced scaffold materials for tissue engineering applications to repair and regenerate living tissues and organs damaged as a result of strain, injury, disease or aging. In tissue engineering, the scaffold with well-defined architecture acts as a temporary support for the cells and guides their proliferation and demarcation into the desired tissues or organs. The scaffold design must meet the demands of tissue engineering such as biocompatibility, sterilizability, mechanical stability and controllable interconnected porosity for healthy growth of tissue. Apart from these requirements, new third generation scaffolds of required molecular modification in order to activate genes as well as cells are under research. These molecularly tailored third generation biomaterials can be the most successful scaffolds to synthesis new tissue of required shape and dimensions. Numerous materials such as natural or synthetic polymers, bioactive glass/glass ceramics, composites of polymers and ceramics as well as metallic materials are being developed to act as synthetic scaffolds for bone tissue engineering applications. Among these materials, bioactive silicate glasses, glass ceramics and their related composites combining bioactive inorganic materials with biodegradable polymers are extensively investigated as bone grafts. These bone graft materials are also being developed for hard tissue engineering applications due to their desired physical properties (mechanical stability and porosity), biocompatibility, osteoconductive as well as osteoinductive nature. Interestingly, no other glass or glass-ceramic compositions have been found to date with excellent biocompatibility and bone bonding ability compared to Na-containing silicate bioactive glass of composition 45S5, discovered by Hench in 1969. The unique atomic structure, structural linkages and local environment of the 45S5 bioglass constituent elements play an important role to stimulate biological activities like bone cell growth and related gene up regulations. It is also noteworthy that 45S5 bio glass is in clinical practice for dental bone regeneration and orthopedic bone grafting. Such potentially important bioglass of various textural properties are so far mostly fabricated by either melt-quenching or sol-gel techniques. The traditional melt-quenching method involves a high temperature process to synthesis materials of limited compositional range. Whereas, the sol-gel method requires toxic organic solvents and follows complicated organometallic synthetic strategies .Various hybrid bio glass scaffold systems with pore sizes ranging from macro to meso are fabricated using biodegradable polymers and inorganic materials. Strong correlations between the porosity and chemical composition of the hybrid bioglass and their biodegradability and bioactivity are discussed in literature. Interestingly, these porous hybrid bioactive glasses of superior surface area and pore volume with favorable chemical compositions are suggested as excellent candidates for scaffolding bone tissue regeneration as well as matrices for local drug delivery systems. Recently, known hierarchical bioglass scaffolds having close similarity to native bones have been shown to offer great promise for effectively restoring the functions of degenerated tissues by significantly inquencing cell behavior including cell adhesion, spreading, proliferation and differentiation in a controlled manner. Additionally, the potential of hierarchical bioactive nano-glass particles is being understood for advanced applications like cell tracking and intracellular delivery of molecules such as therapeutic agents, protein sand [8]. Naturally occurring materials, such as shell, bones and teeth with outstanding architecture and mechanical properties as well as novel nano-sized natural materials like nano-structured silica in diatoms or the bio mineralization of oriented

intracellular chains made of single domain crystals of magnetite or of meta stable ferromagnetic iron sulphide in magnetotactic bacteria, are guided by specific gene products, mostly proteins of natural origin in atmospheric conditions. Here, these bio macromolecules can serve as templates, scaffolds and stabilizers for the material synthesis either by controlling its crystallographic orientation and growth, by imitating the lattice of a two-dimensional face or by stereochemistry of the functional groups at the interface. In addition, heterogeneous nucleation of the hierarchical inorganic materials by simple electrostatic attraction between the charge density of polymeric and inorganic phases is also demonstrated. These biopolymeric templates prefer ambient conditions to retain their original structure in order to nucleate the expected hierarchy in the resulting material. Based on these interesting observations, the bio-inspired route was developed and is well established in the synthesis of nanostructured ceramic oxides below 100°c by using polymers of natural origin. Although few researchers attempted to synthesis bioglass and bioglass scaffolds in ambient conditions, they could not avoid a sintering step to achieve the required mechanical stability of the material. Such a sintering step above ambient temperature may collapse the one architecture of the material for required advanced applications. In this regard, the observed demand for alternative bioglass processing methods in ambient conditions provoked us to introduce the bio-inspired route to synthesis potentially important 45S5 bioglass. Among all natural polymeric templates, c-tab possesses a linear/coiled structure, mechanical rigidity and physicochemical stability and is extensively used for controlled arrangement of molecules and nano-particles into well-defined structured materials by numerous researchers. Extending these interesting readings to the bioglass synthesis, here we report a new procedure for obtaining hierarchical 45S5 glassceramics of various pore sizes in ambient conditions using c-tab as template [9].

CHAPTER 2

MATERIAL & METHOD

2.1 Materials:

The chemicals used in this project are as follows:

S. No.	Chemical Name	Catalogue No./	Structure
		Company	
1.	Sodium acetate	Sigma-Aldrich/	0
		\$8750	H ₃ C [∕] O [©] Na [⊕]
2.	Tetra ethyl Ortho	Sigma-Aldrich/	H ₃ CCH ₃
	Silicate (TEOS)	131903	H ₃ C CH ₃
3.	Calcium acetate	Sigma-Aldrich/	Ca ^{Ca} CH ₃
		402850	H ² C O Ö
4.	Tri ethyl phosphate	Sigma-Aldrich/	O H-O
	(TEP)	538728	
5.	Cetyltrimethyl	SDFCL/37665	CH ₃
	ammonium	KO1	H_3C $N - CH_3$
	bromide(CTAB)		ĊH ₃ Br
6.	Gelatin Powder	Lobachemie/	
		0392000500	
			+ NH2
7.	Glutaraldehyde	CDH	0~~~~0
8.	Distil Water	Millipore	H ₂ O

TABLE 2.1 List of chemicals used

2.2. Synthesis of bioglass particles:

2.2.1. Preparation of TRIZMA buffer solutions

An aqueous solution of 30 mM Tris(hydroxymethy)aminomethane (TRIZMA) buffer at pH8 was prepared by dissolving 1.82 g/l TRIZMA HCl and 2.22 g/l TRIZMA base in Milli-Q water.

2.2.2. Preparation of CTAB solution:

The required amount of material was dissolved in 10 ml of 10 mM TRIZMA buffer solution at pH 8 in a 50 ml vial in such a way to get 10mM of CTAB and kept in an orbital shaker (Thermo Scientific) at 37°C and 180 to 220 rpm overnight.

2.2.3. Bio-inspired synthesis of 45S5 glass-ceramic using CTAB as template

The precursors, namely 9.29 g of tetraethyl orthosilicate, 1.0 g of triethyl phosphate, 6.36 g of sodium acetate, and 4.21 g of calcium acetate, were added slowly and steadily one after another into the 100 ml TRIZMA buffer containing CTAB. For mixing each precursor, half an hour interval was allowed with constant stirring at 37°C in a silicone oil bath for 24 h. After 24 h a white precipitate was formed, centrifuged, washed with Milli-Q water and dried at 40°C in an air oven for 48 h and preserved in desiccators. A similar procedure was also followed to obtain a bioglass sample in the absence of CTAB. Interestingly, the precipitate was not observed after 24h conditioning time at 37°C.

2.3. Synthesis of bioglass doped films

Sample 1: Gelatin type A was used. Gelatin films were prepared from a 20% Conc.(10 gm in 50 ml of water) aqueous gelatin solution. Films were obtained on the bottom of Petri dishes (diameter"6 cm) after water evaporation at room temperature from 50 ml of gelatin solution. After air drying, the film was subjected for different test by cutting in the required proper

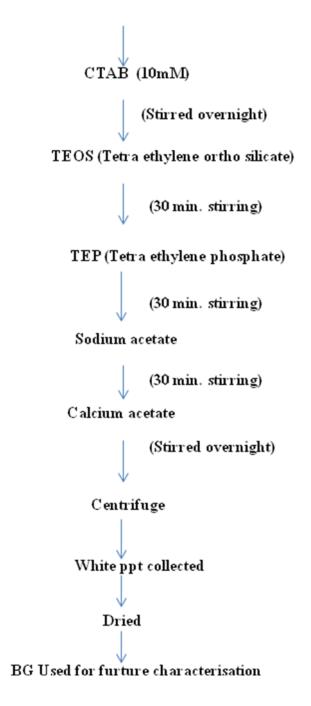
dimensions. The so obtained pure gelatin film was referred as Gel. In another set of experiment, 100 mg pre-synthesized bioglass was added to the 50 ml gelatin solution. The resulted mixture was vigorously stirred slow heating by magnetic stirrer to get uniform mixing of the solution and then poured into the petri-dish. The petri-dish was then left undisturbed until the sample got dried. The resulted Bioglass doped gelatin film was referred as Gel BG.

Sample 2: Gelatin films crosslinked with glutaraldehyde were also prepared. For Glutaraldehyde crosslinked gelatin film, to 20% conc. (10 gm in 50 ml of water) aqueous gelatin solution, 40 μ l of 2.5 % glutaraldehyde was added. The so obtained solution was then poured into the petri-dish. Films were obtained on the bottom of Petri dishes (diameter"6 cm) after water evaporation at room temperature from50 ml of solution. After air drying, the film was subjected for different test by cutting in the required proper dimensions. The so obtained glutaraldehyde crosslinked gelatin film was refereed as Gel glutaraldehyde In another set of experiment,100 mg presynthesized bioglass was added to the 50ml of glutaraldehyde gelatin solution. The resulted mixture was vigorously stirred with slow heating by magnetic stirrer to get uniform mixing of the solution and then poured into the Petri-dish. The Petri-dish was then left undisturbed until the sample got dried. The resulted bioglass doped gelatin film was referred as Gel Glu. BG .

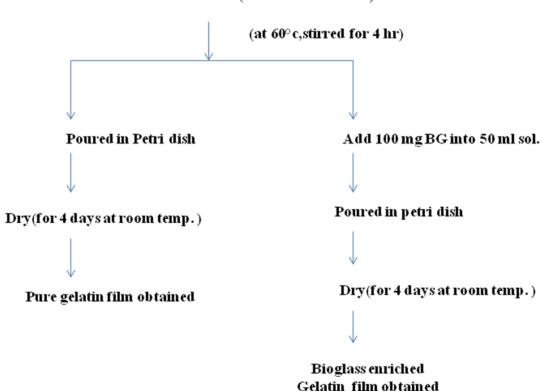
2.4PRODUCTION FLOWCHARTS:

2.4.1. BG formation

TRIZMA BUFFER(10mM)

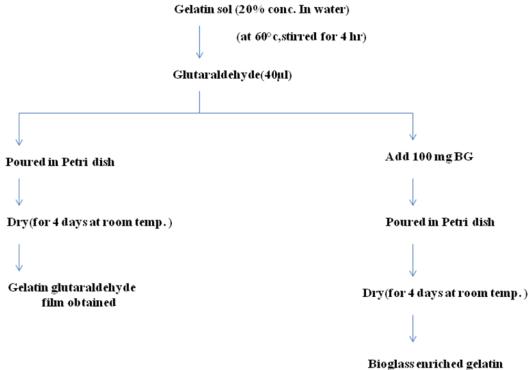


2.4.2. Gelatin & bioglass enriched gelatin film formation



Gelatin sol. (20% conc. In water)

2.4.3. Gelatin glu. & bioglass enriched gelatin glu. film formation



Glutaraldehyde film obtained

CHAPTER 3

CHARACTERIZATION TECHNIQUES/ INSTRUMENTATION

3. Instrumentation:

3.1. FTIR studies

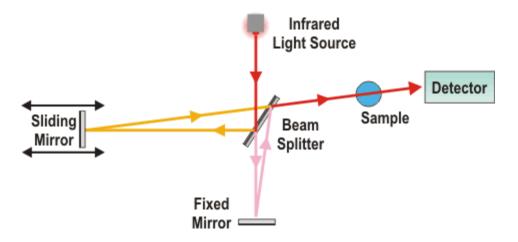
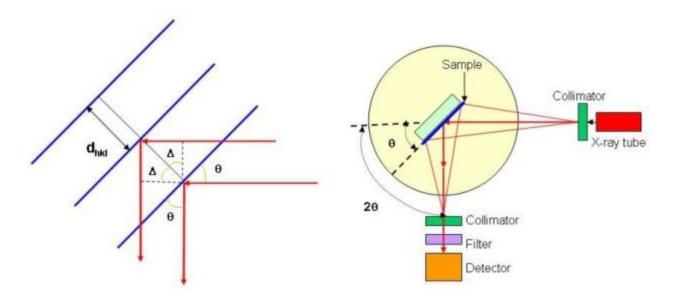


Figure 3.1 working principle diagram of FTIR [10]

FTIR reckons on the fact that most of the molecules absorb light in the infra-red range of the electromagnetic spectrum. This absorption corresponds particularly to the bonds of the molecule .The frequency range is calculated as wave numbers over the range of 4000 – 6000 cm⁻¹ FTIR reckons on the fact that most of the molecules absorb light in the infra-red range of the electromagnetic spectrum. This absorption corresponds particularly to the bonds of the molecule. The frequency range is calculated as wave numbers over the range of 4000 – 600 cm⁻¹. The beckground emission spectrum of the Infra-Red source is recorded, followed by the emission spectrum of the Infra-Red source with the sample in place. The ratio of the sample emission spectrum. The resultant absorption spectrum due to the bond natural vibrational frequencies shows the presence of different chemical bonds and functional groups in the sample. FTIR is specifically useful for identification of organic molecular groups and compounds because of functional groups, side chains and cross-links involve will have characteristic vibrational

frequencies in the infra-red range. The background emission spectrum the Infrared source is recorded, followed by the emission spectrum of the Infra-Red source with the sample in place. The ratio of the sample emission spectrum to the background emission spectrum is directly related to the sample& absorption spectrum. The resultant absorption spectrum due to the bond natural vibrational frequencies shows the presence of different chemical bonds and functional groups in the sample. FTIR is specifically useful for identification of organic molecular groups and compounds because of functional groups, side chains and cross-links involve will have characteristic vibrational frequencies in the infra-red range [11].



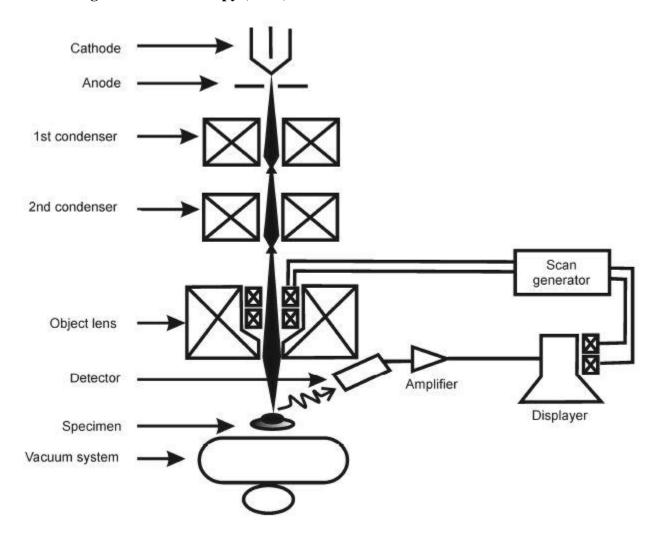
3.2 X-ray diffraction (XRD):

Figure 3.2. Working principle diagram of X-ray diffraction (XRD)

X-ray powder diffraction (XRD) is a high-speed analytical technique mostly used for phase determination of a crystalline material and can also be used to get information about unit cell dimensions. The analyzed material is finely ground, homogenized, and average bulk composition is calculated. X-ray diffraction is based on constructive interference of monochromatic X-rays

with a crystalline sample. These X-rays are produced by a cathode ray tube, which is then filtered to generate monochromatic radiation, collimated to condense, and directed to the sample. The interaction of the incident rays with the sample cause constructive interference and a diffracted ray is produced, when conditions satisfy Braggs Law [12].

$$n\lambda = 2d \sin \theta$$
 Eq.1



3.3. Scanning electron microscopy (SEM):

Figure 3.3. Working principle diagram of SEM

A scanning microscope (SEM) is one type of electron microscope which produces images of the samples by scanning it using a focused beam of electrons. The electrons are interacts with the

atoms in the sample and produces various signals that contain information regarding the sample& surface topography and composition. The electron beam is usually scanned in an exceedingly formation scan pattern and therefore the beam's position is combined with the detected signal to provide a picture. SEM can be able to do resolution higher than 1 nanometer. Specimens will be observed in high vacuum, in low vacuum, in wet conditions and at a wide range of cryogenic or elevated temperatures [13].

3.4. TENSILE TEST:

UTM

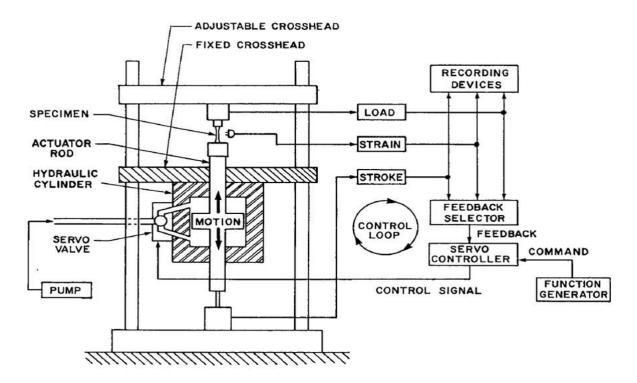


Figure 3.4: Working principle diagram of UTM

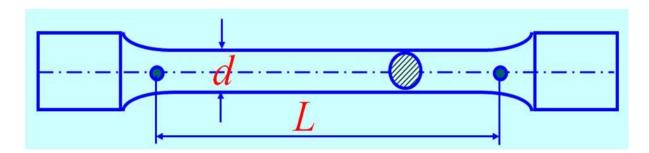


Figure 3.5: A Tensile Testing Specimen

3.4.1. Introduction

A tension test is commonly used for determining the quasi-static properties of a material type. Results of tension tests are tabulated in handbooks and, through the use of failure theories, these data can be used to predict failure of parts subjected to more generalized stress states. Theoretically, this is a good test because of the apparent simplicity with which it can be performed and because the uniaxial loading condition results in a uniform stress distribution across the cross section of the test specimen [14].

3.4.2. Principle

When a prismatic bar is subjected to two equal and opposite forces along the bar axis, a change of interaction force among various parts of the bar is introduced. This change of interaction force is referred to as internal force. In terms of deformation, tensile loads cause the bar to extend longitudinally (axially) and contract transversely. Prior to the eventual breakage, bar deformation can be classified into four distinct stages, i.e. elastic, yielding, hardening and necking. The latter three are collectively termed as inelastic deformation.

3.4.3Objective

- Identify the elastic, plastic and strength properties of a testing specimen
- Understand the stress-strain curve of typical ductile materials [15].



RESULT & DISCUSSION

4.1. XRD:

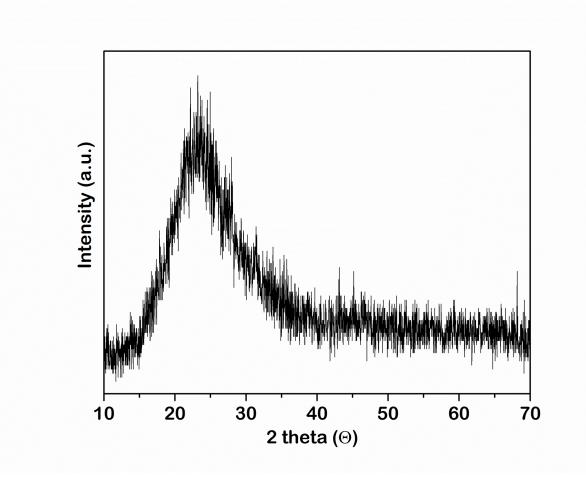


Figure 4.1: XRD result of bioglass sample

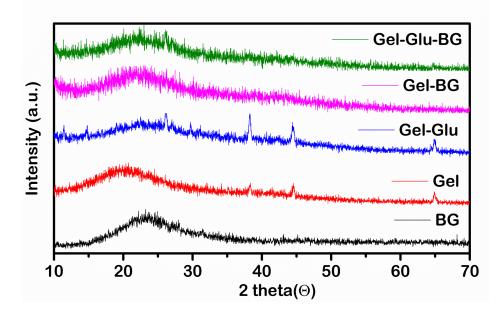


Figure 4.2: XRD result of BG, Gel, glutaraldehyde crosslinked gelatin (Gel-Glu) and bioglass enriched gel & gel-glutaraldehyde

For pure bioglass, a broad hump appeared in the XRD graph (Figure 1) ranging from 15° to 30° depicting the amorphous nature of the material. However, a sharp peak at 39° , 44° and 65° 20 values while a broad hump was observed from 15° to 25° in case of gelatin film (Figure 2) depicting the semi-crystalline nature of the film through XRD. A similar spectra was observed in case of glutaraldehyde crosslinked gelatin film (Gel Glu) with only the increase in intensity. However, for bioglass doped films (GB, GGB), sharp peaks were no more visible but only the broad peak ranging from 15° to 30° 20 degree values observed in the diffraction pattern indicating that the bioglass must have doped into the films.

4.2 FTIR:

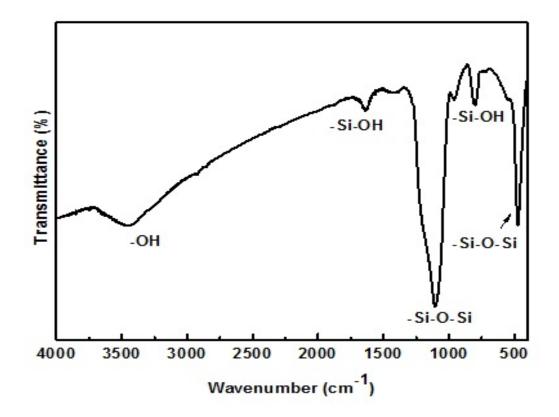


Figure 4.3: FTIR Result for pure bioglass sample

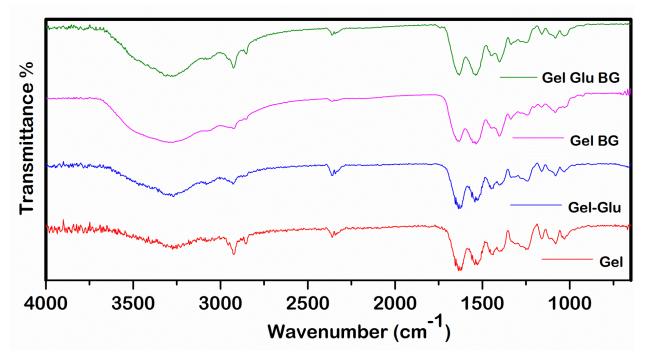


Figure4.4: FTIR result of Gel, glutaraldehyde crosslinked gelatin (Gel-Glu) and bioglass enriched gel & gel-glutaraldehyde.

FTIR spectra for the as-syntheisized CTAB templated bioglass was recorded raging from 4000 to 400 cm⁻¹. In the recoded FTIR spectra (Figure 4.4), characteristic broad intense peak at 1200-1000 cm⁻¹ and a small sharp peak at 467 cm⁻¹ attributed to Si–O–Si asymmetric stretching and bending vibrations respectively are observed. A small shoulder at 956 cm⁻¹ assigned to non-bridging oxygen together with the surface active silanols (Si–OH) groups, which enhances the rate of apatite formation are visualized. The peak at 803 cm⁻¹ could be attributed to characteristic silicate network ring structures. Peak at 644 cm⁻¹ corresponding to P-O bond bending vibration and indicating that phosphate enter as network former is observed.

FTIR graph of films were recorded in ATR mode from 4000-650 cm⁻¹ as shown in figure 4. C-H skeletal vibrations are observed at 2918 cm⁻¹ in gelatin film. Additionally, peaks at 1245 cm⁻¹, 1637 cm⁻¹ and 1530 cm⁻¹ could be attributed to C-N, amide I (C=O group) and amide II, respectively in case of gelatin and glutaraldehyde crosslinked gelatin films which are observed to get weaker in case of bioglass doped films. It is pertinent to observe the emergence of new peak corresponding to Si-OH and Si-O-Si asymmetric stretch at 1401 cm⁻¹ and 1085 cm⁻¹ respectively due to embedding of bioglass into film Gel and Gel-Glu alongwith the amide 1 (C=O group) peak retention at 1637 cm⁻¹ due to gelatin.

4.5 Gelatin and glutaraldehyde Cross-Linking Chemistry:

Due to its unique amino acid sequences and numerous functional groups, gelatin is well-suited for producing chemical hydro gels in the form of sheets, films, or membranes by reacting with small molecules containing reactive functional groups, such as an aldehyde group. As mentioned before, it is generally accepted that the mechanism of gelatin cross-linking mediated by glutaraldehyde can be explained through the reaction of the aldehyde functional groups with free nonprotonated ε -amino groups (-NH2) of lysine or hydroxyl sine through a nucleophilic addition-type reaction. Although such reaction normally requires sub acid conditions, neutral to slightly alkaline pH values are more favorable for gelatin crosslinking[16].

More specifically, the first step of the reaction involves the nucleophilic addition of the ε -NH2 groups to the carbonyl groups (CdO) of the aldehyde to form a tetrahedral unstable intermediate called carbinolamine. In a second step, protonation of the -OH group followed by loss of a water molecule yields the conjugated Schiff bases . Such a mechanism results in the formation of new covalent bonds between gelatin molecules at either intramolecular (short-range) or intermolecular scale (long-range). The longdistance bridges form through the polymerization of glutaraldehyde in aqueous solution or aldol condensation reaction[17].

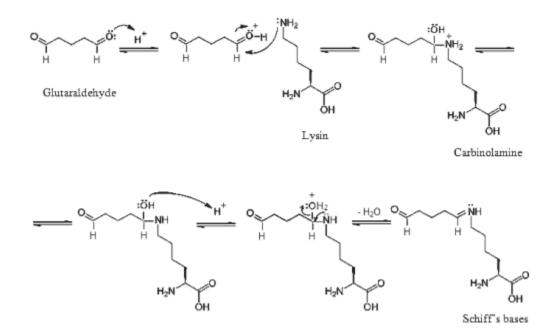
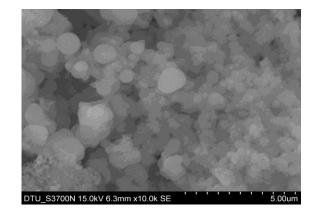
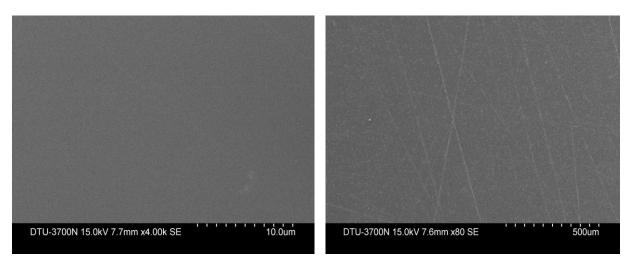


Figure 4.5. Reaction mechanism between amino groups of lysine and carbonyl groups of glutaraldehyde for the formation of Schiff [18].

4.3. SEM:







(b)

(**c**)

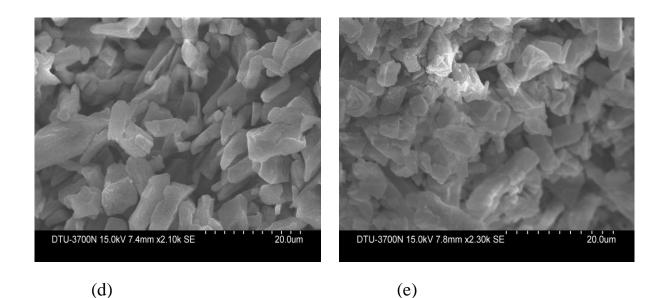


Figure 4.6:Sem images of samples: a).pure bioglass(BG) sample b). pure gelatin sample c).gelatin glutaraldehyde d).gelatin BG e). gelatin glutaraldehyde BG

SEM image of the pure bioglass (Figure 4.6.a) depicts the spherical morphology of the bioglass particles. Gelatin and Glutaraldehyde crosslinked gelatin films are observed to possess smooth surface morphology as shown in (figure 4.6.b) and (4.6.c) respectively. While embedded bioglass particles into gelatin and Glutaraldehyde crosslinked gelatin films after doping could be observed in figure (4.6.d) and (4.6.e), respectively. The doped bioglass are observed to be of bigger size could be due to the aggregation of the particles during processing.

4.4. Tensile result:

4.4.1. UTM

DELHI COLLEGE OF ENGG.

Graph 1

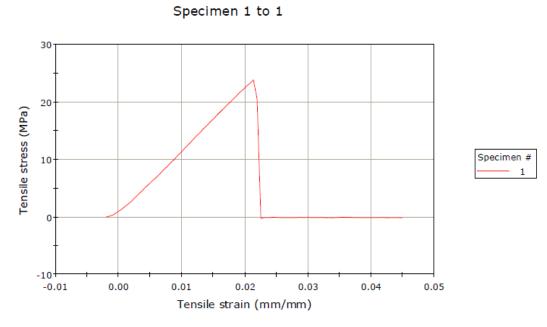


Figure 4.7: UTM result of pure gelatin film

DELHI COLLEGE OF ENGG.

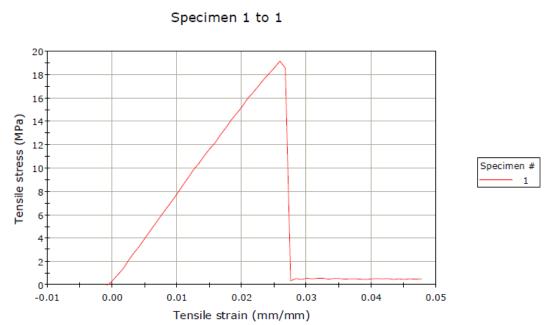


Figure 4.8: UTM result of bioglass enriched gelatin film

Stress/Strain curve were plotted for both the samples on UTM by properly cutting the samples of required length and Thickness of the samples measured by using vernier caliper so that to maintain the equal area for both the samples. The stress/strain curves for both the samples are shown in figure (4.7) & (4.8). The tensile stress vs strain curve in fig 1 (pure gelatin film) shows a sharp peak at break as compared to fig 2 (bioglass enriched gelatin film). Because bioglass particles incorporated in gelatin film provides a resistance during break and so crack have to propogate a longer path which results in higher elongation at break and does not show a sharp peak. Hence incorporation of bioglass particles results in better creep properties & strain modulus which widen the application area of gelain film.

CHAPTER 5

CONCLUSIONS & FUTURES SCOPE OF WORK

5.1. CONCLUSION:

As revealed from the results of XRD, FTIR and SEM that the synthesize bioglass was successfully doped in gelatin and glutaraldehyde crosslinked gelatin films. Also, using solution casting method successfully glutaraldehyde crosslinked gelatin films were obtained. In addition to this the result from the UTM study reveals that bioglass incorporated gelatin film have better creep property and strain modulus as compared to gelatin film without bioglass particles, so provide a better scope for various biomedicals applications like in vascular grafts or scaffolds.

5.2 FUTURE SCOPE

The present piece of work needs to be further studied in depth for practical use, in consideration of the following points :

- Effect of different conc. Of Glutaraldehyde as a cross-linker for gelatin film with the proposed procedure need to be evaluated.
- Biocompatibility of the different bioglass doped and undoped gelatin and Glutaraldehyde crosslink gelatin film needs to be explored for making its application area wider.
- Intensive study of its mechanical and visco-elastic properties need to be carried out to look forth its application as an implant material.
- Its processing techniques needs to be future optimize for its future scaffolds application.

REFERENCES:

[1] Veis A. The macromolecular chemistry of gelatin. New York,London: Academic Press, 1964.

[2] Rose PJ, Mark HF, Bikales NM, Overberger CG, Menges G, Kroschwitz JI. Encyclopedia of polymer science and engineering,2nd ed., vol. 7. New York: Wiley Interscience, 1987.

[3] "Global Gelatin market". Retrieved 11 MAY 2016.

[4] <u>"Natural Health Products Ingredients Database: Hydrolyzed Collagen"</u>. Government of Canada, Health Canada, Health Products and Food Branch, Natural Health Products Directorate. Retrieved 9 May 2016.

[5] Hastings GW, Ducheyne P. Macromolecular biomaterials. Boca Raton: CRC Press, 1984.

[6] Pezron I, Djabourov M, Leblond J. Polymer 1991.

[7] Ross-Murphy SB. Polymer 1992;33:2622}.

[8] Digenis GA, Gold TB, Shah VP. Cross-linking of gelatin capsules and its relevance to their in vitro}in vivo performance. J Pharm Sci 1994;83:915}.

[9] Esposito E, Cortesi R, Nastruzzi C. Gelatin microspheres: in#uence of preparation parameters and thermal treatment on chemico-physical and biopharmaceutical properties. Biomaterials 1995;20:2009}.

[10] Otani Y, Tabata Y, Ikada Y. Hemostatic capability of rapidly curable glues from gelatin poly(L-glutamic acid) and carbodiimide. Biomaterials 1998;19:2091}.

[11] Davis, Joseph R. (2004), Tensile testing (2nd ed.), ASM international.

[12] Lapitsky, Y.; Zahir, T.; Shoichet, M. S. Modular biodegradable biomaterials from surfactant and polyelectrolyte mixtures. Biomacromolecules 2008.

[13] Metzke, M.; Guan, Z. Structure-property studies on carbohydratederived polymers for use as protein-resistant biomaterials. Biomacromolecules 2008.

[14] Hoare, T. R.; Kohane, D. S. Hydrogels in drug delivery: progress and challenges. Polymer 2008, 49, 1993–2007.

[15] Turgeon, S. L.; Schmitt, C.; Sanchez, C. Protein-polysaccharide complexes and coacervates.Curr. Opin. Colloid Interface Sci. 2007,

[16] Khademhosseini, A.; Langer, R. Microengineered hydrogels for tissue engineering.Biomaterials 2007, 28, 5087–5092.

[17] Lee, K. Y.; Mooney, D. J. Hydrogels for tissue engineering. Chem. Rev. 2001, 101, 1869– 1879.

[18] Hoffman, A. S. Hydrogels for biomedical applications. Biocompatible and biodegradable ultrafine fibrillar scaffold materials for tissue engineering by facile grafting of l-lactide onto chitosan. Adv. Drug Delivery Rev. 2002.